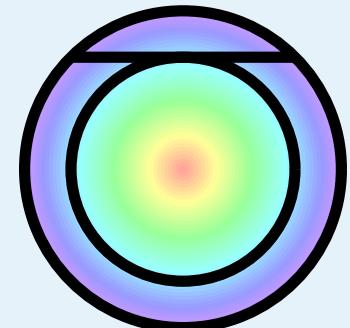
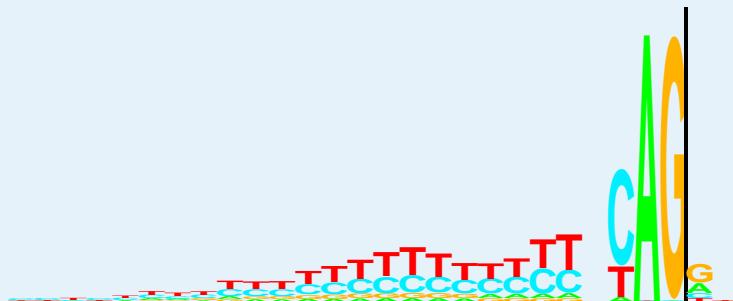




Information Theory in Biology

Thomas D. Schneider, Ph.D.

Molecular Information Theory Group
Center for Cancer Research
Gene Regulation and Chromosome Biology Laboratory
National Cancer Institute
Frederick, MD 21702-1201

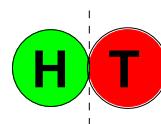


Information Theory: One-Minute Lesson

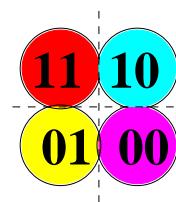
number of symbols	number of bits	example
-------------------	----------------	---------

M	B
---	---

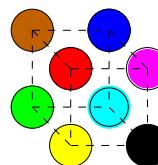
2	1
---	---



4	2
---	---



8	3
---	---



$$M=2^B$$

$$B=\log_2 M$$

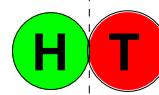


Information Theory: One-Minute Lesson

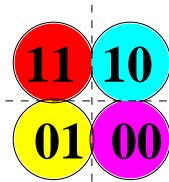
number of symbols	number of bits	example
-------------------	----------------	---------

M	B	
---	---	--

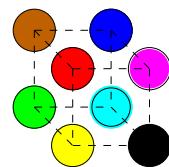
2	1	
---	---	--



4	2	
---	---	--



8	3	
---	---	--



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$$B=\log_2 M$$

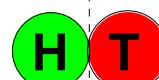


Information Theory: One-Minute Lesson

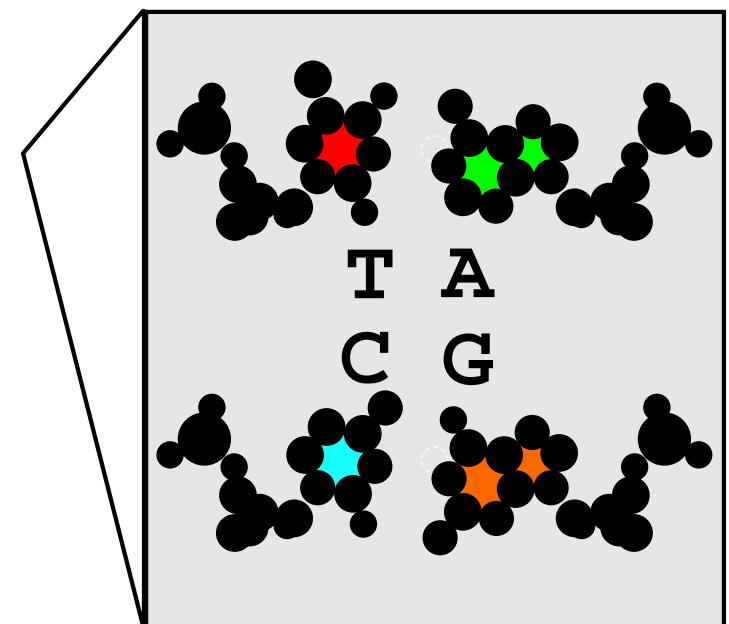
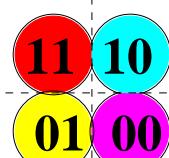
number of symbols	number of bits	example
-------------------	----------------	---------

M	B
---	---

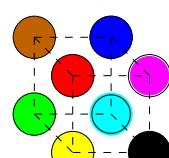
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---	---



4	2
---	---



8	3
---	---



$$M=2^B \quad B=\log_2 M$$

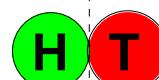


Information Theory: One-Minute Lesson

number of symbols	number of bits	example
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M	B	
---	---	--

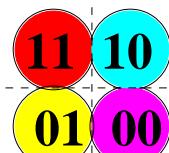
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---	---	--



H T

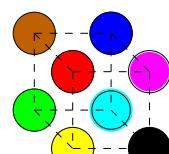


4	2	
---	---	--



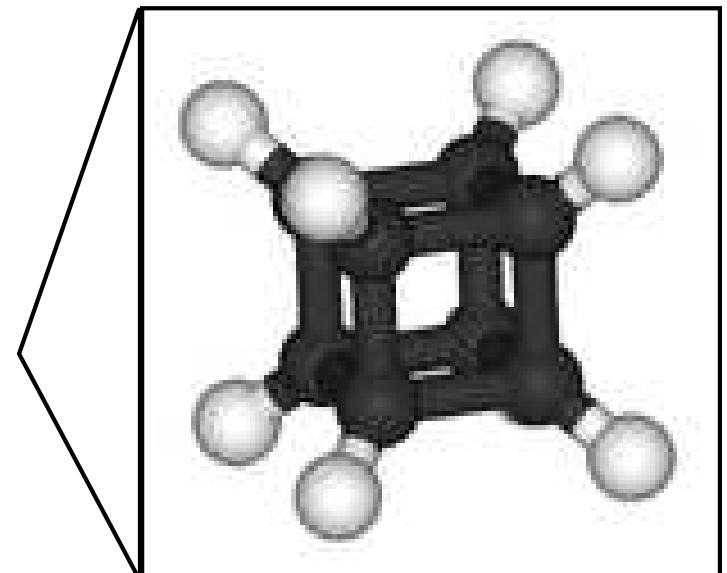
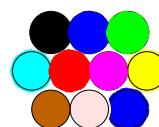
11 10
01 00

8	3	
---	---	--



000 001 010 011
100 101 110 111

$$M=2^B \quad B=\log_2 M$$

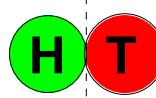


Information Theory: One-Minute Lesson

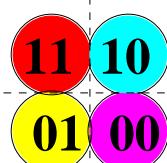
number of symbols	number of bits	example
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M	B	
---	---	--

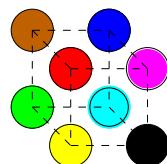
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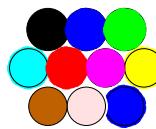
4	2	
---	---	--



8	3	
---	---	--



$$M=2^B \quad B=\log_2 M$$

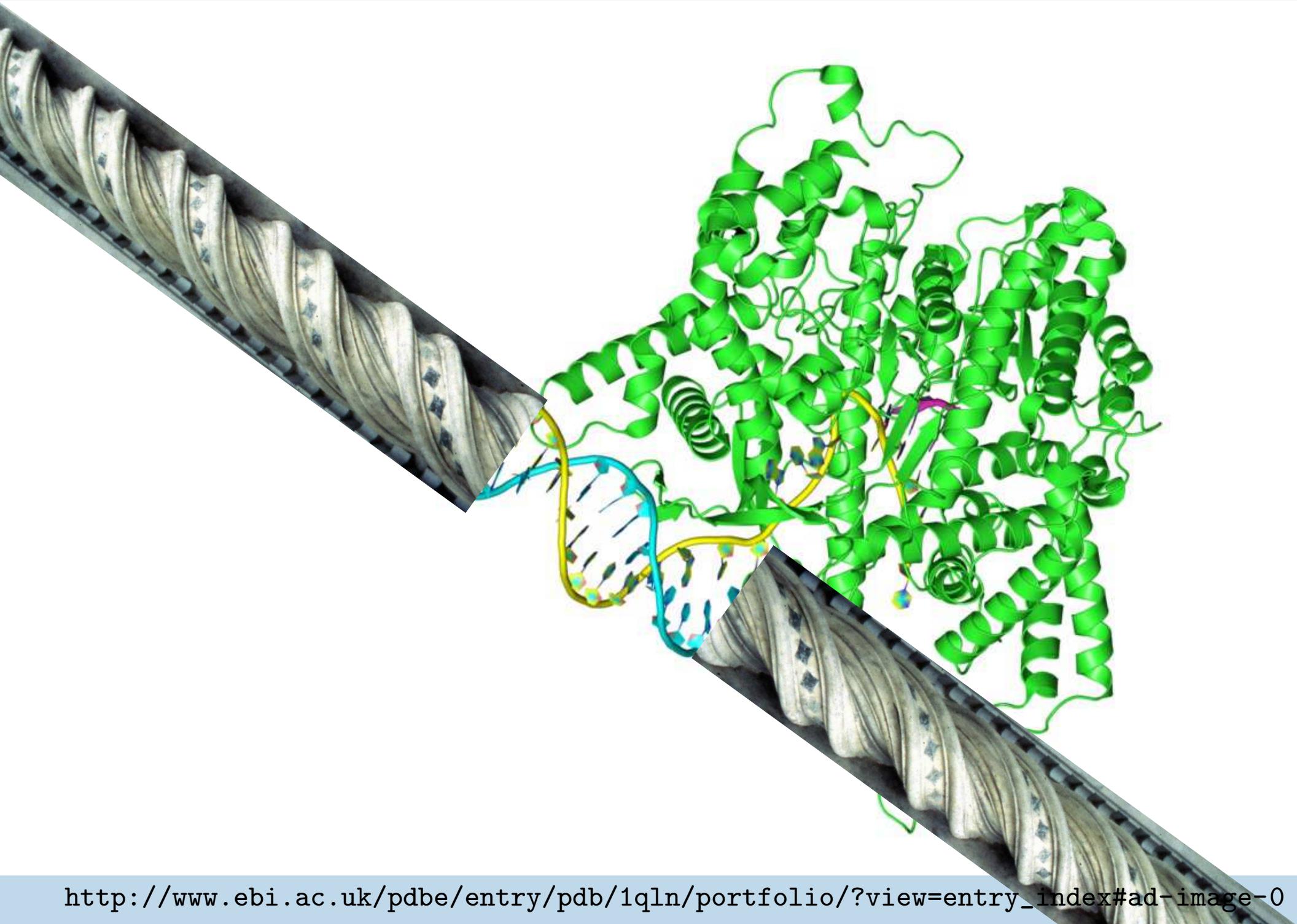


El Duomo, Florence, Italy



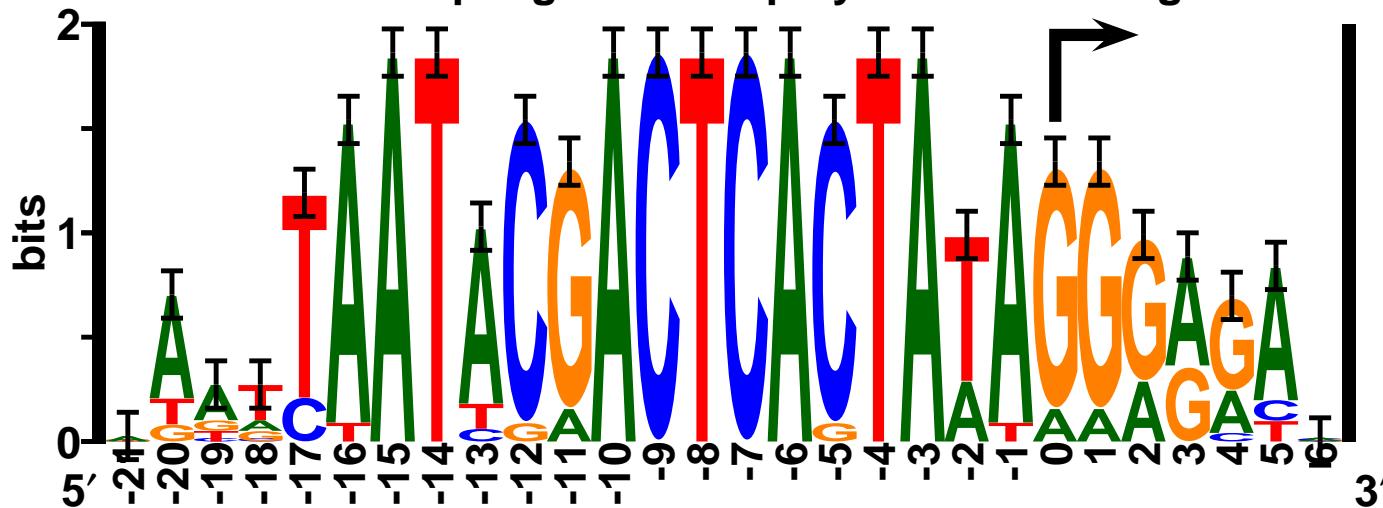
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May not be used for commercial purposes.

T7 RNA polymerase + DNA



Sequence Logo

17 Bacteriophage T7 RNA polymerase binding sites



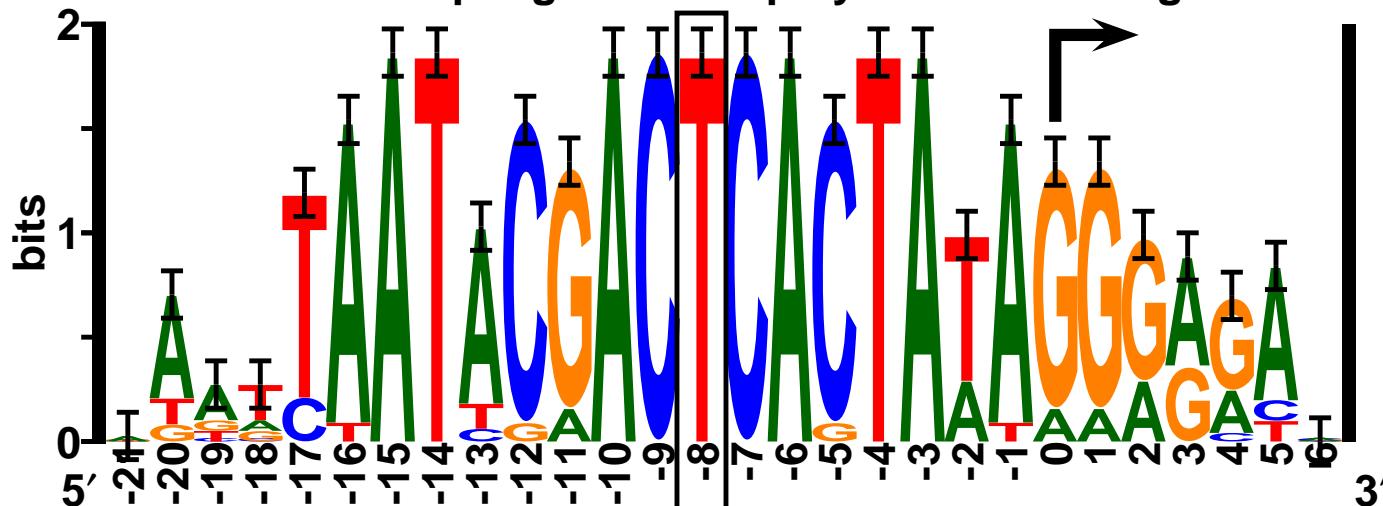
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3 cggtaataacgactcactataaggagaac
4 gaagtaataacgactcagtatagggacaa
5 taatttaatttgaactcactaaaggagac
6 cgcttaataacgactcactaaaggagaca

6 of 17 sites

Schneider &
Stephens
Nucl. Acids Res.
18: 6097-6100
1990

Sequence Logo

17 Bacteriophage T7 RNA polymerase binding sites



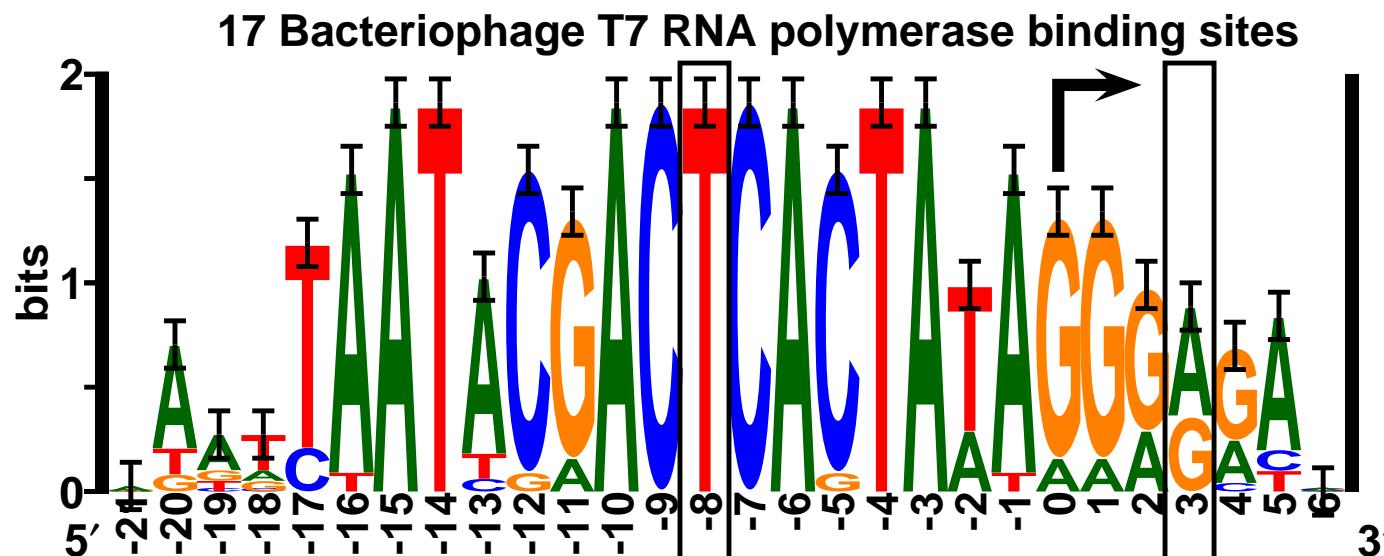
1 ttatttaataacaactcactataaggagag
2 aaatcaataacgactcactataggggac
3 cggtaataacgactcactataaggagaac
4 gaagtaataacgactcagtatagggacaa
5 taatttaattgaactcactaaaggagac
6 cgcttaataacgactcactaaaggagaca

6 of 17 sites

2 bits/base

Schneider &
Stephens
Nucl. Acids Res.
18: 6097-6100
1990

Sequence Logo



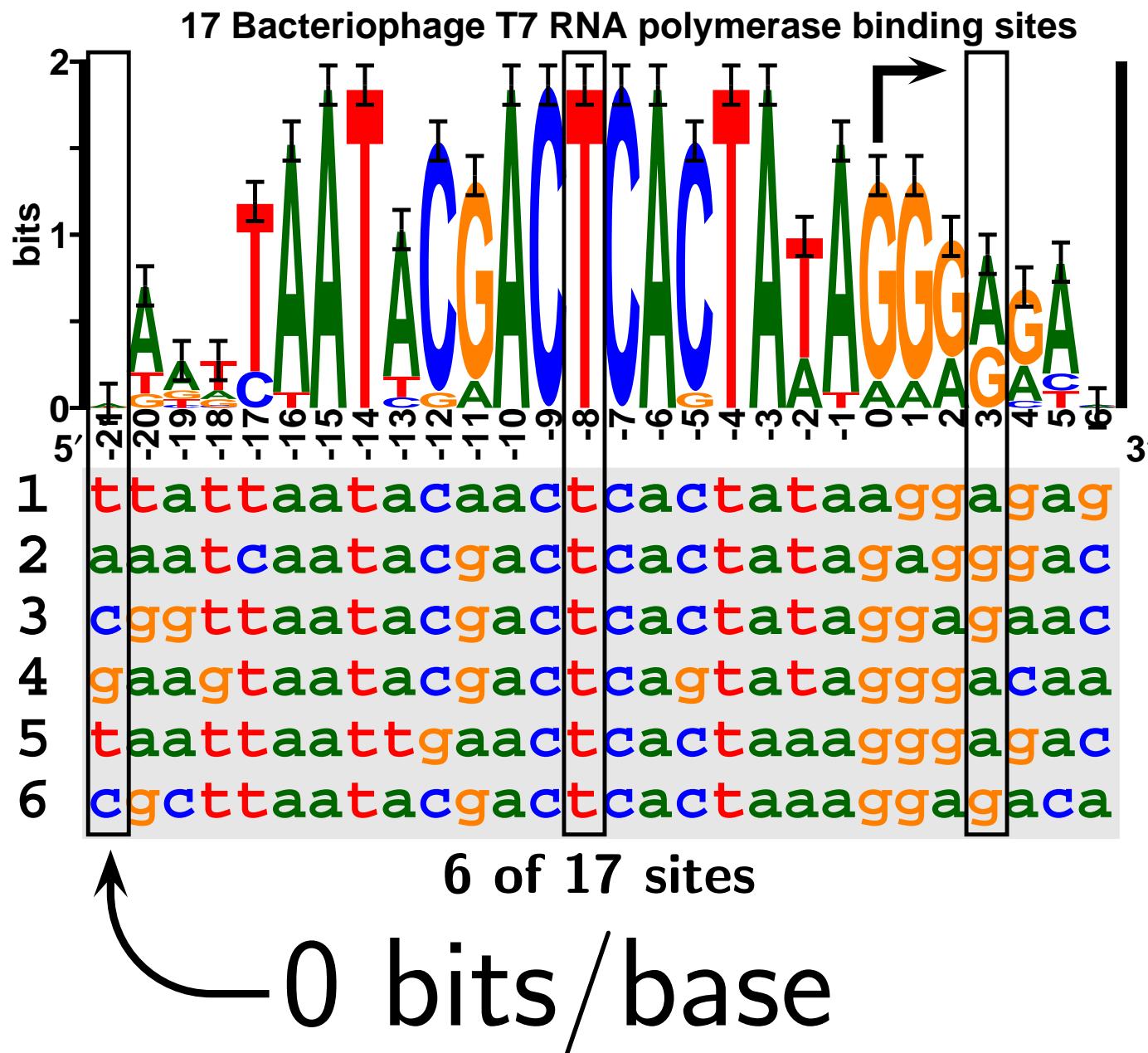
1 ttatttaataacaactcactataaggagag
2 aaatcaataacgactcactataagggac
3 cggtttaataacgactcactataaggagaac
4 gaagtaataacgactcagtataaggacaa
5 taatttaatttgaactcactaaaggagac
6 cgcttaataacgactcactaaaggagaca

6 of 17 sites

1 bit/base

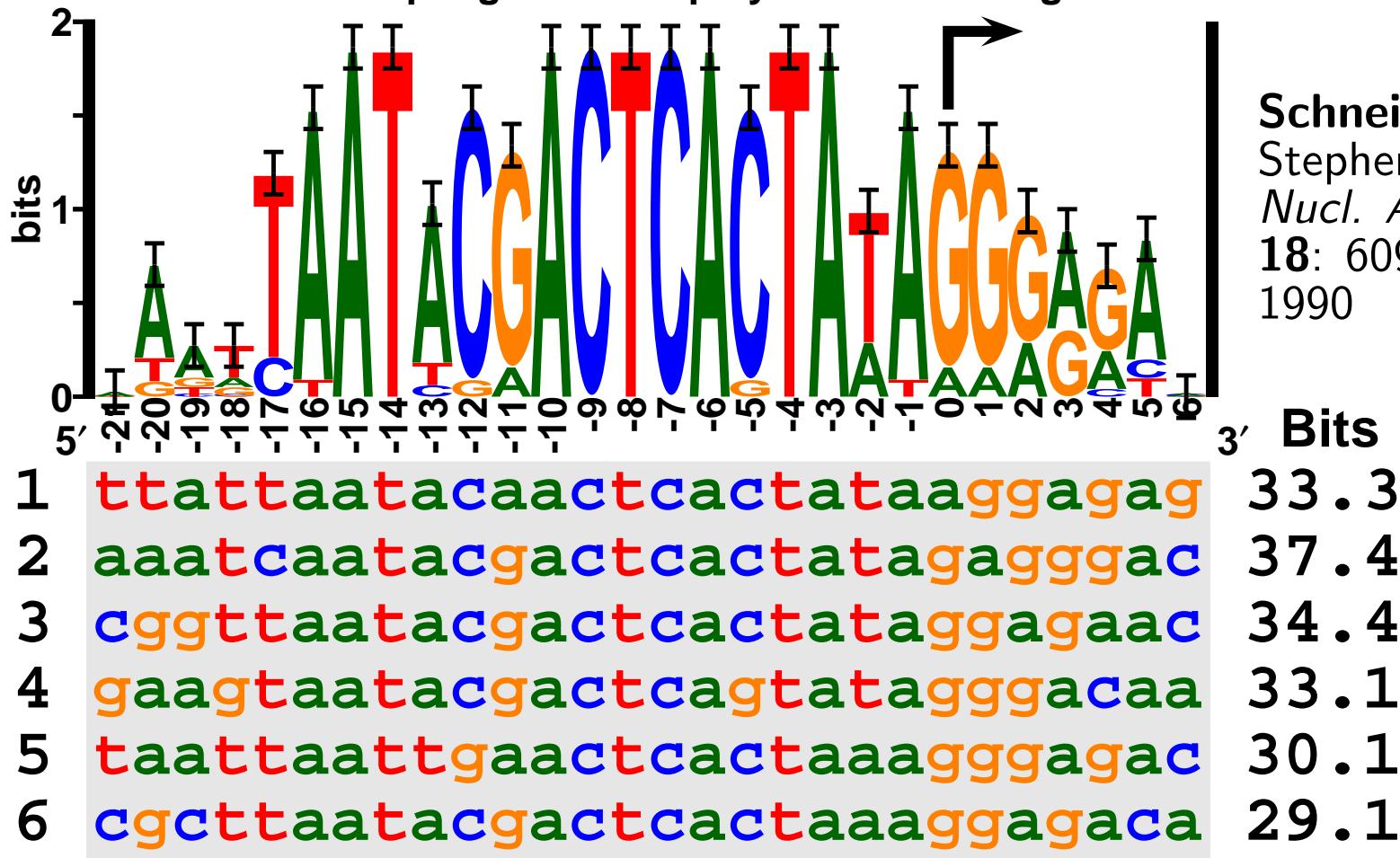
Schneider &
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Sequence Logo



Sequence Logo

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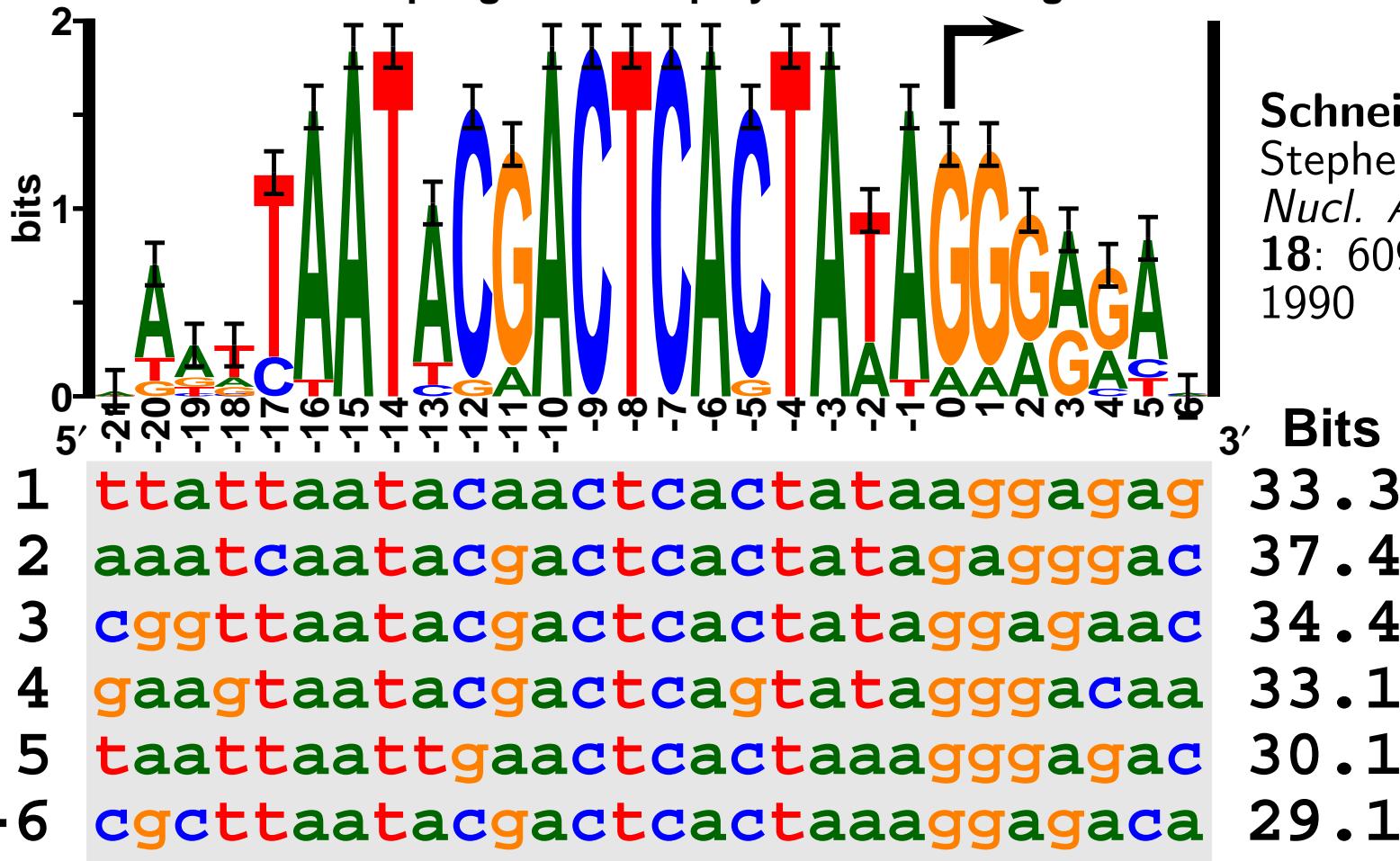


Schneider &
Stephens
Nucl. Acids Res.
18: 6097-6100
1990

Individual Information

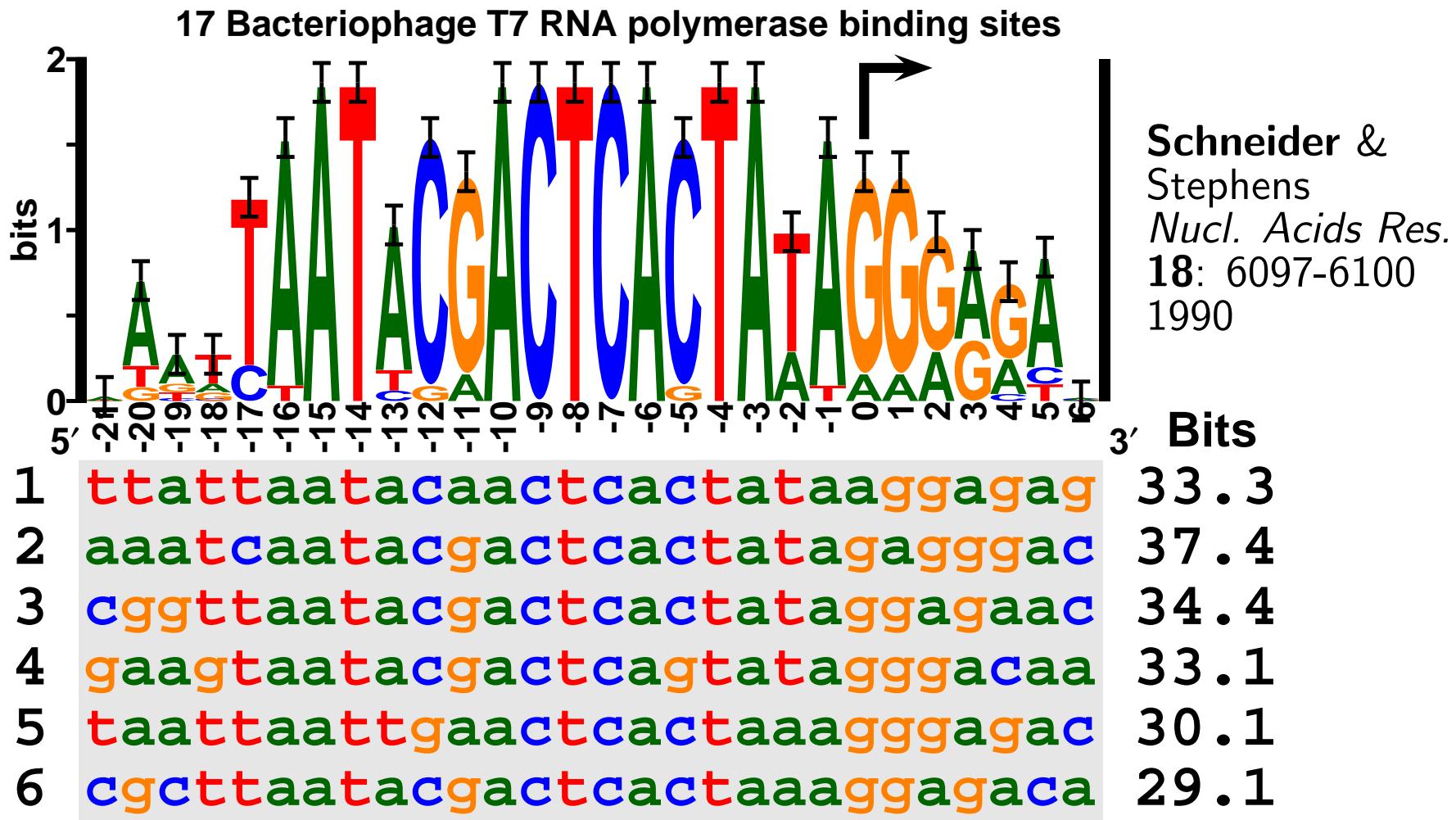
Sequence Logo and Sequence Walker

17 Bacteriophage T7 RNA polymerase binding sites



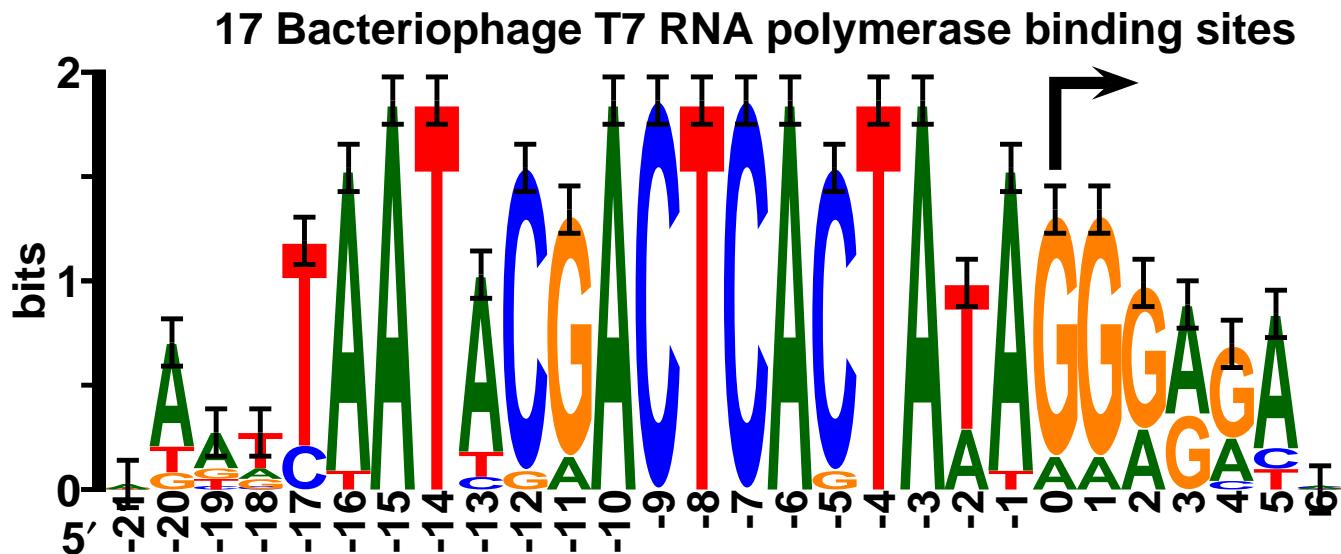
Sequence
Walker
Patent
5,867,402

Sequence Logo and Sequence Walker and Rsequence



Rsequence is the average: 35.0 ± 0.6 bits

Sequence Logo and Sequence Walker and Rsequence



Schneider &
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Nucl. Acids Res.
18: 6097-6100
1990

1	ttattaatacacaactcaactataaggagag	33.3
2	aaatcaataacgactcaactataaggagac	37.4
3	cggtaataacgactcaactataaggagaac	34.4
4	gaagtaataacgactcaactataaggacaa	33.1
5	taatttaatttgaactcaactaaaggagac	30.1
6	cgcttaataacgactcaactaaaggagaca	29.1

Rsequence is the average: 35.0 ± 0.6 bits
= “area under the logo”

**Information required
to find a set of binding sites**

$$G = \# \text{ of potential binding sites}$$

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= genome size in some cases

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Information required to find a set of binding sites

$G = \# \text{ of potential binding sites}$
 $= \text{ genome size in some cases}$

$\gamma = \text{ number of binding sites on genome}$

$$\begin{aligned} R_{frequency} &= H_{before} - H_{after} \\ &= \log_2 G - \log_2 \gamma \end{aligned}$$

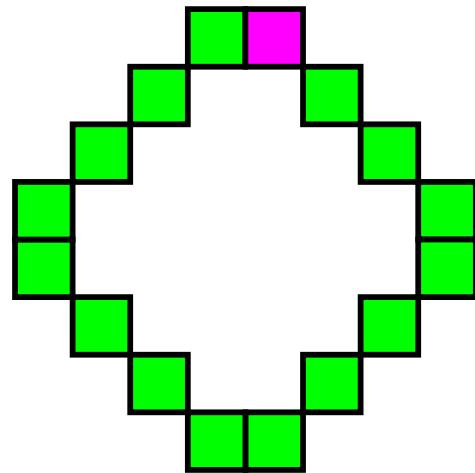
Information required to find a set of binding sites

$G = \# \text{ of potential binding sites}$
 $= \text{ genome size in some cases}$

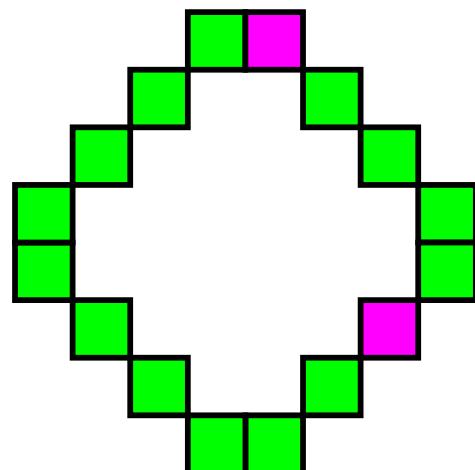
$\gamma = \text{ number of binding sites on genome}$

$$\begin{aligned} R_{frequency} &= H_{before} - H_{after} \\ &= \log_2 G - \log_2 \gamma \\ &= -\log_2 \gamma/G \end{aligned}$$

Information required to find a set of binding sites in a genome



16 positions
1 site
 $\log_2 16/1 = 4$ bits



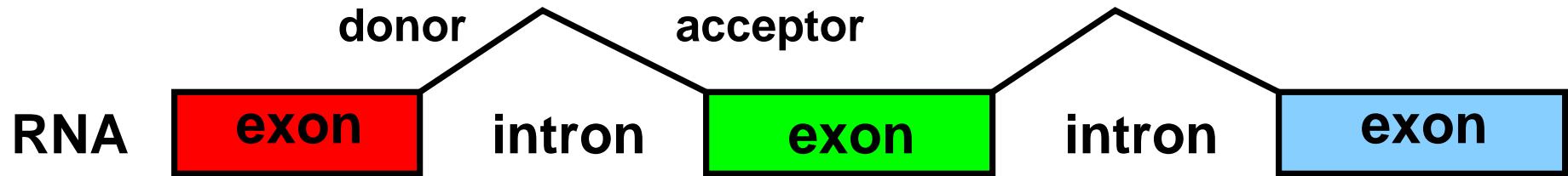
16 positions
2 sites
 $\log_2 16/2 = 3$ bits

RNA Splicing

DNA



Copy DNA (transcription)

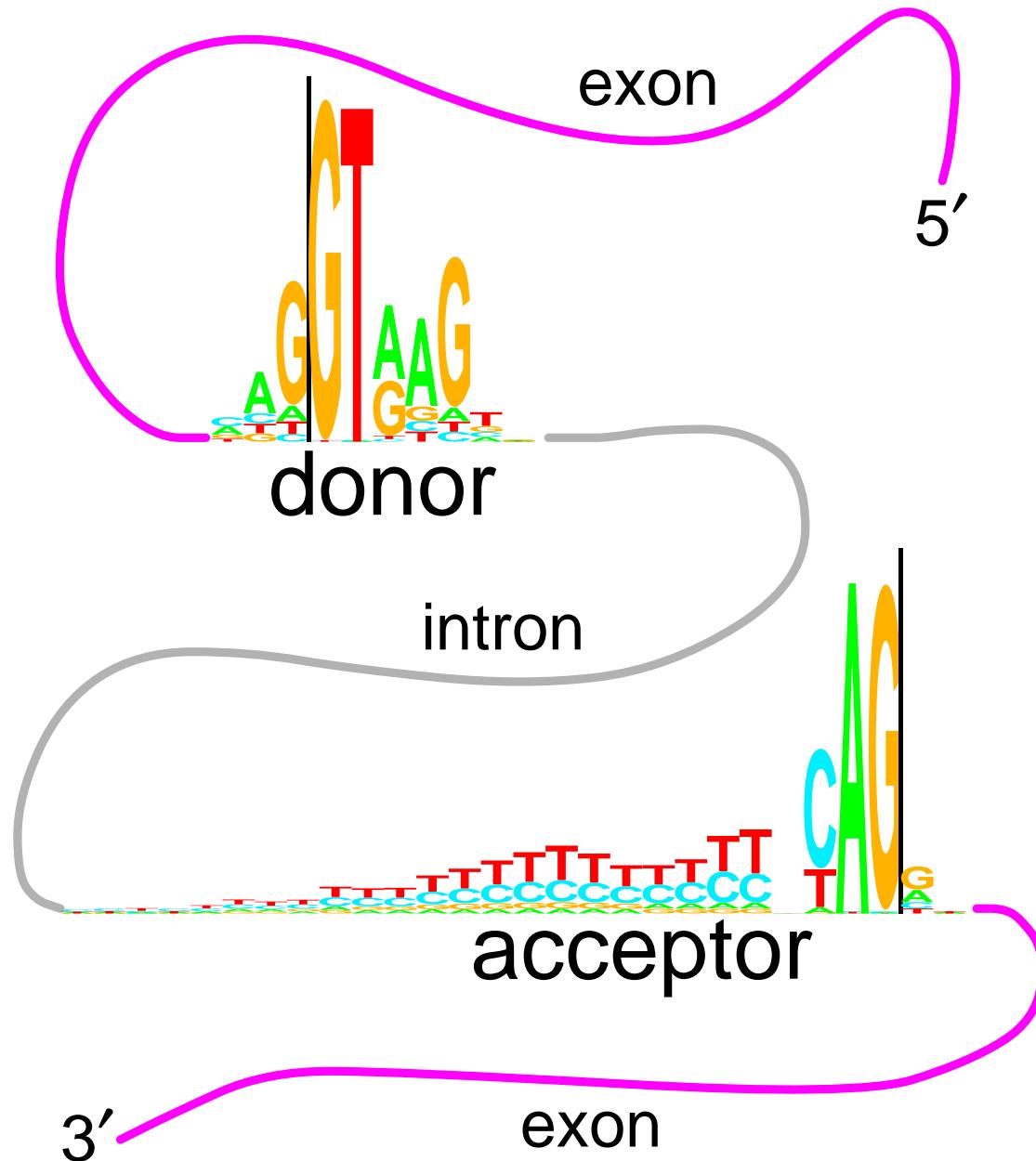


RNA Splicing

Spliced RNA

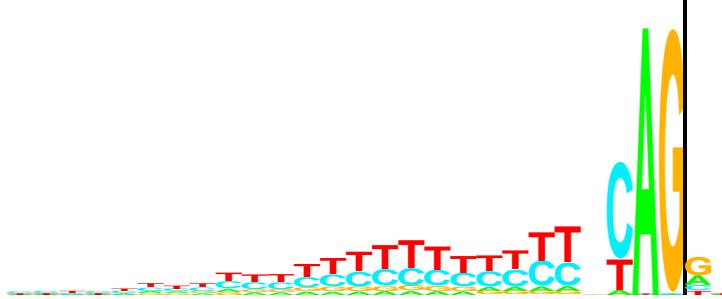


Donor and acceptor logos



Rsequence and Rfrequency for Splice Acceptors

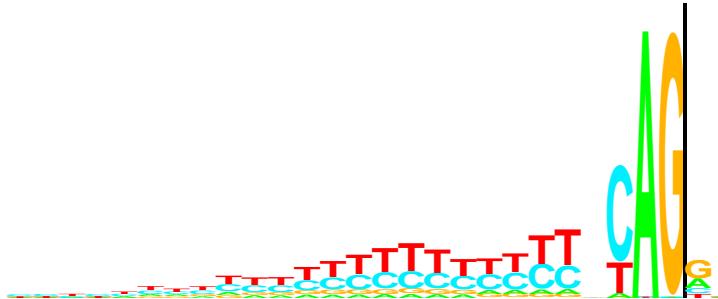
$R_{sequence}$



- Information at binding site sequences (area under sequence logo)
- from: binding site sequences
- 9.4 bits per site

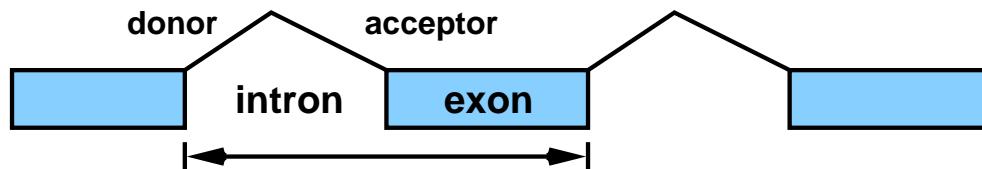
Rsequence and Rfrequency for Splice Acceptors

$R_{sequence}$



- Information at binding site sequences (area under sequence logo)
- from: binding site sequences
- 9.4 bits per site

$R_{frequency}$



- Information needed to locate the sites
- from: size of genome and number of sites (length of intron+exon)
- 9.7 bits per site

$$R_{frequency}/R_{sequence} = 0.97$$

Hypothesis:

The information in
binding site patterns
is just sufficient
for the sites to be found
in the genome

Rsequence versus Rfrequency

Binding Site Recognizer ¹	Total Pattern Information = R_{sequence} (bits)	Information needed to Locate Site in Genome = $R_{\text{frequency}}$ (bits)	Pattern Info / Location Info = $\frac{R_{\text{sequence}}}{R_{\text{frequency}}}$
Spliceosome acceptor ²	9.35 ± 0.12	9.66	0.97 ± 0.01
Spliceosome donor	7.92 ± 0.09	9.66	0.82 ± 0.01
Ribosome	11.0	10.6	1.0
λ cl/cro	17.7 ± 1.6	19.3	0.9 ± 0.1
LexA	21.5 ± 1.7	18.4	1.2 ± 0.1
TrpR	23.4 ± 1.9	20.3	1.2 ± 0.1
LacI	19.2 ± 2.8	21.9	0.9 ± 0.1
ArgR	16.4	18.4	0.9
O (λ Origin)	20.9	19.9	1.0
Ara C	19.3	19.3	1.0
Transcription at TATA ³	3.3	~ 3	~ 1
T7 Promoter	35.4	16.5	2.1

¹T. D. Schneider, G. D. Stormo, L. Gold, and A. Ehrenfeucht. J. Mol. Biol., 188:415-431, 1986.

²R. M. Stephens and T. D. Schneider. J. Mol. Biol., 228:1124-1136, 1992.

³F. E. Penotti. J Mol Biol, 213:37-52, 1990.

$R_{sequence}$ **versus** $R_{frequency}$ - meaning

The information in the binding site pattern ($R_{sequence}$)
is close to

The information needed to find the binding sites ($R_{frequency}$)

$R_{sequence}$ versus $R_{frequency}$ - meaning

The information in the binding site pattern ($R_{sequence}$)
is close to

The information needed to find the binding sites ($R_{frequency}$)

But for a species in a stable environment:

- size of genome (G) is fixed (e. g. *E. coli* has 4.7×10^6 bp)
- number of binding sites (γ) is fixed (e. g. there are ~ 50 *E. coli* LexA sites)

so $R_{frequency} = \log_2 G/\gamma$ is fixed

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so $R_{frequency} = \log_2 G/\gamma$ is fixed

Rsequence must evolve towards Rfrequency!

Evolution of Binding Sites

- $R_{frequency}$ is fixed relative to $R_{sequence}$

Evolution of Binding Sites

- $R_{frequency}$ is fixed relative to $R_{sequence}$
- Does $R_{sequence}$ evolve toward $R_{frequency}$?

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Setup a Computer Model, ‘Ev’:
A population of “creatures” with

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- genomes containing 4 bases (A, C, G, T)

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Setup a Computer Model, ‘Ev’:

A population of “creatures” with

- genomes containing 4 bases (A, C, G, T)
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- predetermined binding site locations (γ)
(to fix the frequency of sites)

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Evolution of Binding Sites

- $R_{frequency}$ is fixed relative to $R_{sequence}$
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Setup a Computer Model, ‘Ev’:

A population of “creatures” with

- genomes containing 4 bases (A, C, G, T)
 - a defined genome size (G)
 - predetermined binding site locations (γ)
(to fix the frequency of sites)
 - a recognizer gene encoded in the sequence:
use a weight matrix
- $R_{frequency}$
is fixed

How A Weight Matrix Works

Sequence matrix, $s(b, l, j)$ for sequence j

base b	position 1										
	C	A	G	G	T	C	T	G	C	A	
	-3	-2	-1	0	1	2	3	4	5	6	
A	0	1	0	0	0	0	0	0	0	1	
C	1	0	0	0	0	1	0	0	1	0	
G	0	0	1	1	0	0	0	1	0	0	
T	0	0	0	0	1	0	1	0	0	0	

Individual information weight matrix, $R_{iw}(b, l)$

base b	position 1											
	-3	-2	-1	0	1	2	3	4	5	6		
A	+0.4	+1.3	-1.4	-8.8	-5.8	+1.1	+1.5	-1.8	-0.7	+0.0		
C	+0.6	-0.8	-2.4	-7.8	-5.5	-3.7	-1.6	-2.2	-0.5	-0.2		
G	-0.6	-1.0	+1.6	+2.0	-6.2	+0.7	-1.1	+1.7	-0.3	+0.4		
T	-1.0	-0.9	-1.7	-5.8	+2.0	-3.4	-1.6	-2.2	+0.9	-0.5		

How A Weight Matrix Works

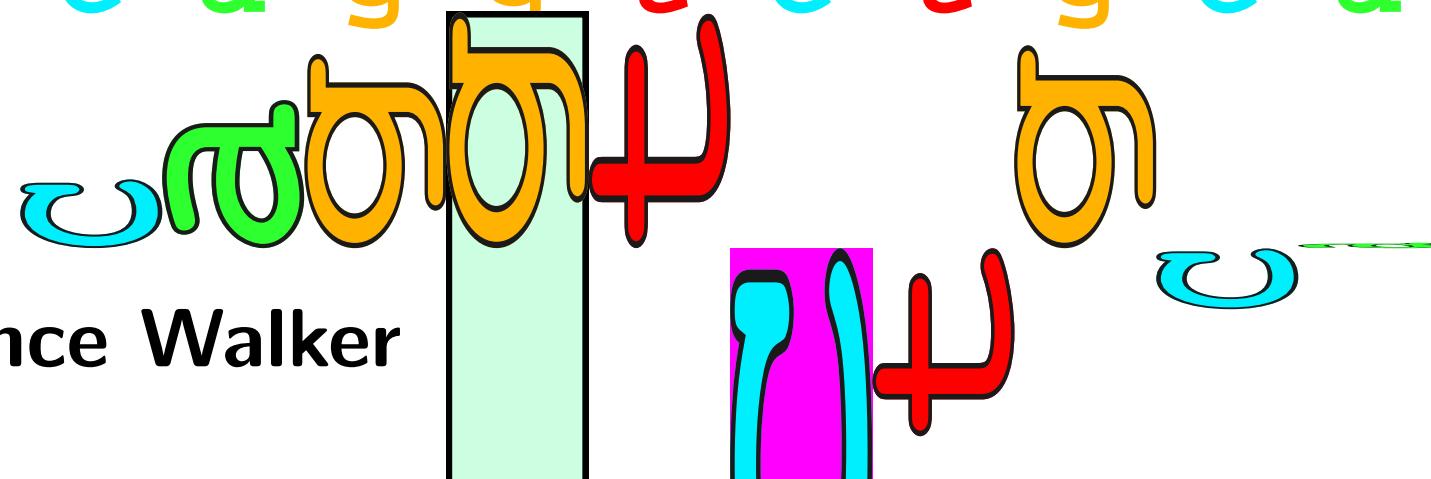
Sequence matrix, $s(b, l, j)$ for sequence j

base b	position 1										
	C	A	G	G	T	C	T	G	C	A	
-3	-2	-1	0	1	2	3	4	5	6		
A	0	1	0	0	0	0	0	0	0	1	
C	1	0	0	0	0	1	0	0	1	0	
G	0	0	1	1	0	0	0	1	0	0	
T	0	0	0	0	1	0	1	0	0	0	

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	-3	-2	-1	0	1	2	3	4	5	6		
A	+0.4	+1.3	-1.4	-8.8	-5.8	+1.1	+1.5	-1.8	-0.7	+0.0		
C	+0.6	-0.8	-2.4	-7.8	-5.5	-3.7	-1.6	-2.2	-0.5	-0.2		
G	-0.6	-1.0	+1.6	+2.0	-6.2	+0.7	-1.1	+1.7	-0.3	+0.4		
T	-1.0	-0.9	-1.7	-5.8	+2.0	-3.4	-1.6	-2.2	+0.9	-0.5		

5' c a g q t c t g c a 3'



Sequence Walker

Unevolved Ev Creature



Unevolved Ev Creature



“blue”
gene
weight
matrix:
6 bp
wide

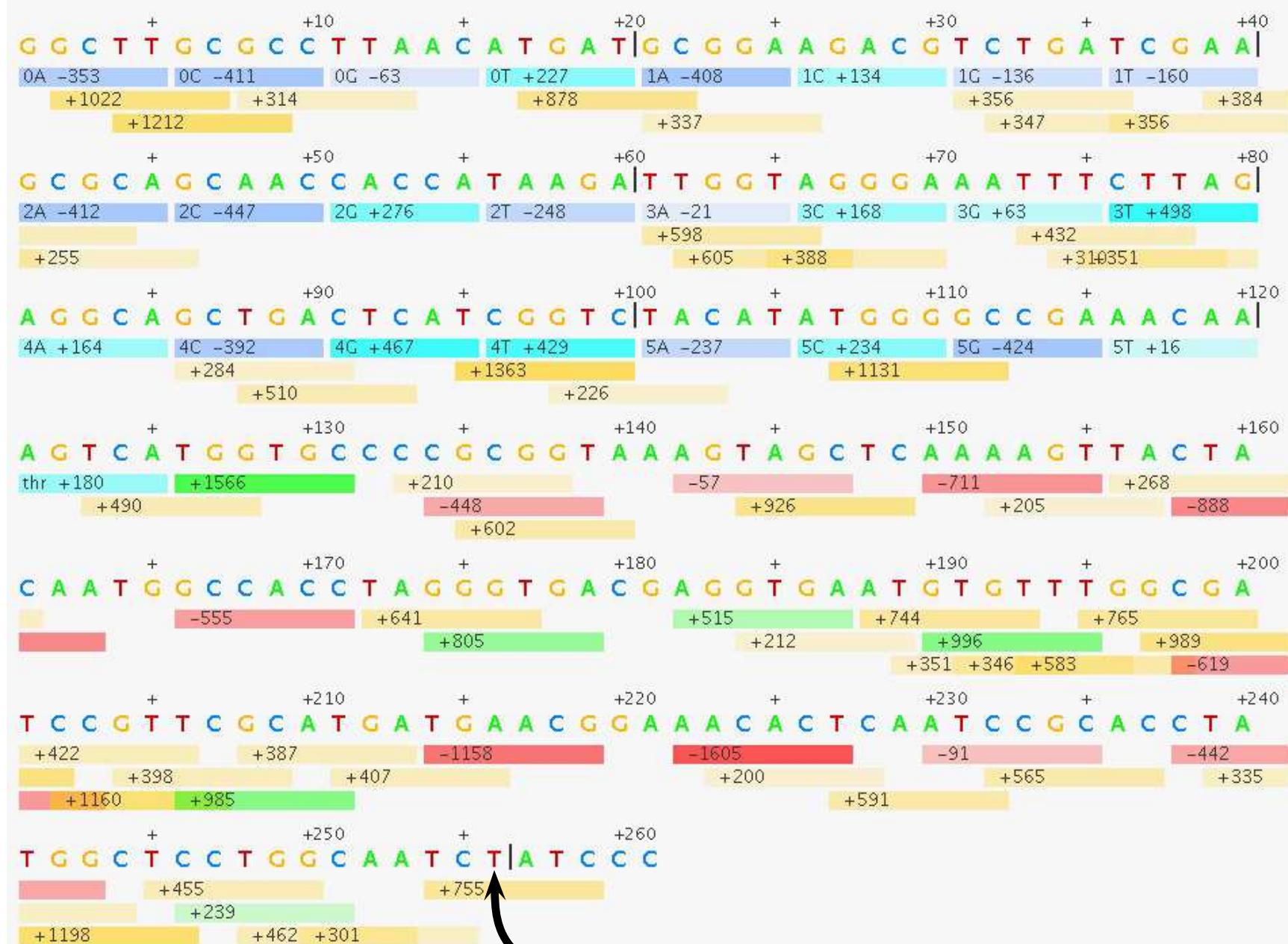
Unevolved Ev Creature



Genome positions available $G = 256$ bases

"blue"
gene
weight
matrix:
6 bp
wide

Unevolved Ev Creature

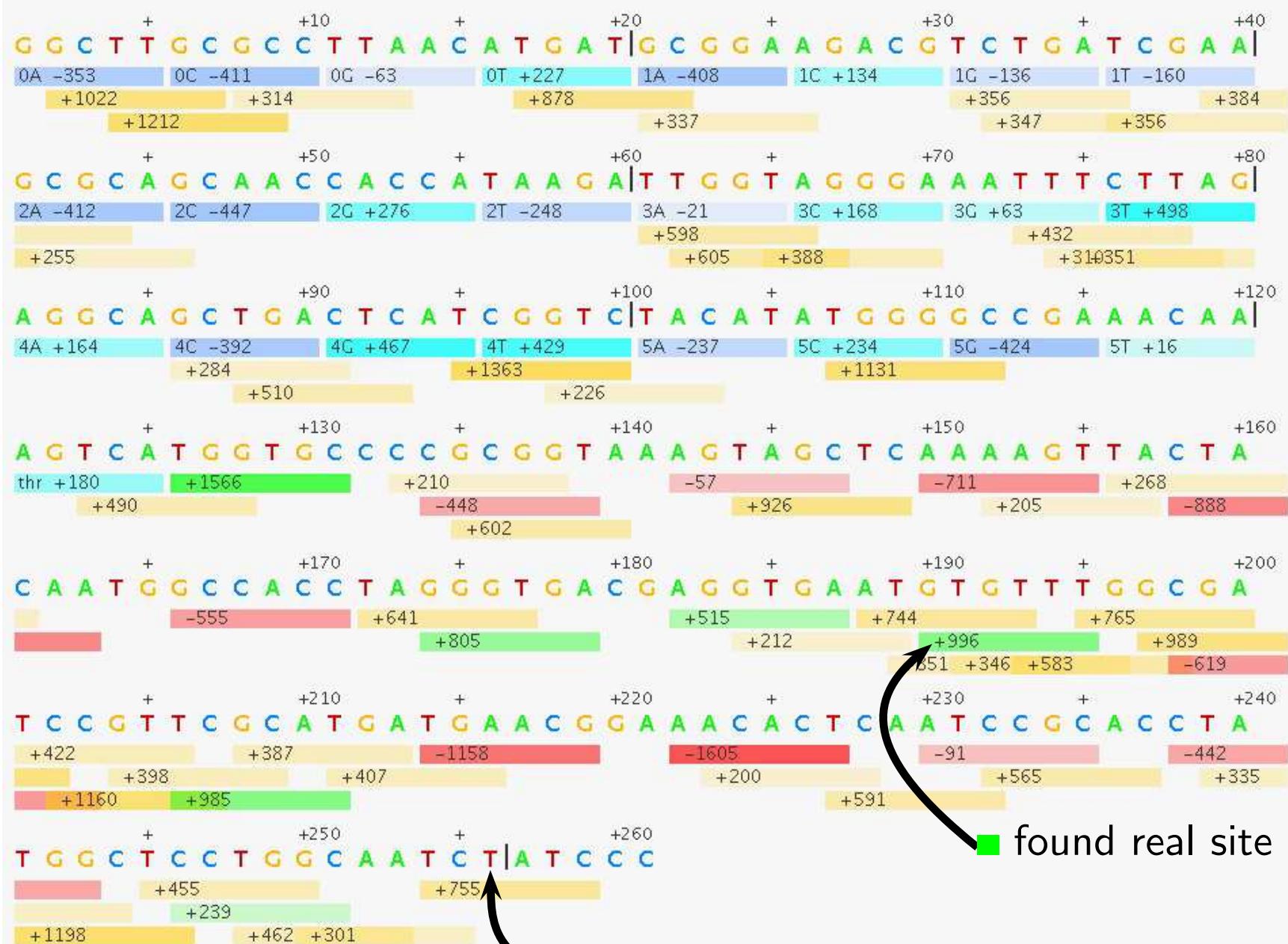


“blue”
gene
weight
matrix:
6 bp
wide

$\gamma = 16$
binding sites

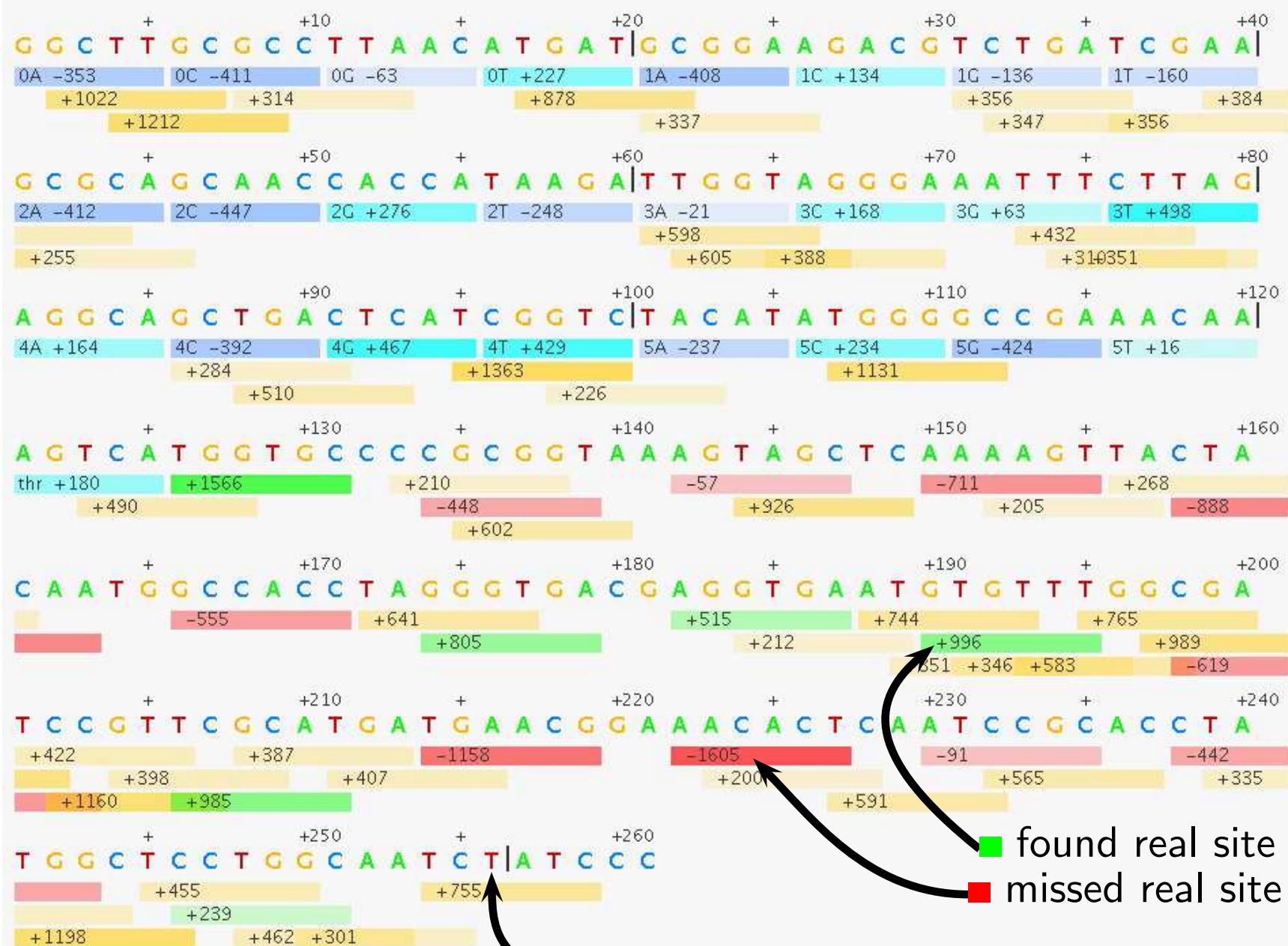
- Genome positions available $G = 256$ bases
 $R_{frequency} = \log_2 256/16 = 4$ bits

Unevolved Ev Creature



Genome positions available $G = 256$ bases
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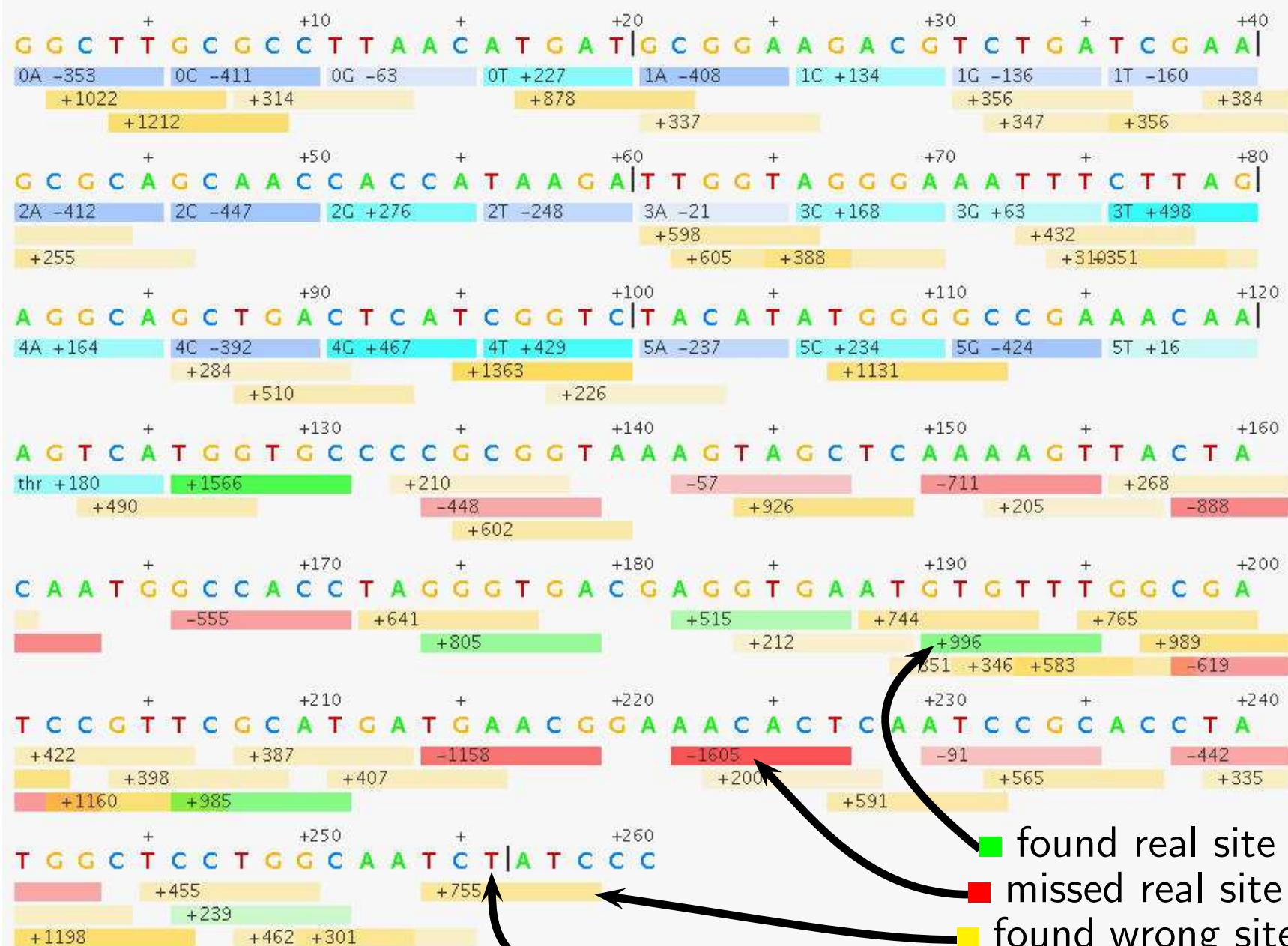
Unevolved Ev Creature



"blue"
gene
weight
matrix:
6 bp
wide

$\gamma = 16$
binding
sites

Unevolved Ev Creature



“blue”
gene
weight
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6 bp
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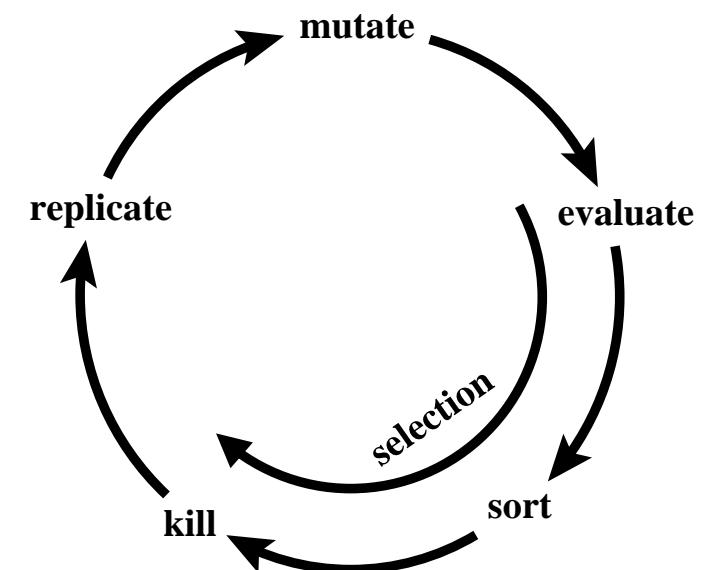
$\gamma = 16$
binding
sites

- found real site
- missed real site
- found wrong site

Genome positions available $G = 256$ bases
 $R_{frequency} = \log_2 256/16 = 4$ bits

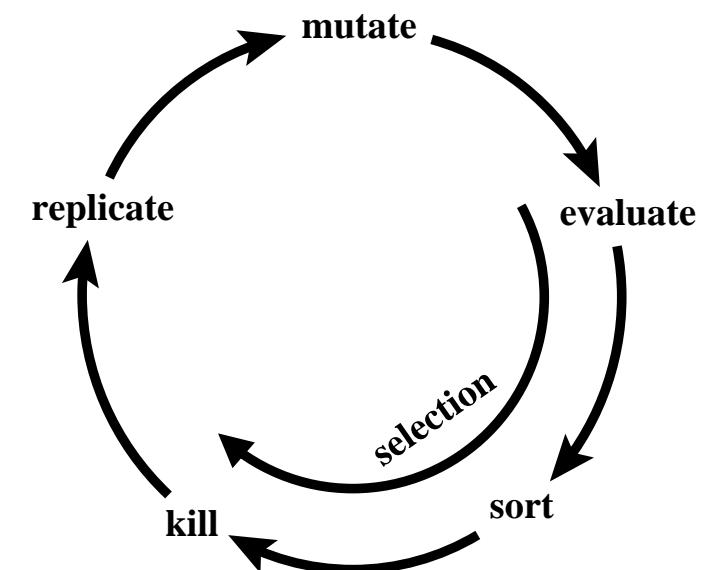
Evolution Cycle

- EVALUATE each creature



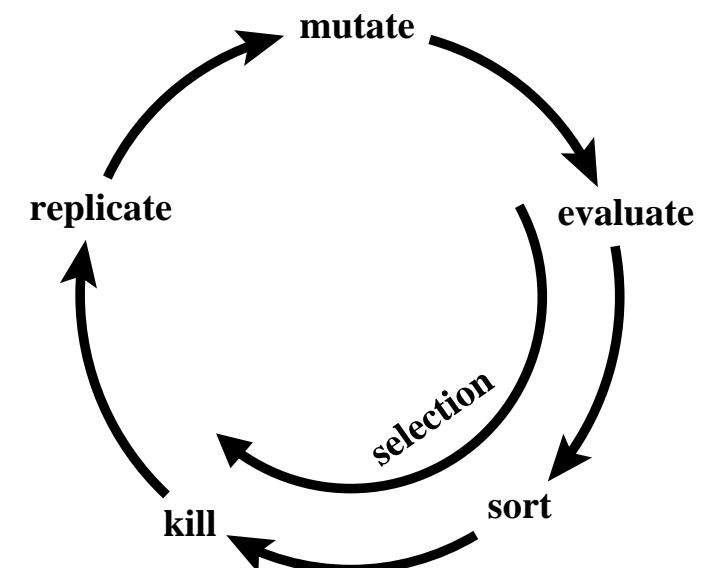
Evolution Cycle

- EVALUATE each creature
 - translate the recognizer gene into a weight matrix



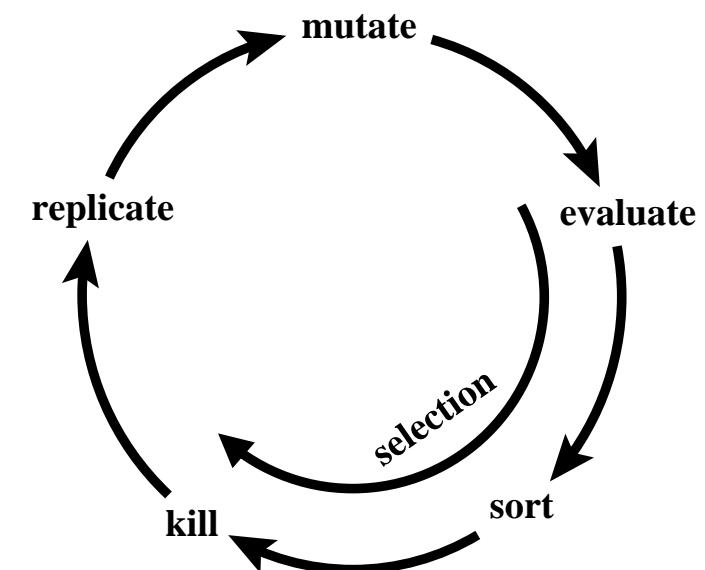
Evolution Cycle

- EVALUATE each creature
 - translate the recognizer gene into a weight matrix
 - scan the weight matrix across the genome



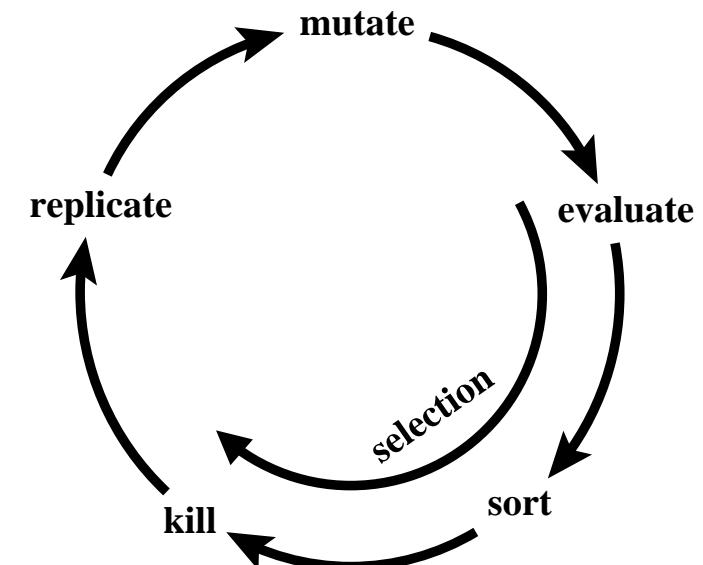
Evolution Cycle

- EVALUATE each creature
 - translate the recognizer gene into a weight matrix
 - scan the weight matrix across the genome
 - count the number of mistakes:



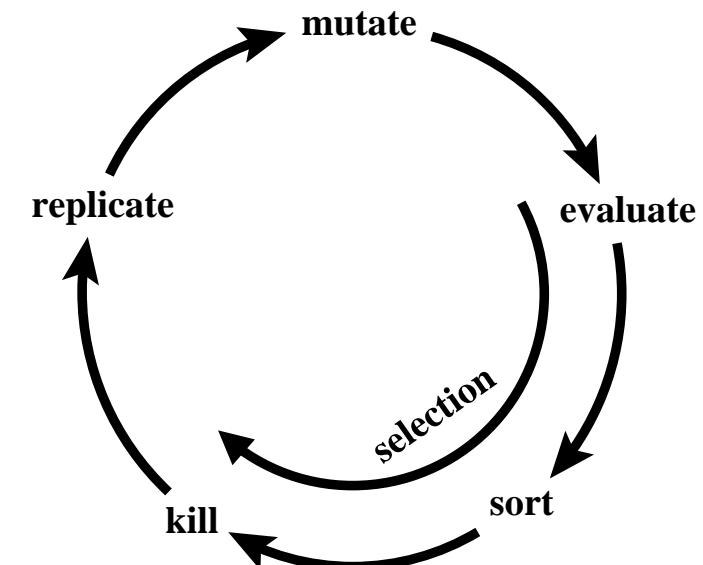
Evolution Cycle

- EVALUATE each creature
 - translate the recognizer gene into a weight matrix
 - scan the weight matrix across the genome
 - count the number of mistakes:
 - missing a site at a right place



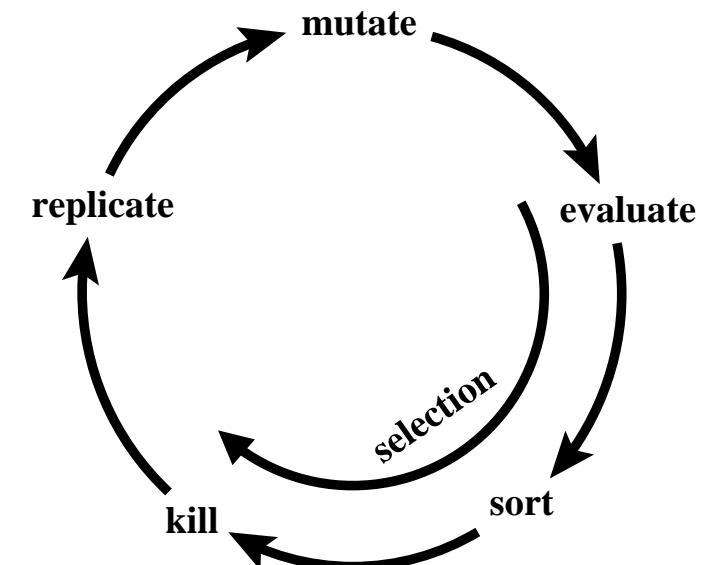
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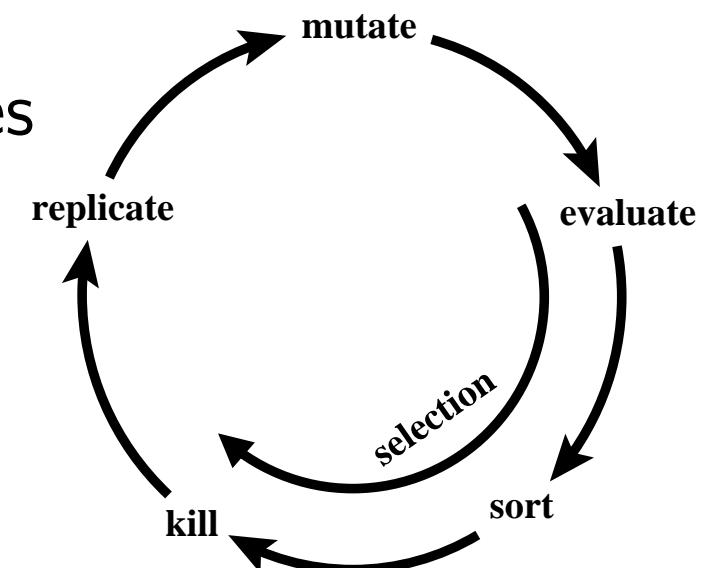
Evolution Cycle

- EVALUATE each creature
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 - missing a site at a right place
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 - Sort the creatures by their mistakes



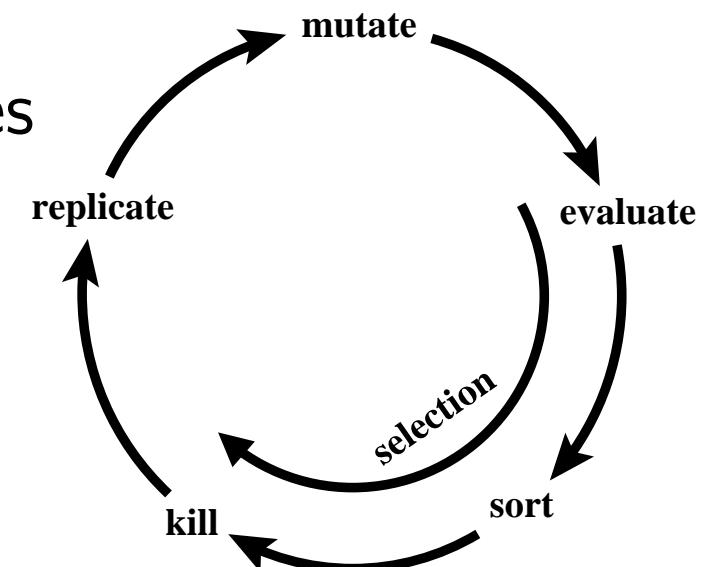
Evolution Cycle

- EVALUATE each creature
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- REPLICATE: the best creatures are duplicated and replace the worst ones

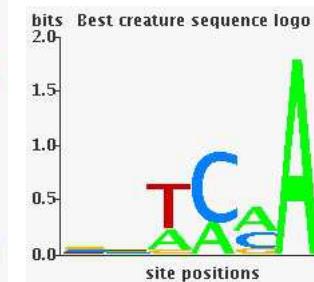
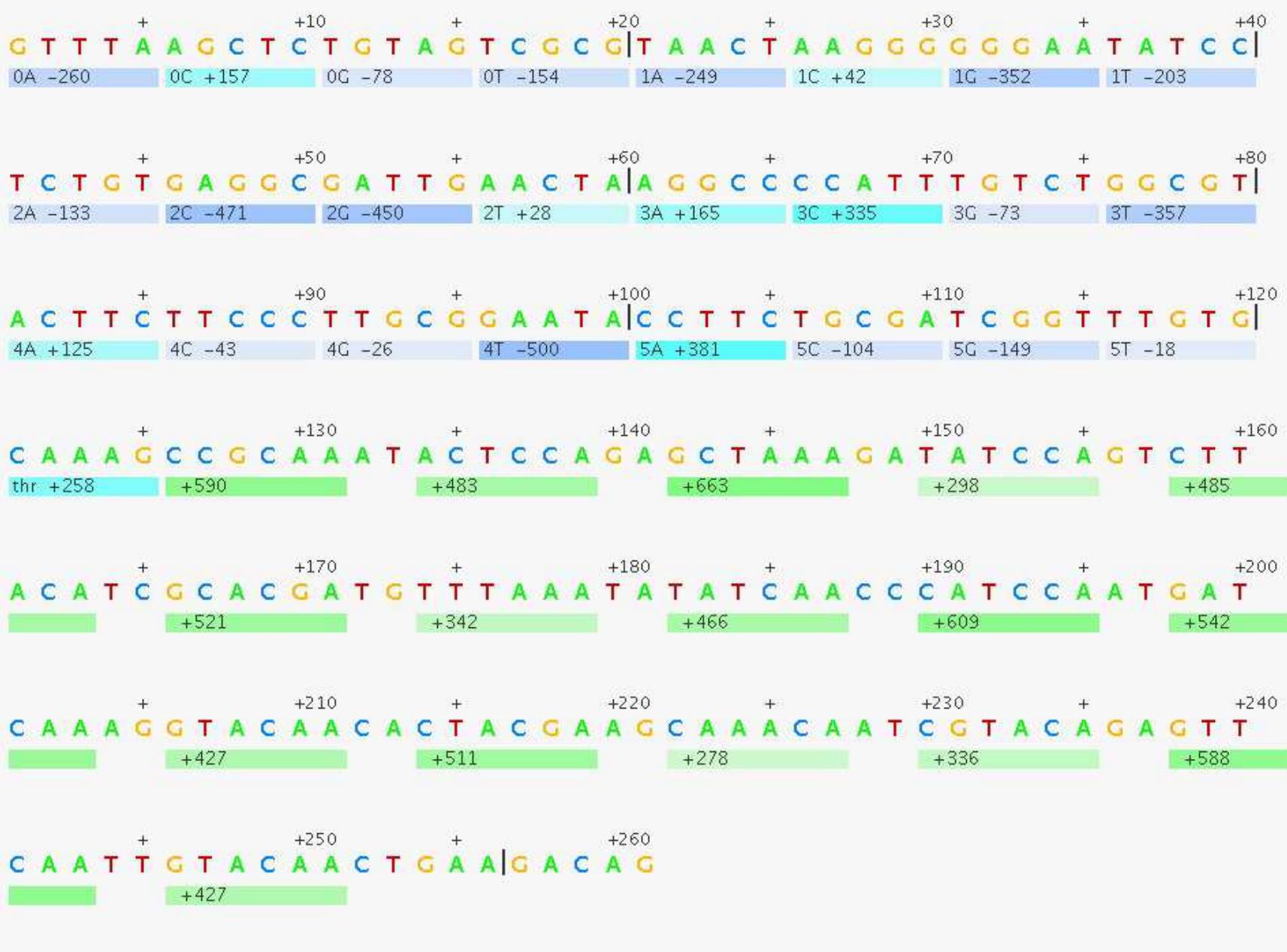


Evolution Cycle

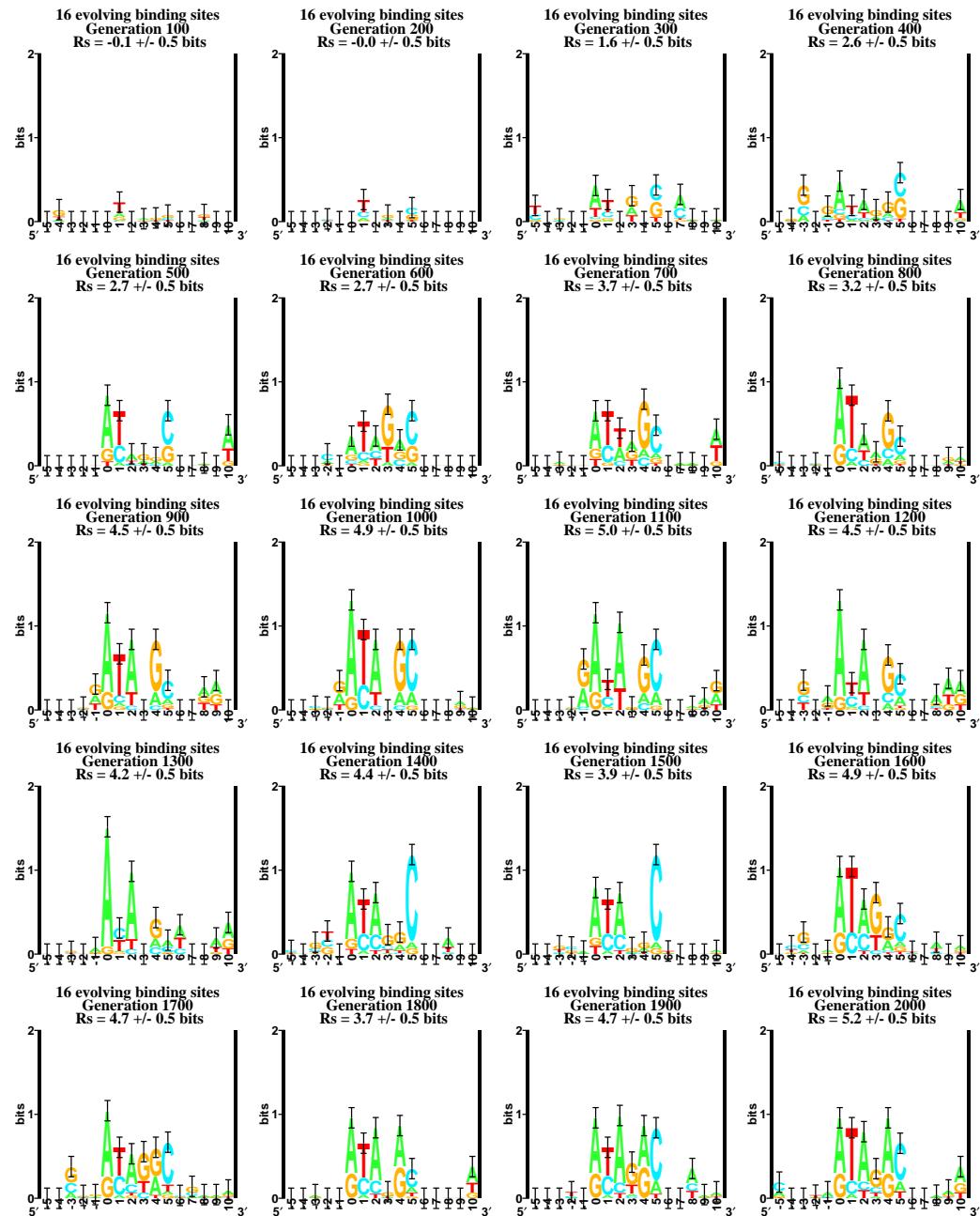
- EVALUATE each creature
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 - missing a site at a right place
 - finding a site at a wrong place
 - Sort the creatures by their mistakes
- REPLICATE: the best creatures are duplicated and replace the worst ones
- MUTATE all genomes randomly



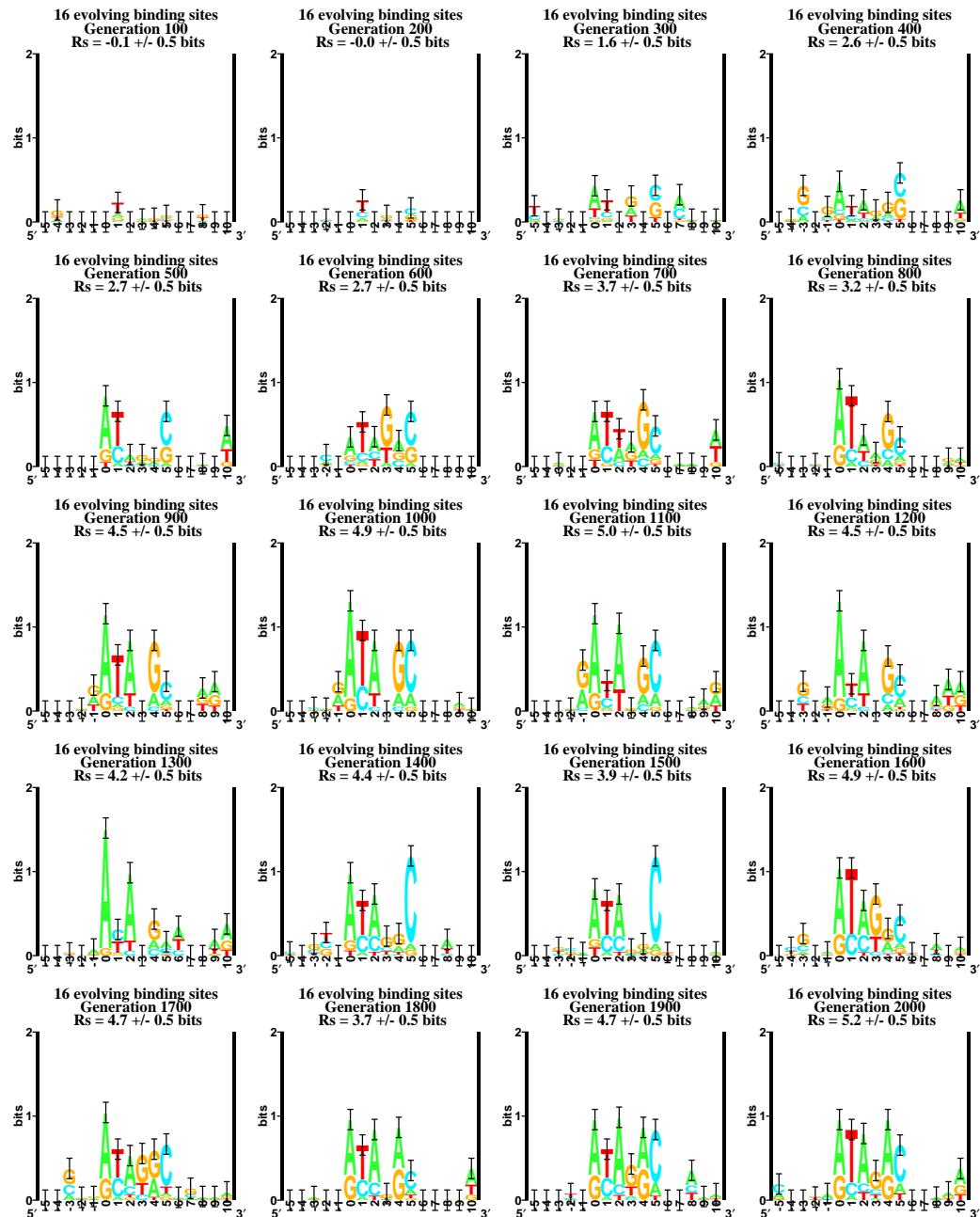
Evolved Ev Creature



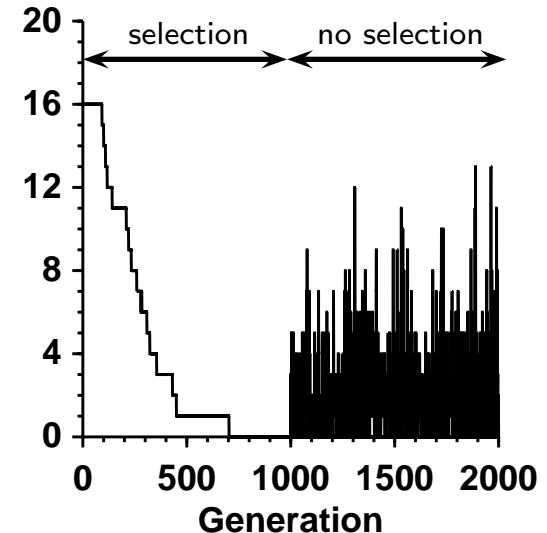
Evolution of Binding Sites



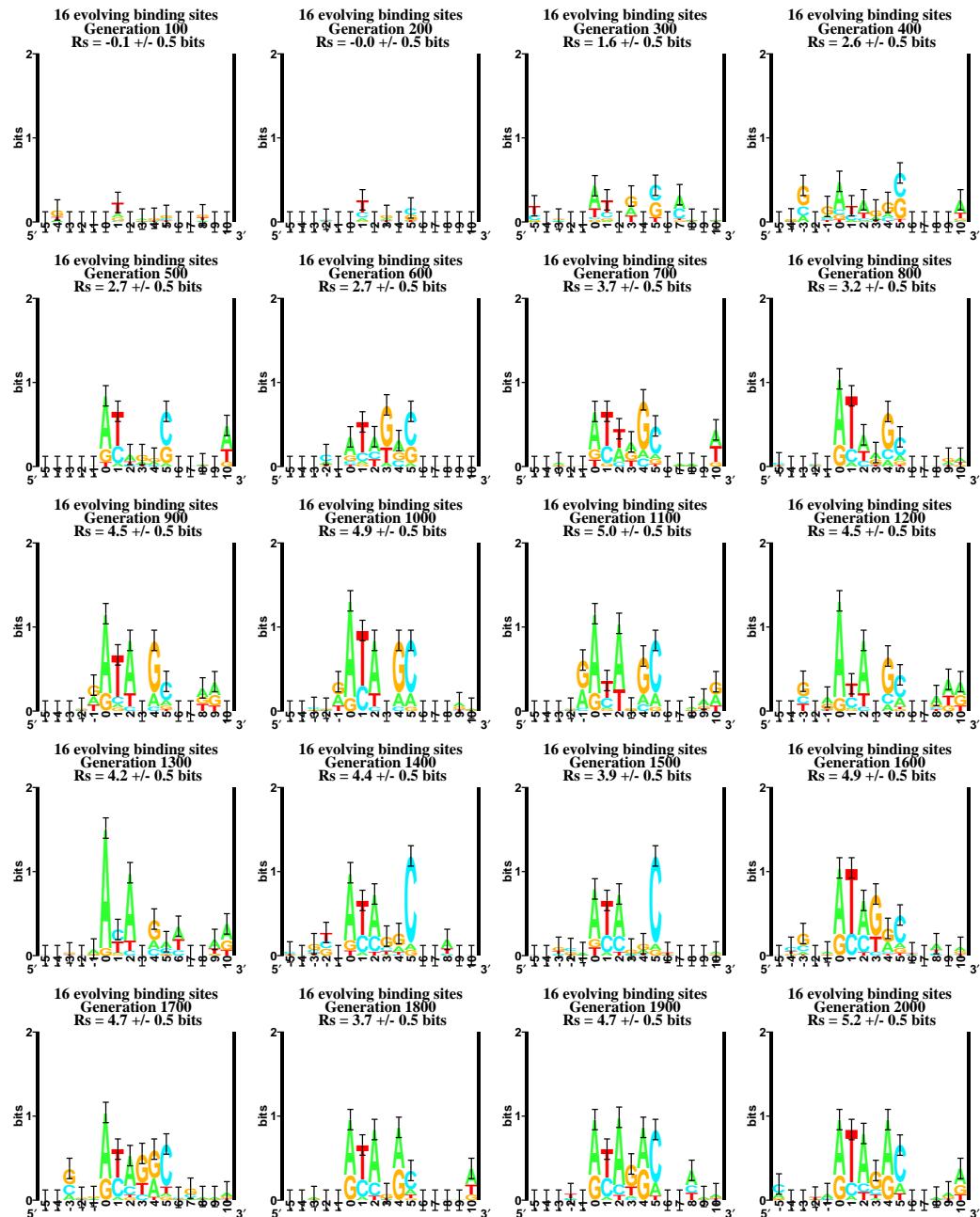
Evolution of Binding Sites



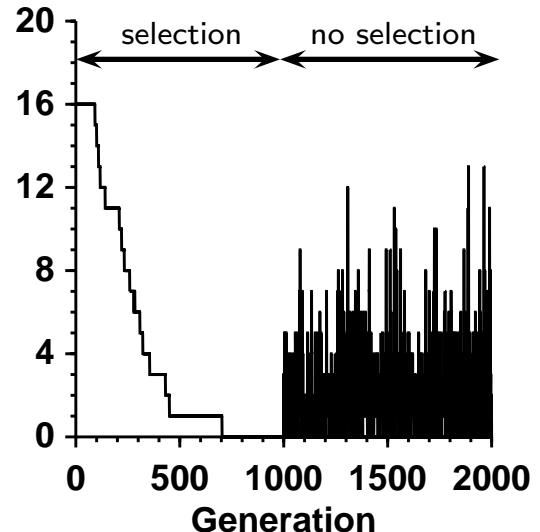
Mistakes of Best Organism



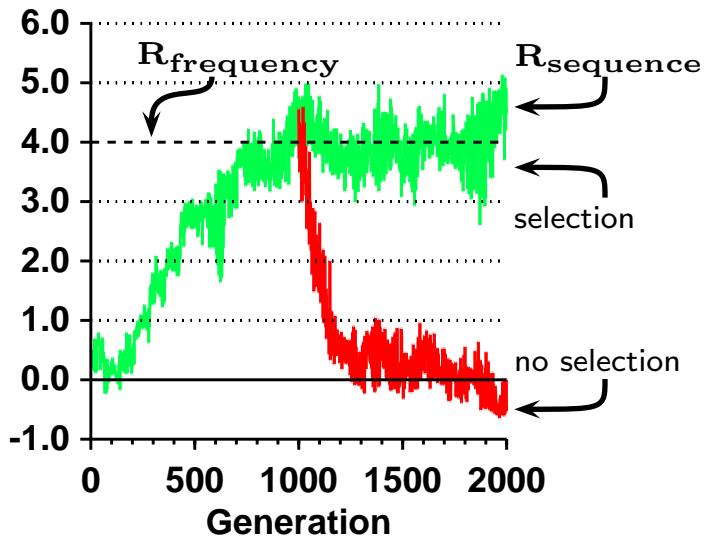
Evolution of Binding Sites



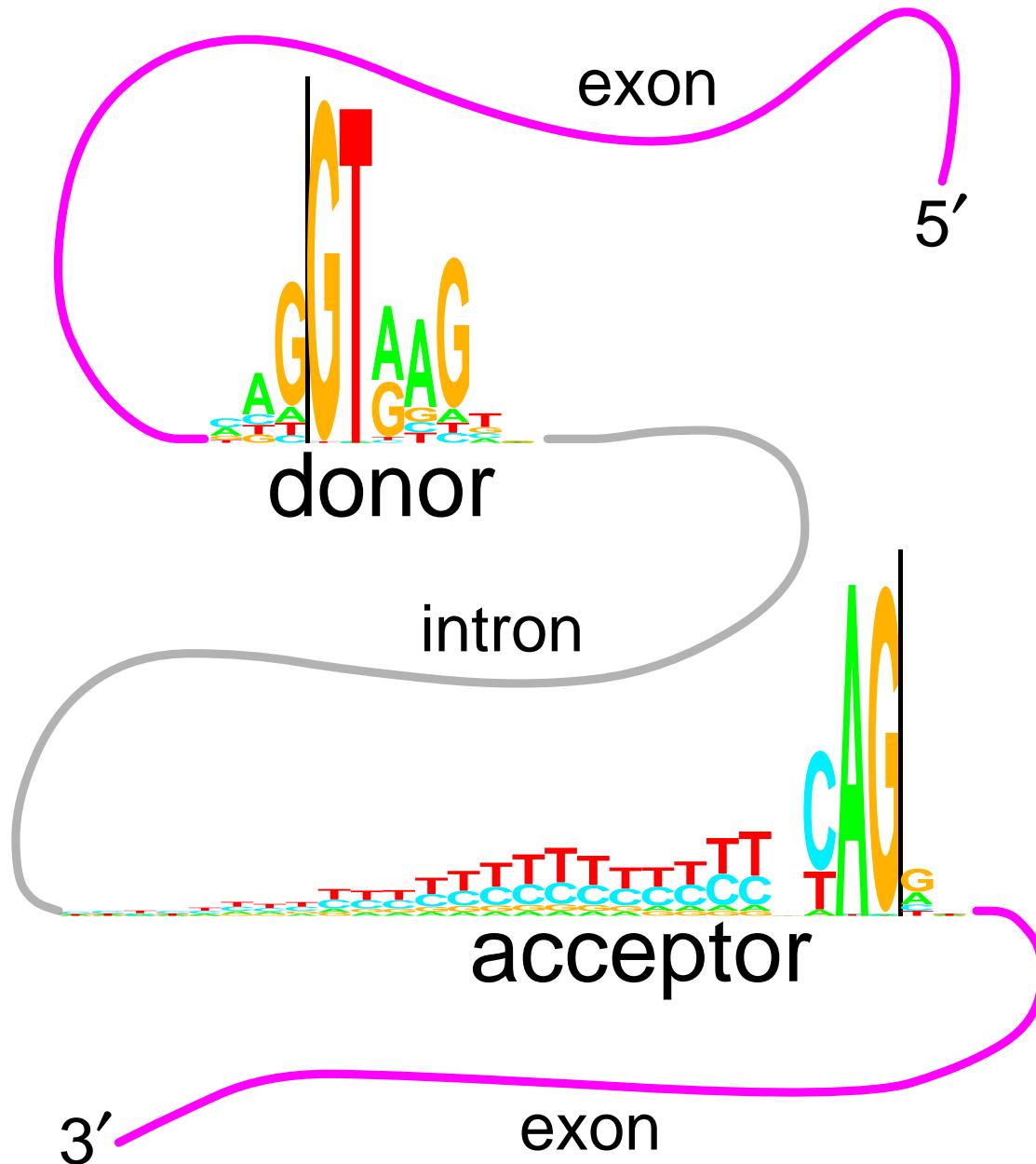
Mistakes of Best Organism



Information (bits per site)

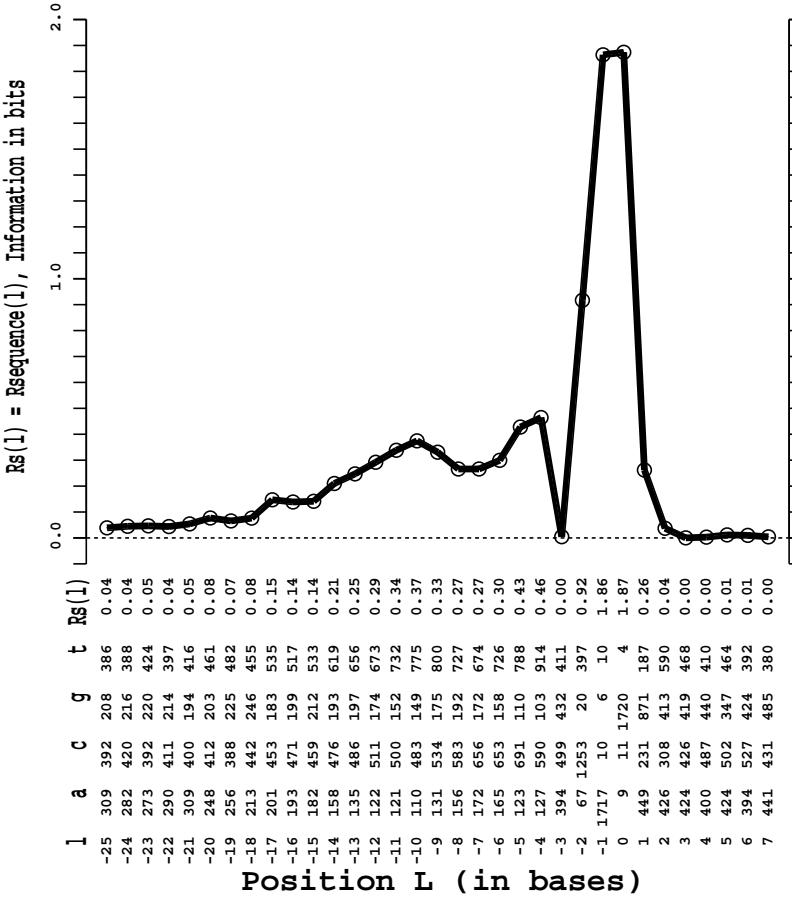
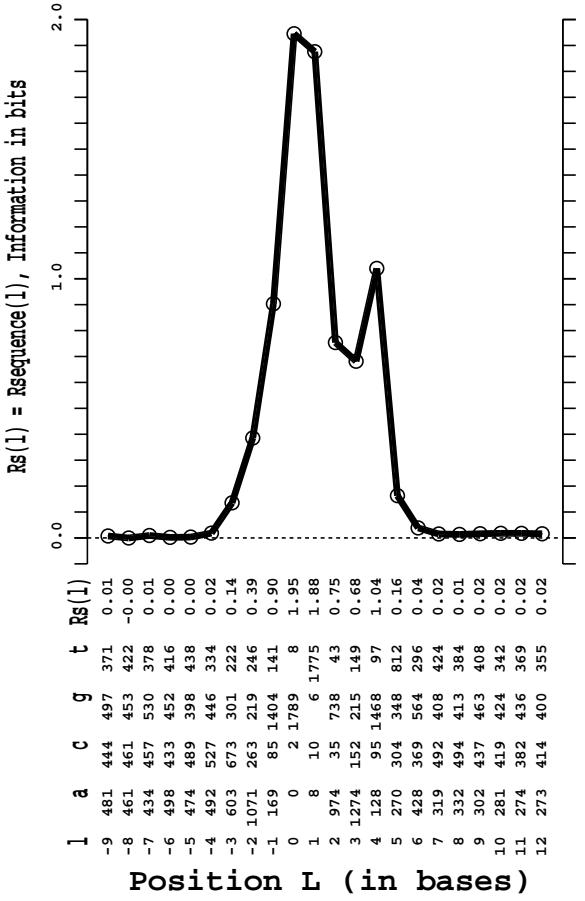


Donor and acceptor logos



Sequence Conservation → in bits per base

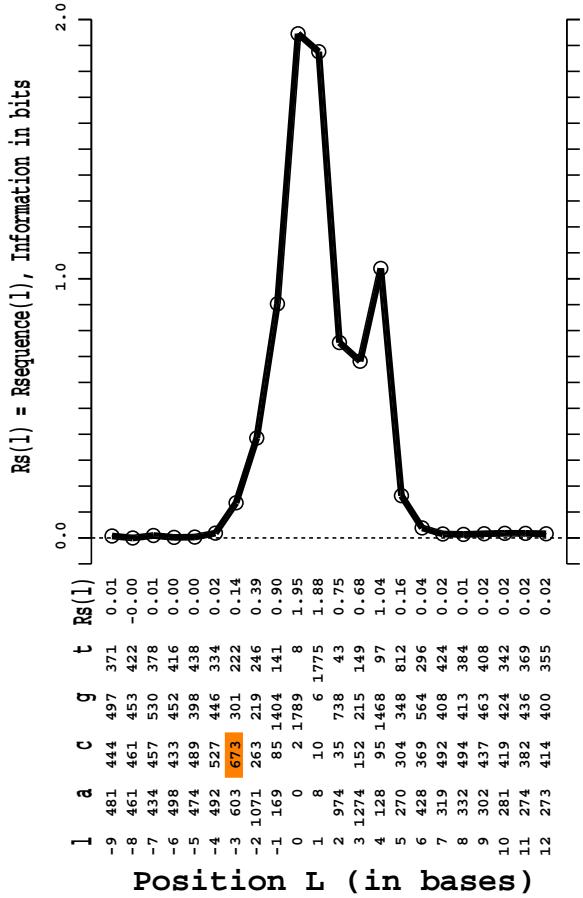
Donor



Acceptor

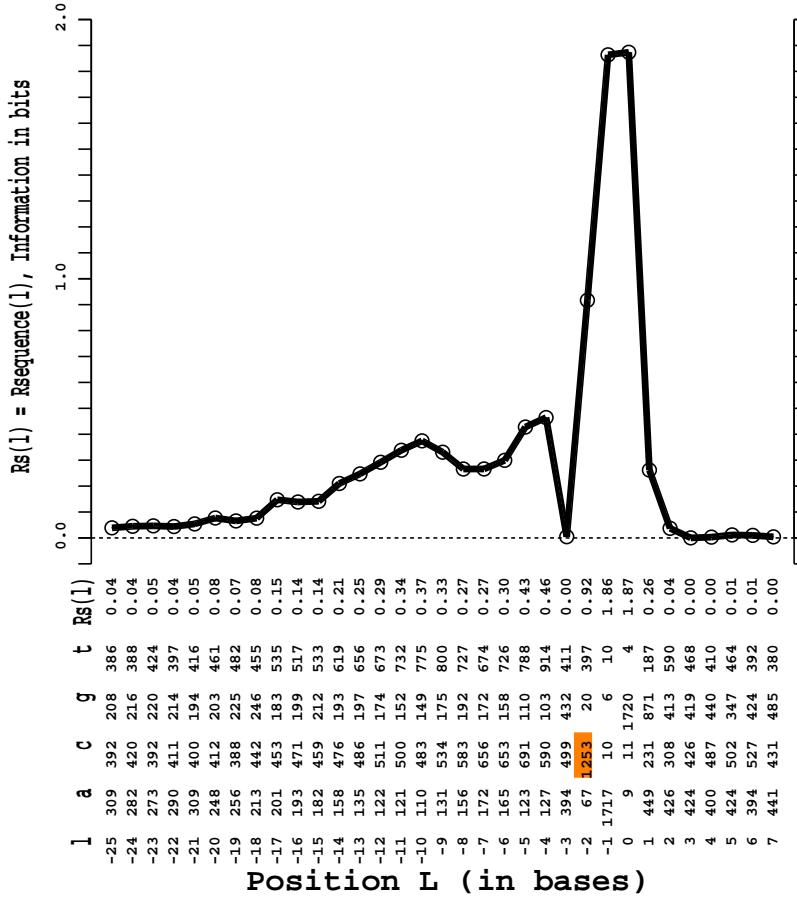
Sequence Conservation → in bits per base

Donor



C

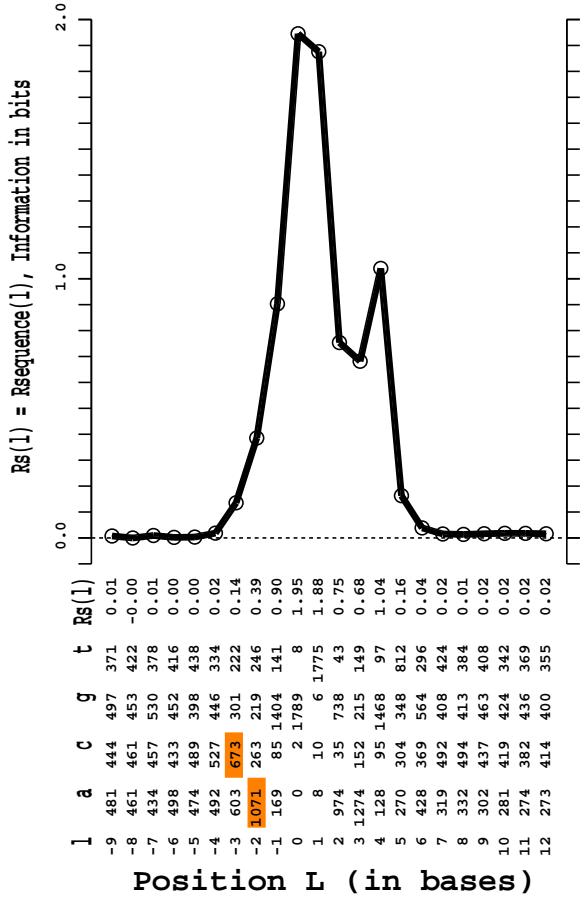
- The consensus sequences match ...



Acceptor

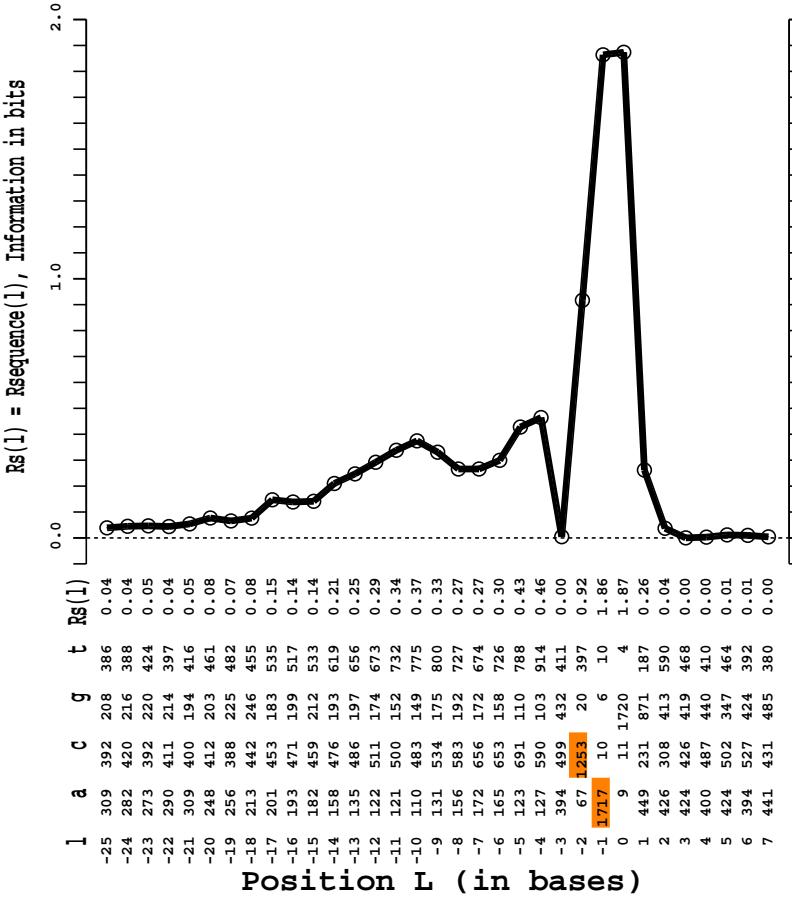
Sequence Conservation → in bits per base

Donor



C A

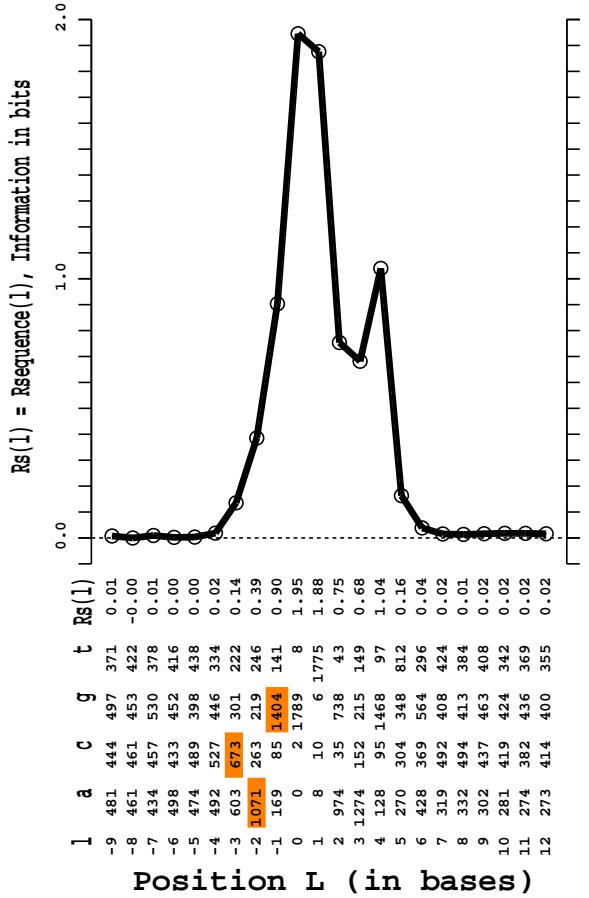
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Acceptor

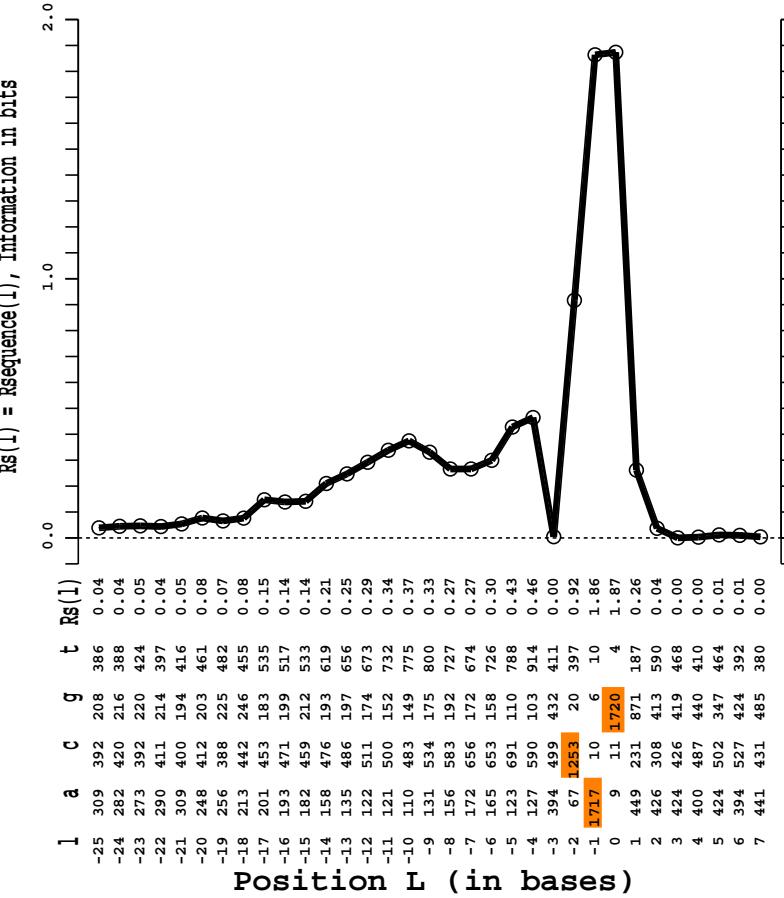
Sequence Conservation → in bits per base

Donor



C A G

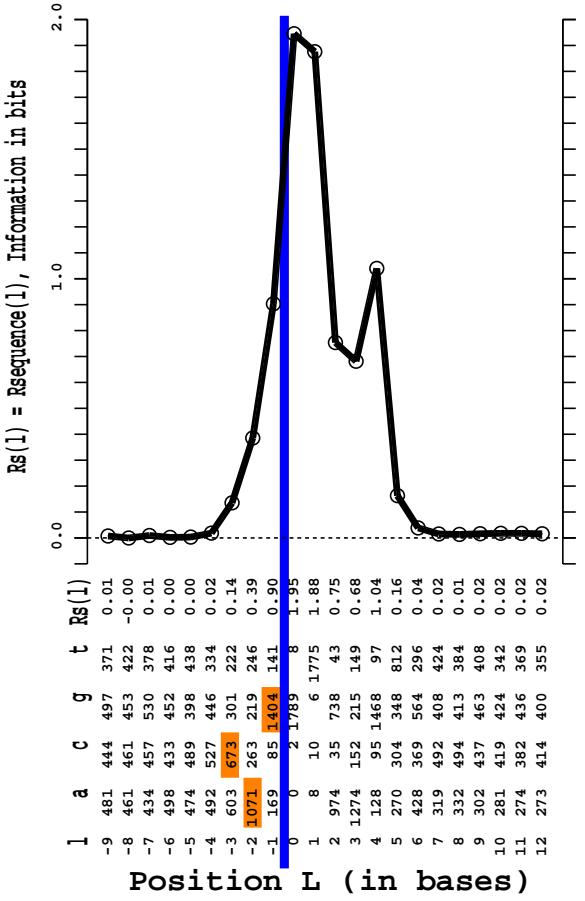
- The consensus sequences match ...



Acceptor

Sequence Conservation → in bits per base

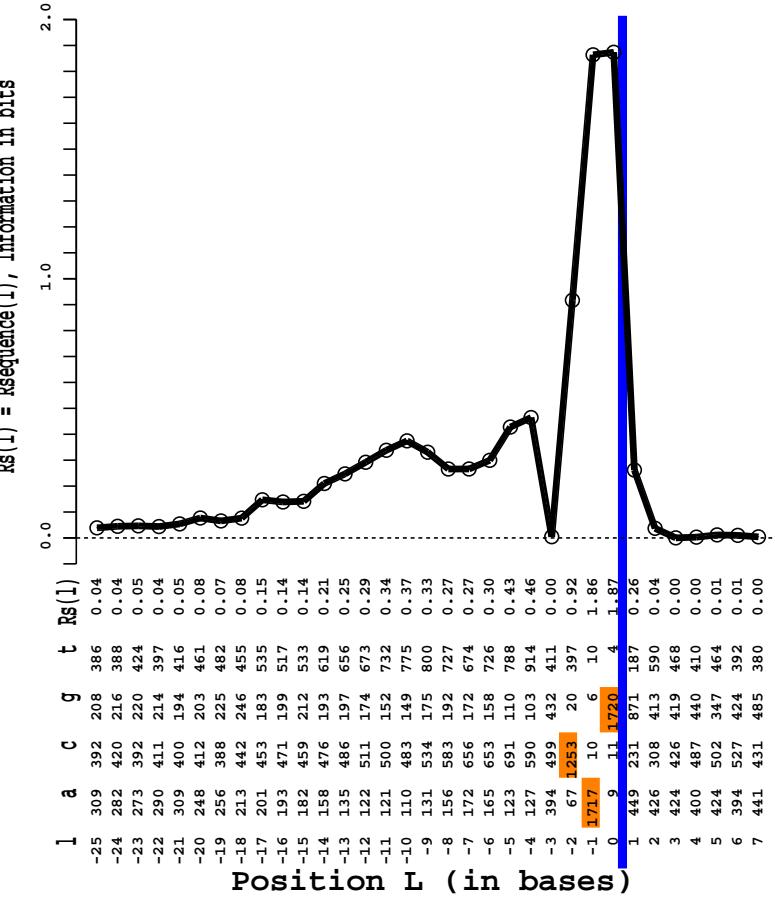
Donor



C A G —

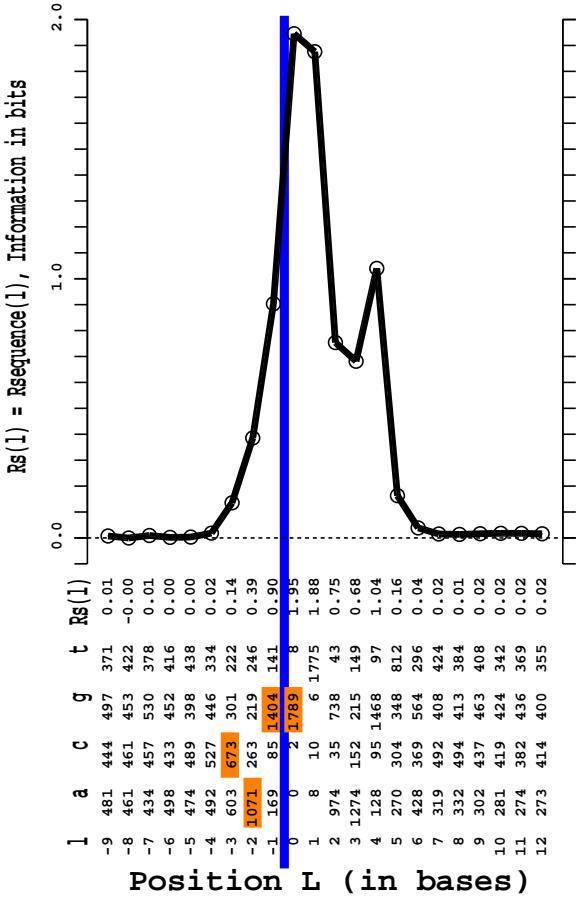
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Acceptor



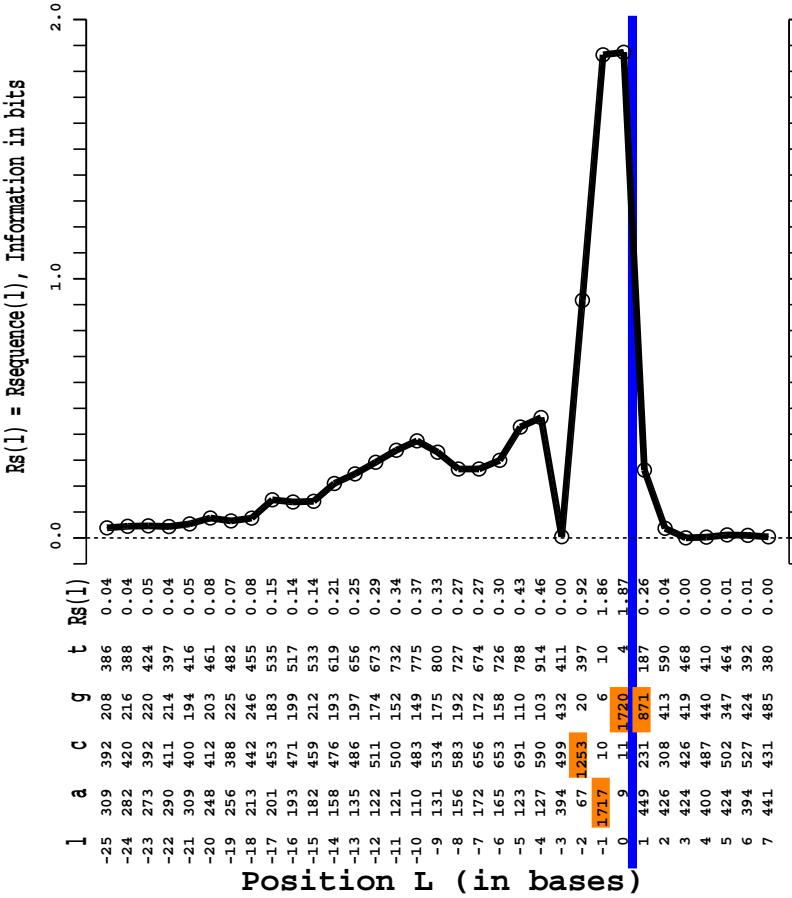
Sequence Conservation → in bits per base

Donor



C A G — G

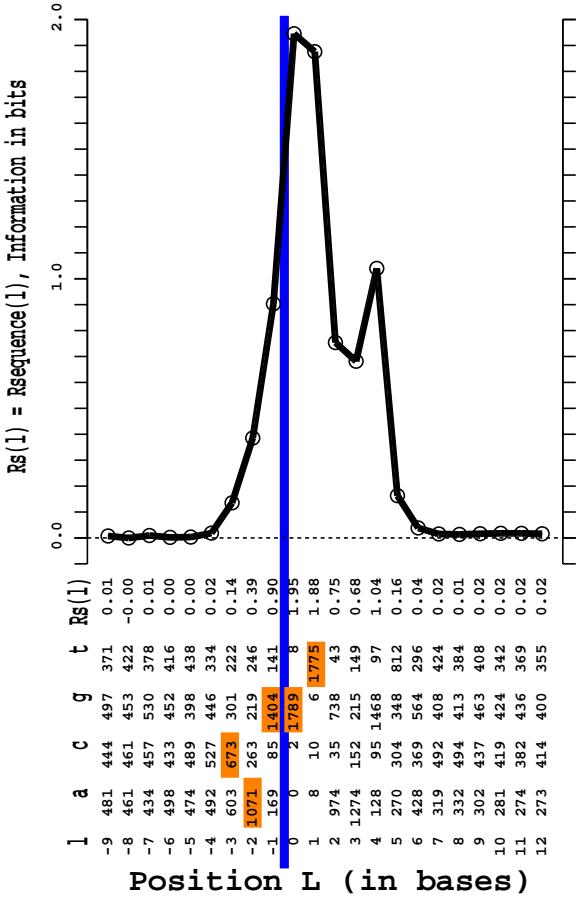
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Acceptor

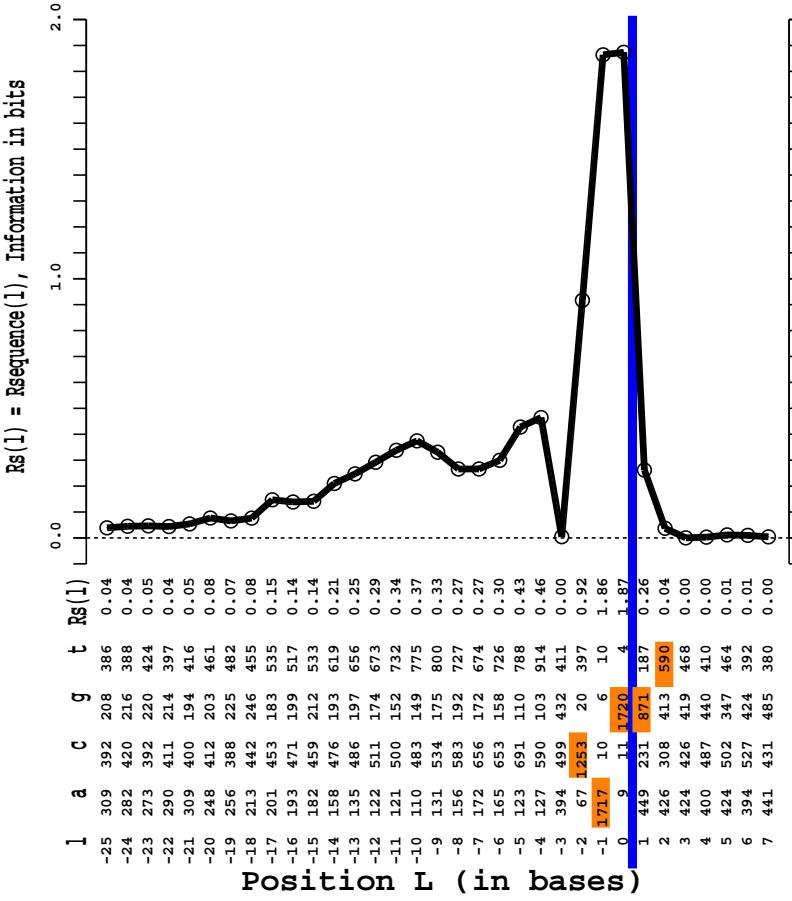
Sequence Conservation → in bits per base

Donor



C A G — G T

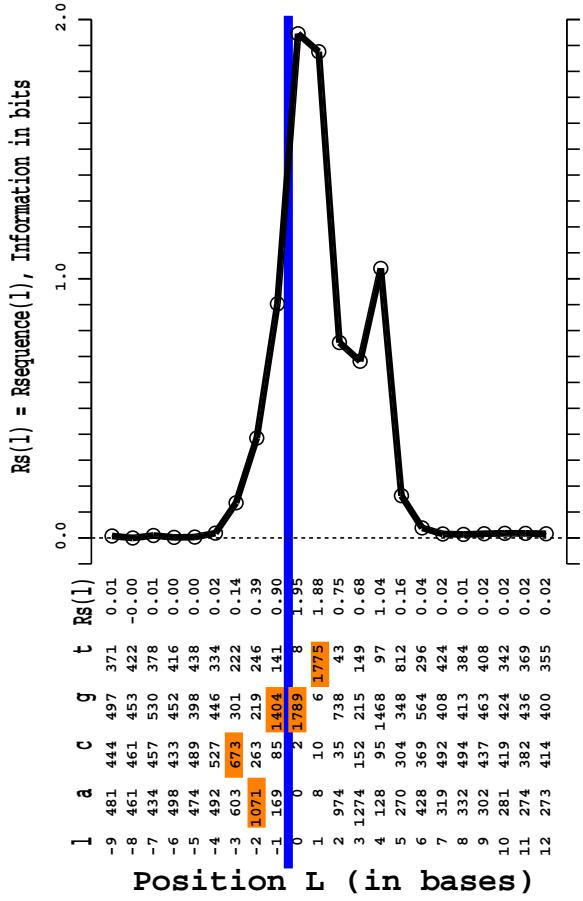
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Acceptor

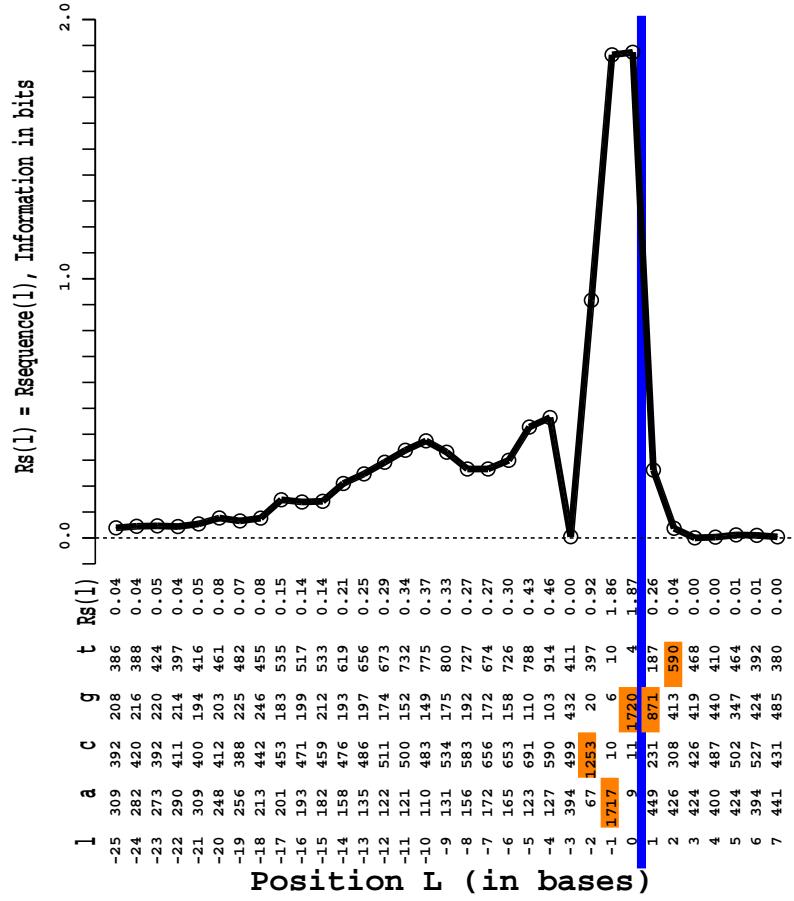
Sequence Conservation → in bits per base

Donor



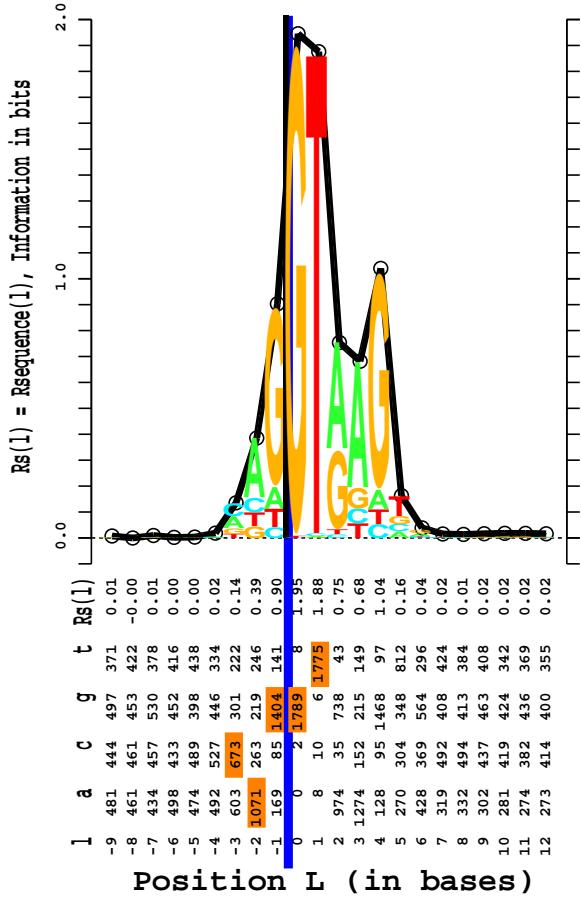
C A G — G T

- The consensus sequences match ...
- BUT the information curves (sequence conservation) differ!

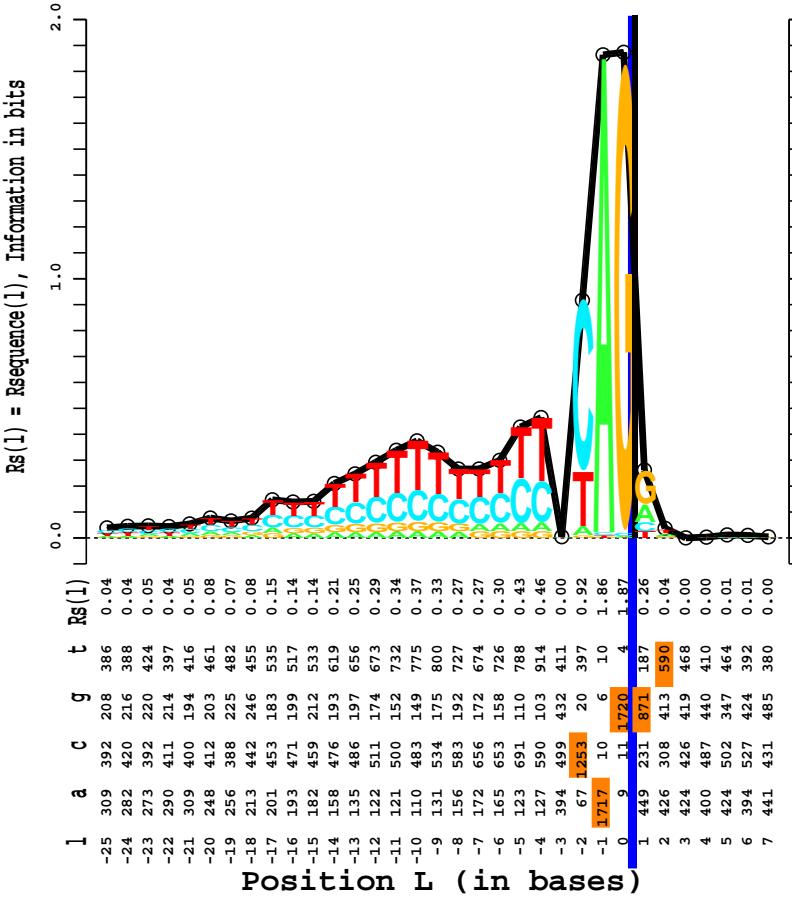


Acceptor

Sequence Conservation → Donor



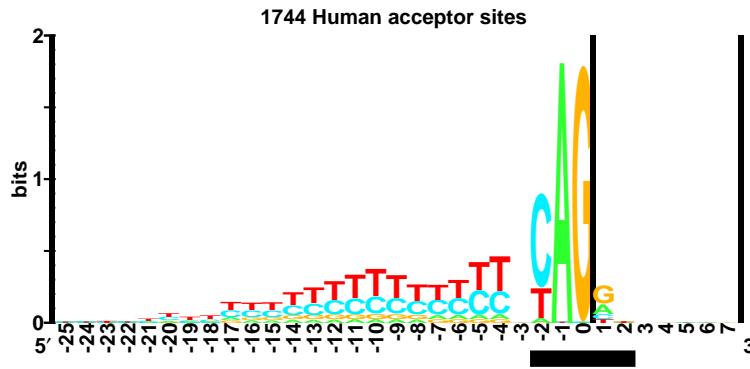
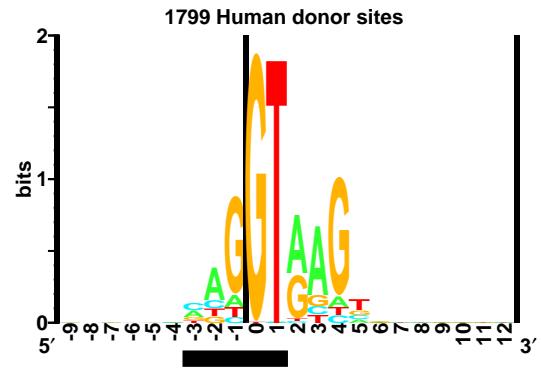
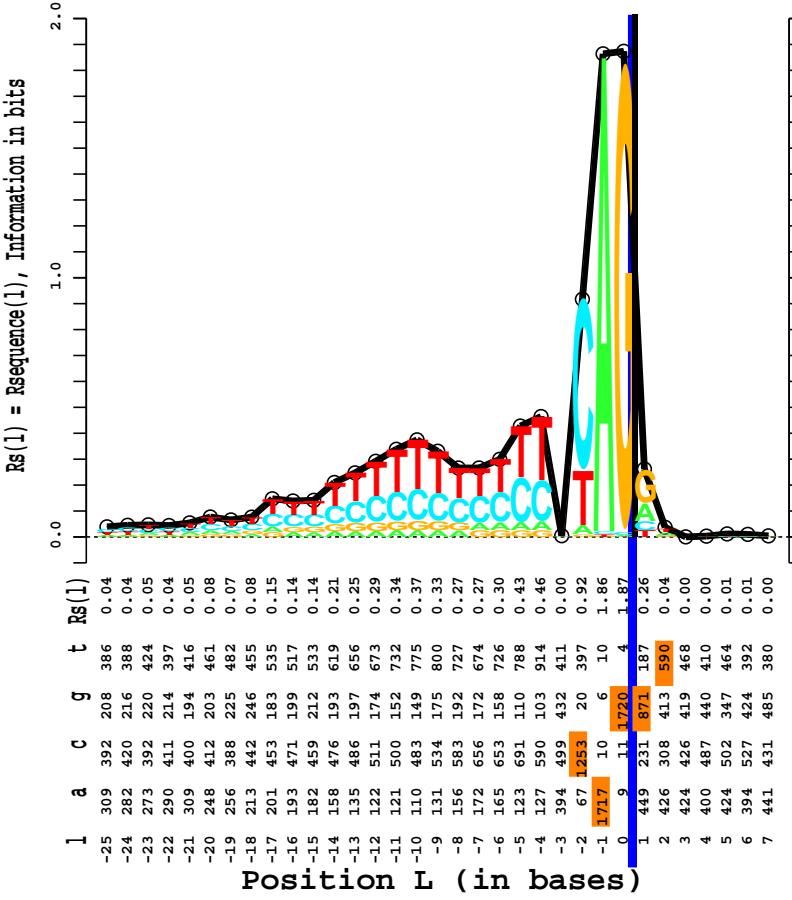
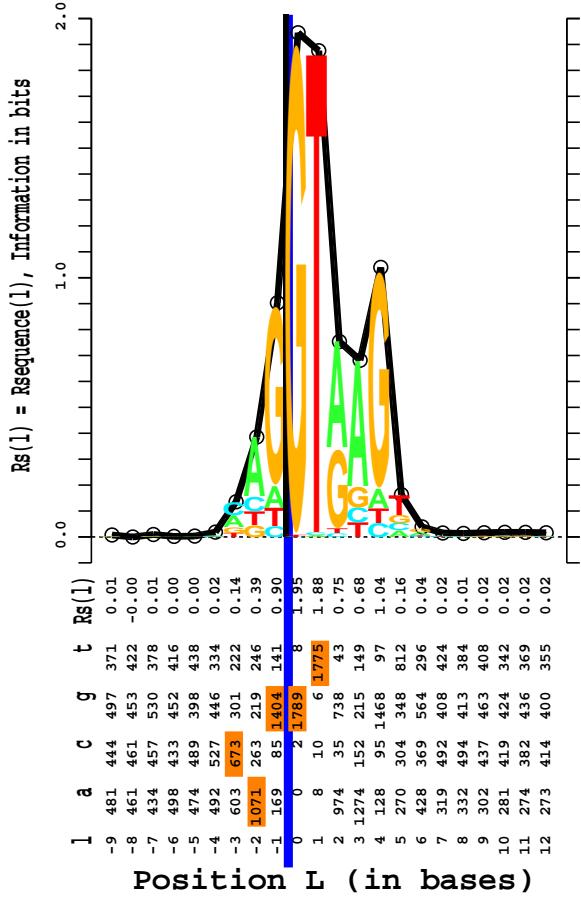
C A G — G T



Accepter

- The consensus sequences match ...
- BUT the information curves (sequence conservation) differ!
- Put letters into the graph proportional to their frequency!

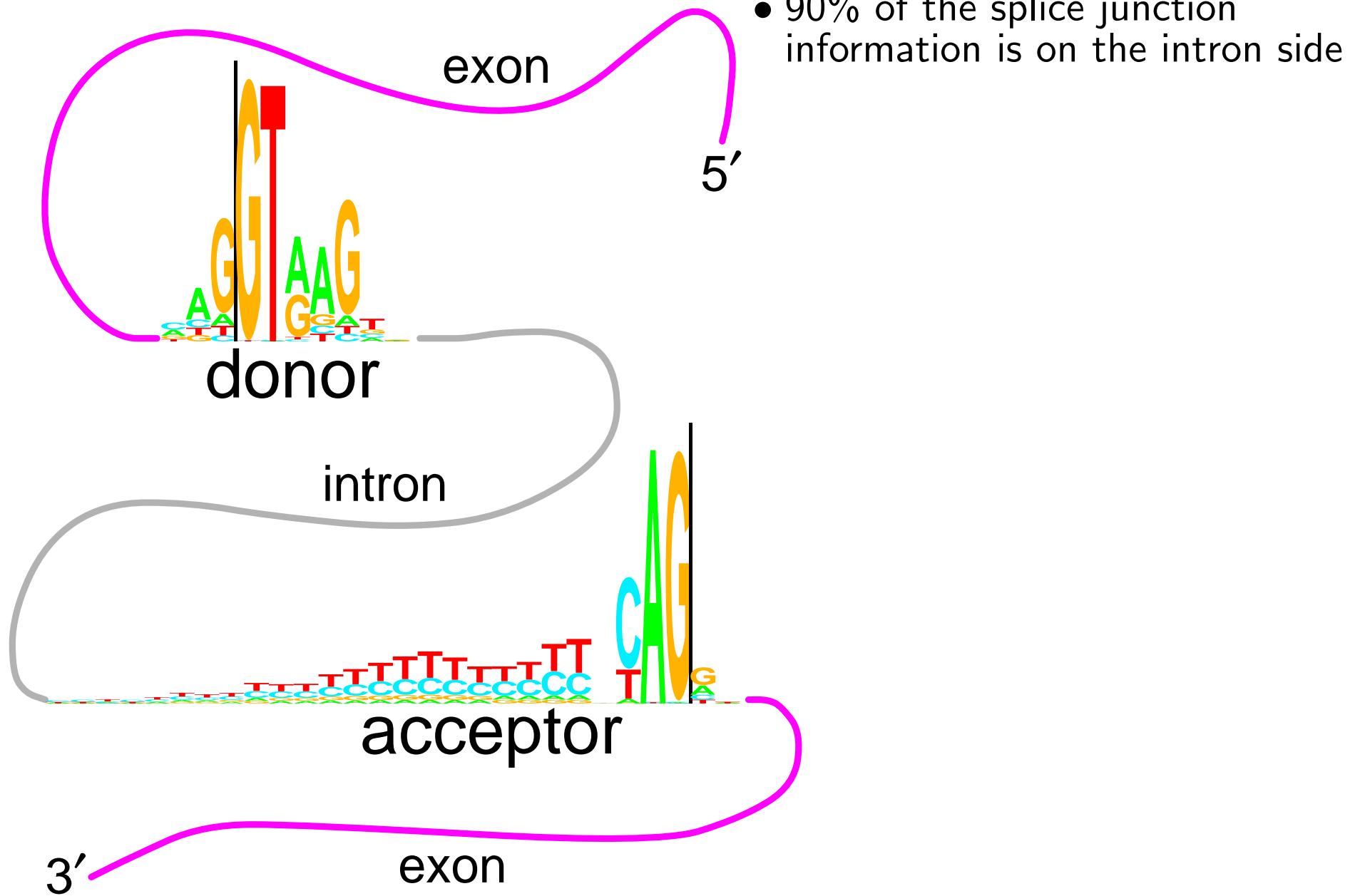
Sequence Conservation →
in bits per base
Donor



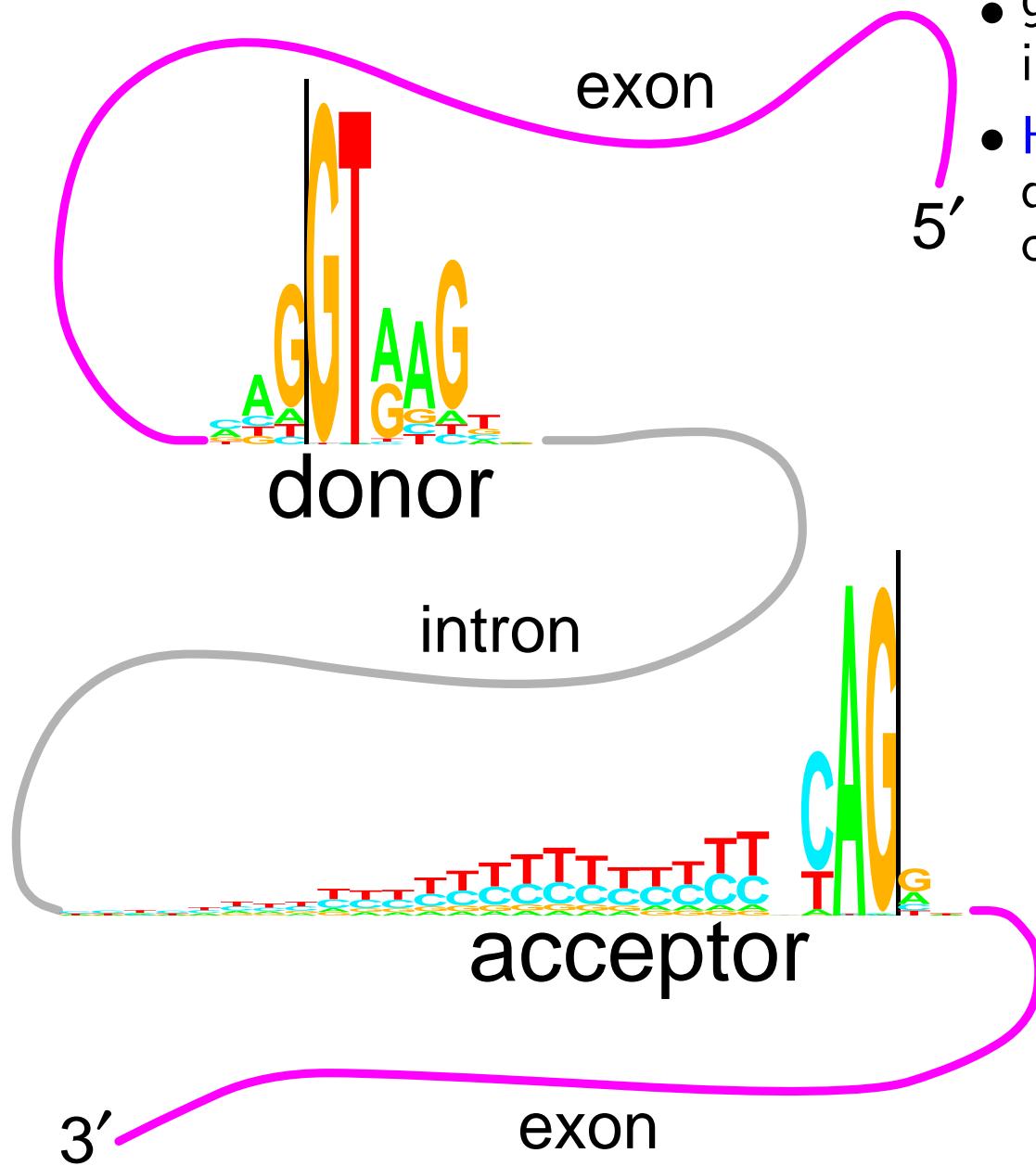
- That's how and why we invented sequence logos!

Acceptor

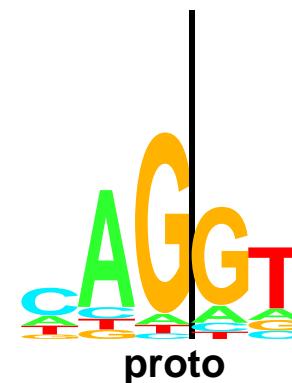
Splice Junction Sequence Logos



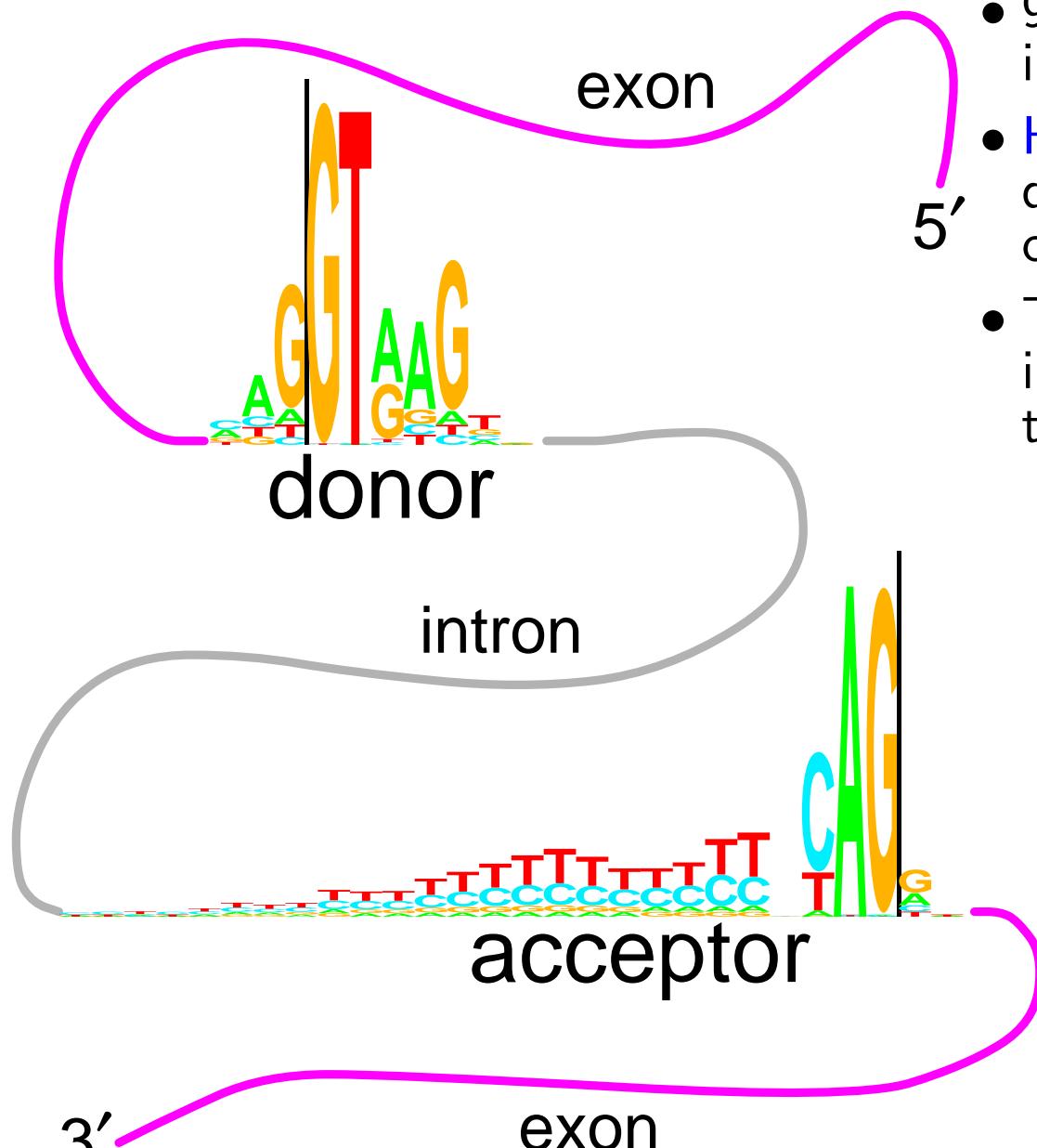
Splice Junction Sequence Logos



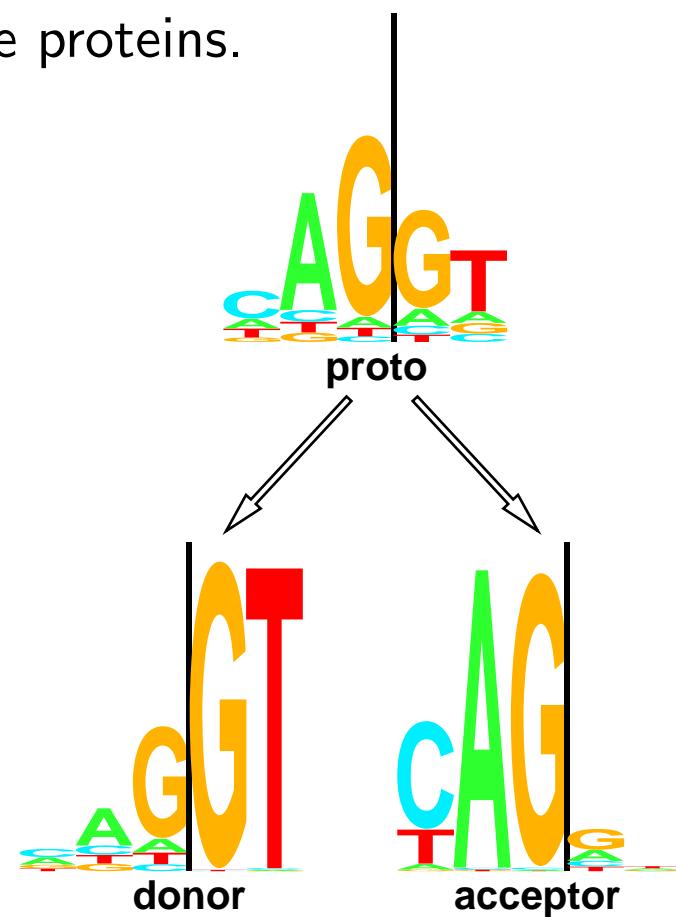
- 90% of the splice junction information is on the intron side
- **Hypothesis:** donor and acceptor sites had a common ancestor that duplicated



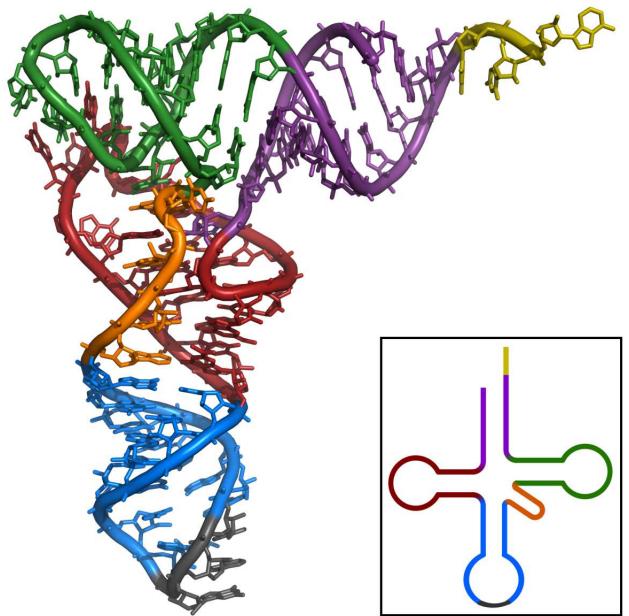
Splice Junction Sequence Logos



- 90% of the splice junction information is on the intron side
- **Hypothesis:** donor and acceptor sites had a common ancestor that duplicated
- They evolved to put the information into the intron. This avoids affecting the proteins.

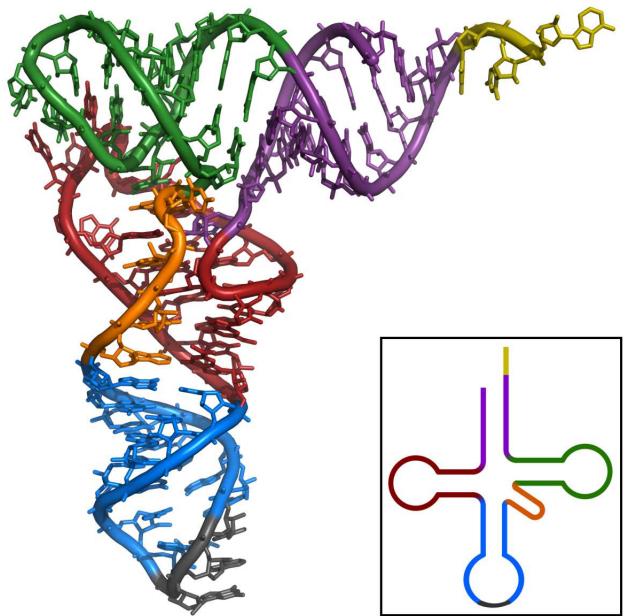


3D Sequence Logos for tRNA Correlations



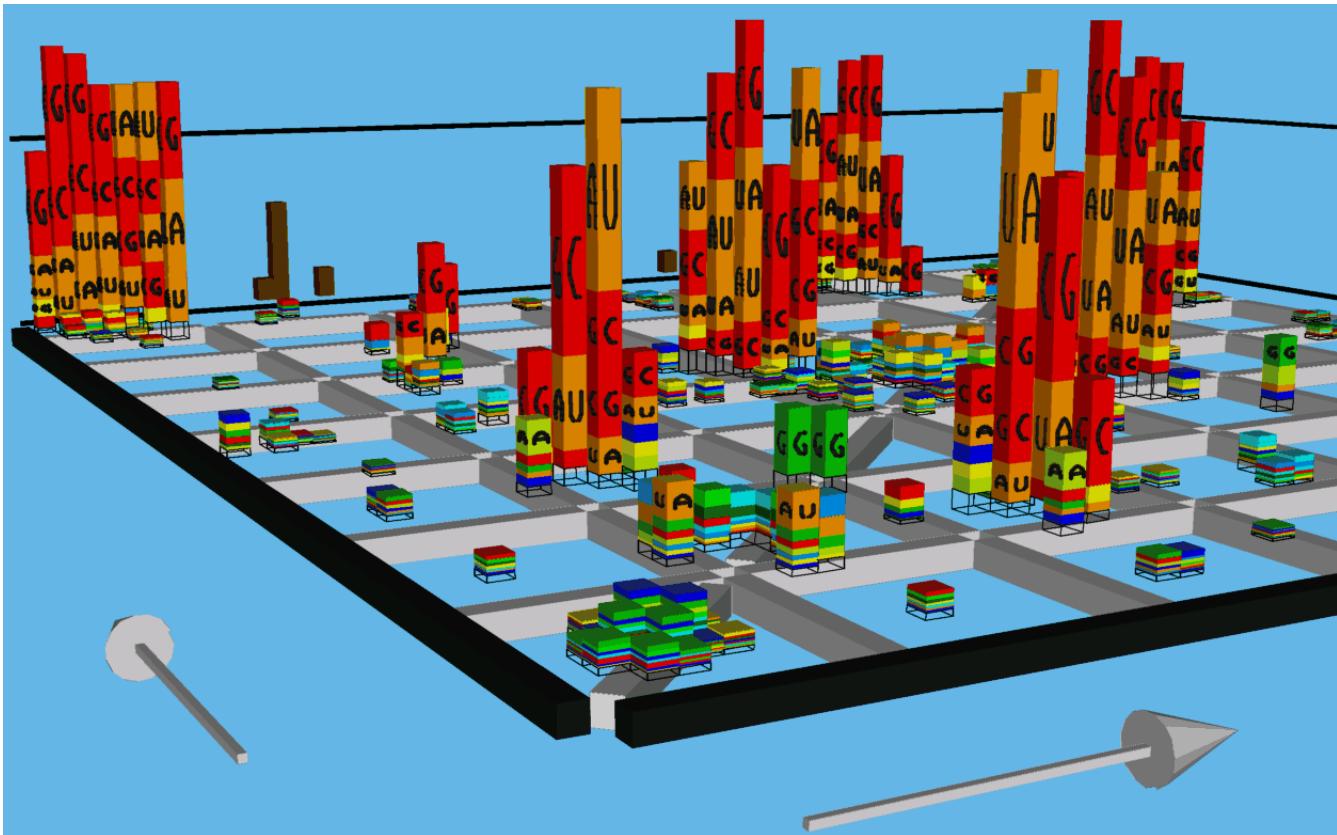
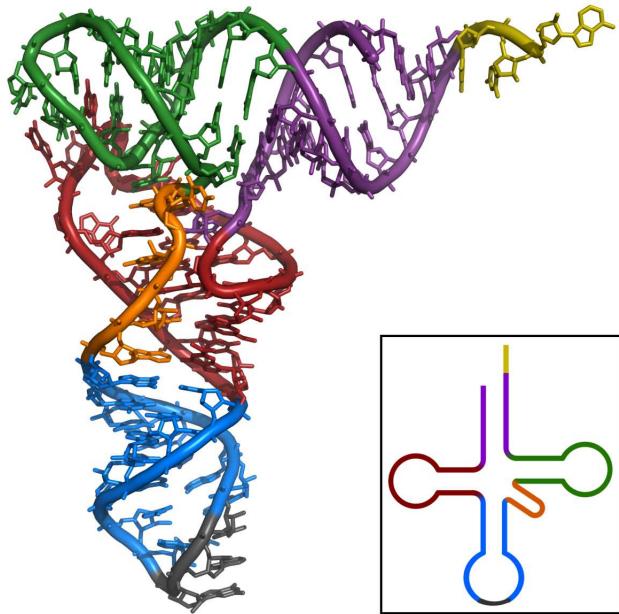
- tRNA reads RNA
to make protein

3D Sequence Logos for tRNA Correlations



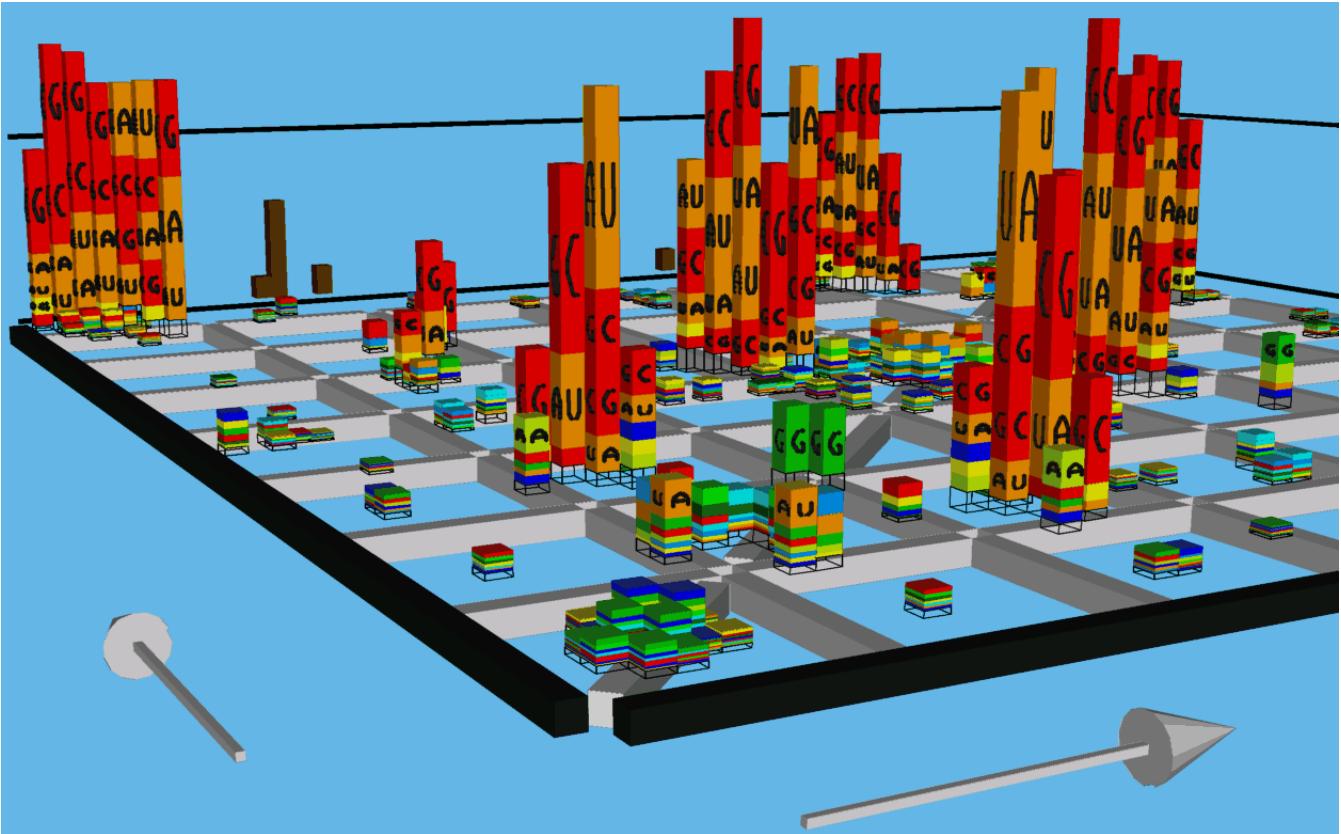
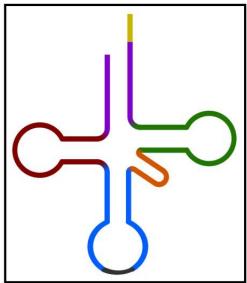
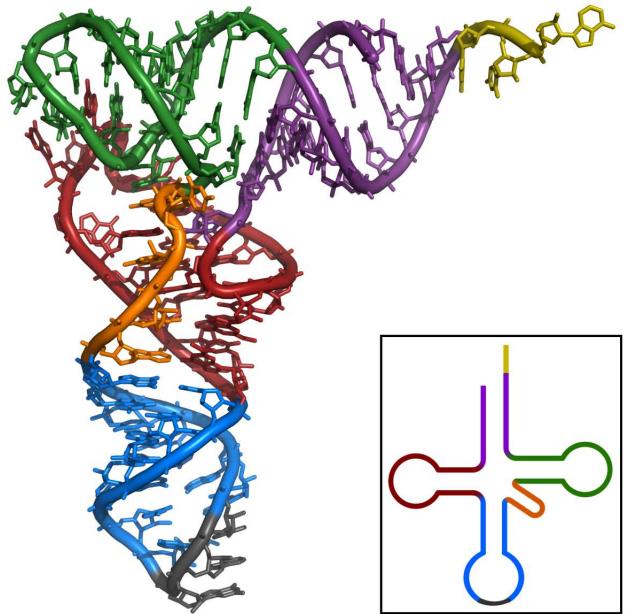
- tRNA reads RNA to make protein
- Correlations can be measured in bits!

3D Sequence Logos for tRNA Correlations



- tRNA reads RNA to make protein
- Correlations can be measured in bits!
- 3D Sequence logo

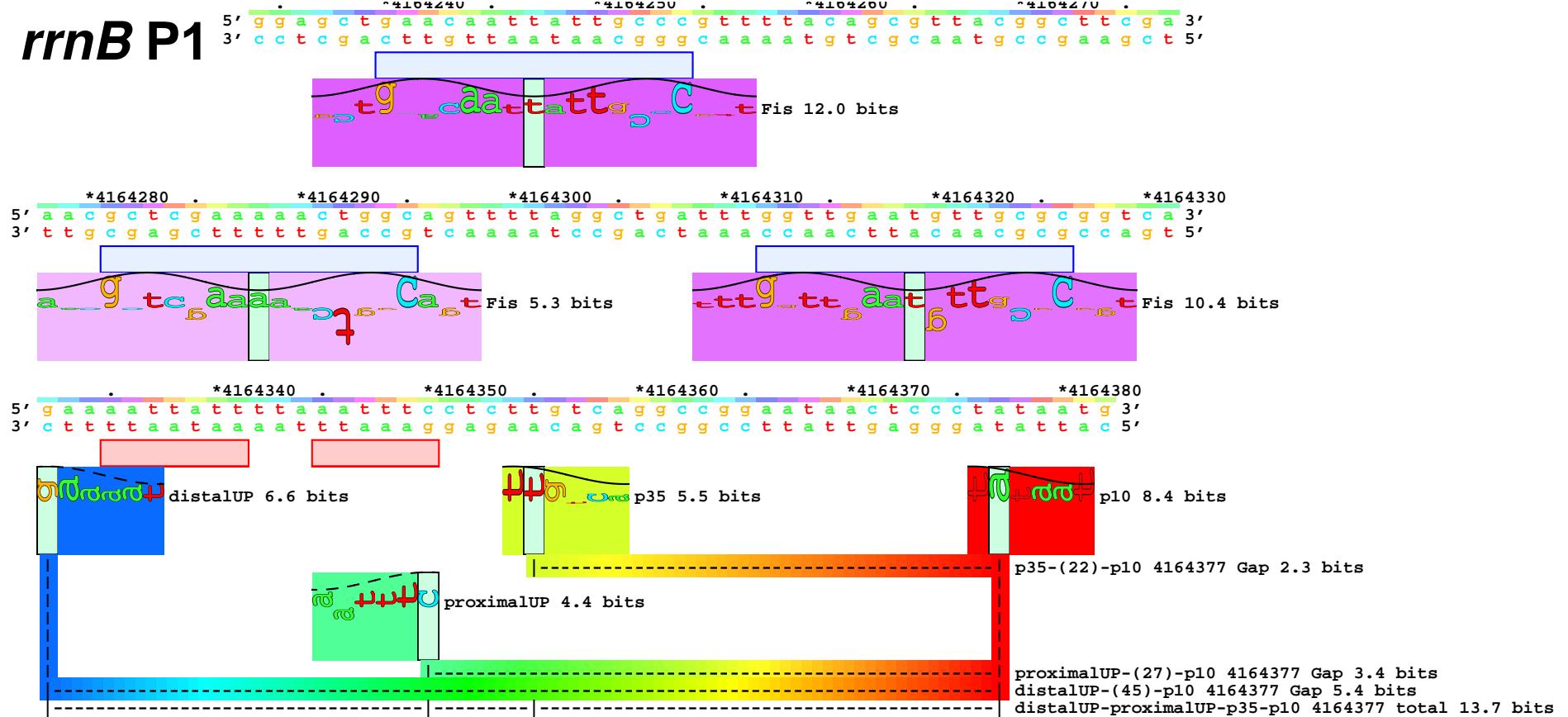
3D Sequence Logos for tRNA Correlations



- tRNA reads RNA to make protein
- Correlations can be measured in bits!
- 3D Sequence logo
- OBSERVED: tRNA stems

Sequence Walker example: rrnB P1

rrnB P1



Complex Sequence Walker Example

- σ^{70} promoters have a -35 and a -10

Complex Sequence Walker Example

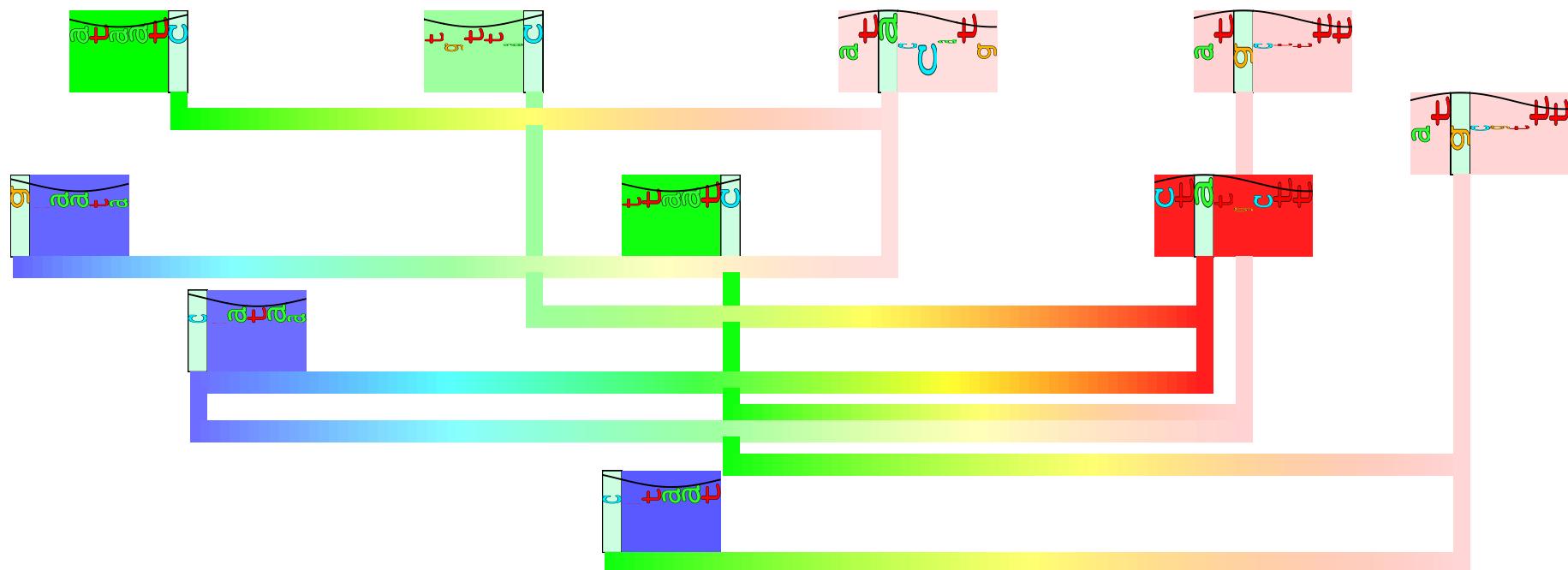
- σ^{70} promoters have a -35 and a -10
- Using information theory we discovered that stress-response σ^{38} promoters do not have a -35

Complex Sequence Walker Example

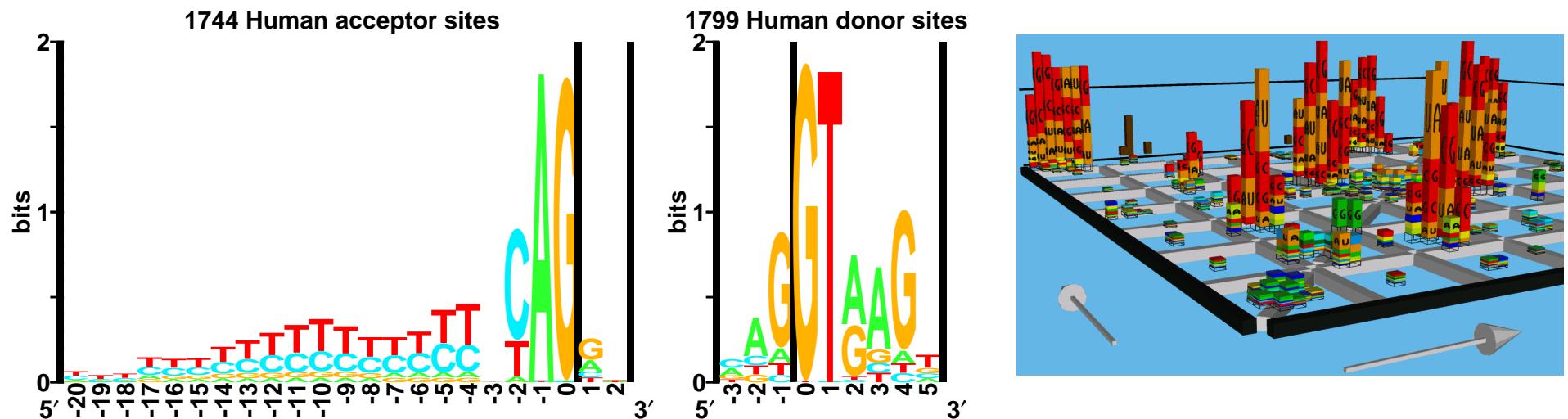
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Complex Sequence Walker Example

- σ^{70} promoters have a -35 and a -10
- Using information theory we discovered that stress-response σ^{38} promoters do not have a -35
- Instead, they have a -10 and two UP elements
- σ^{38} promoter *talA* P1 is complex!

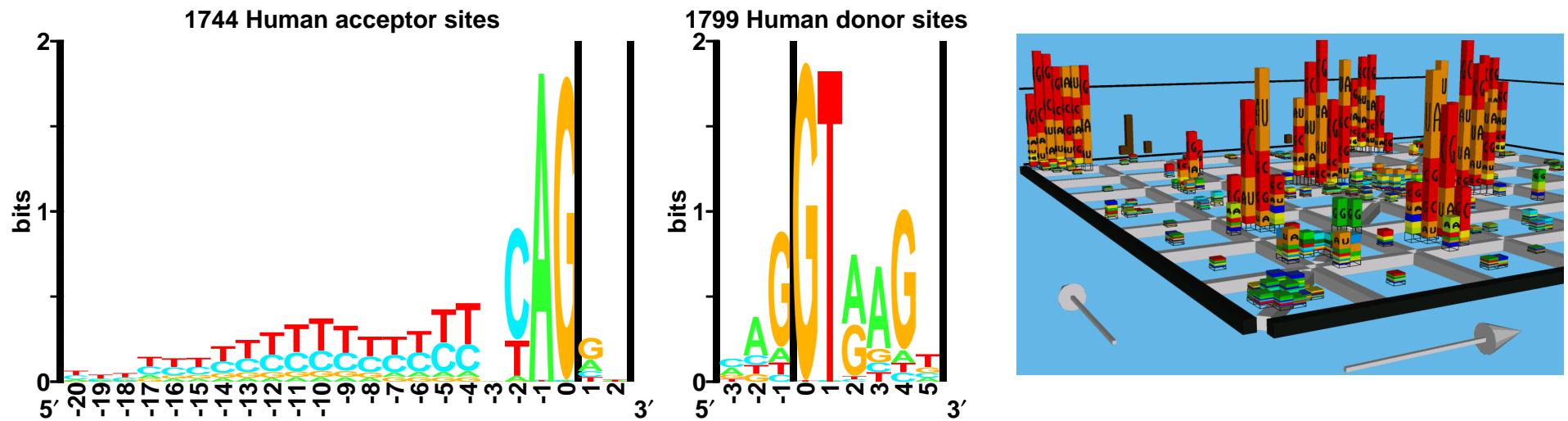


Important Discovery



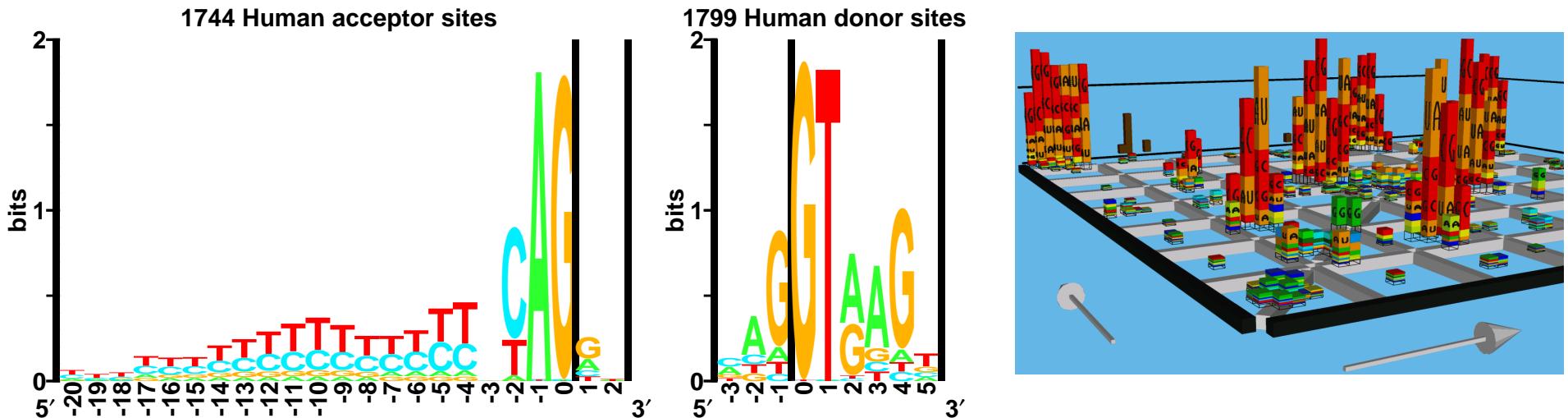
- Area under a sequence logo is the total information.

Important Discovery



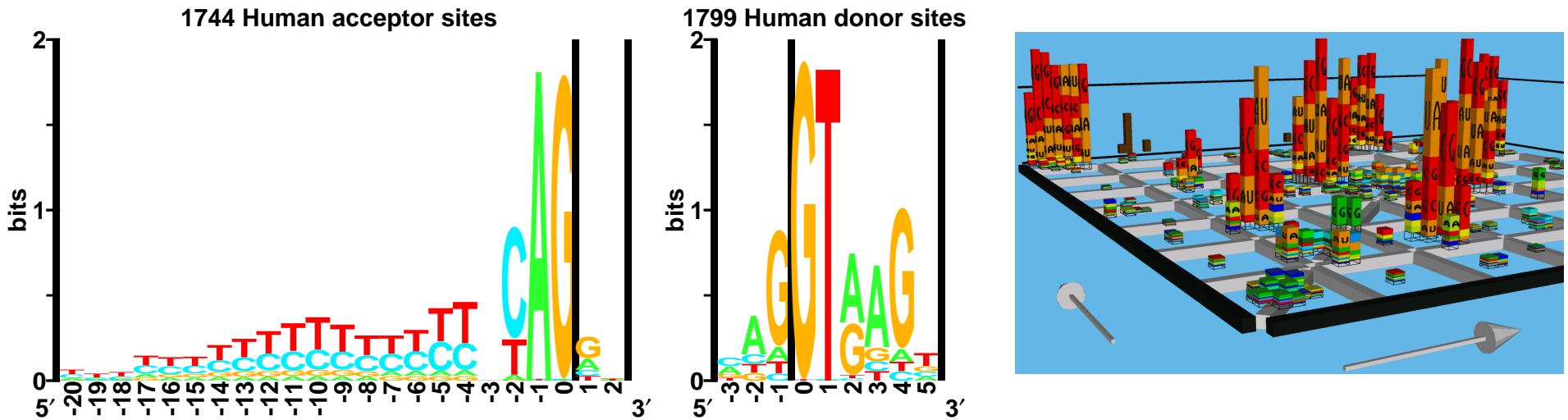
- Area under a sequence logo is the total information.
- How is that related to the binding energy?

Important Discovery



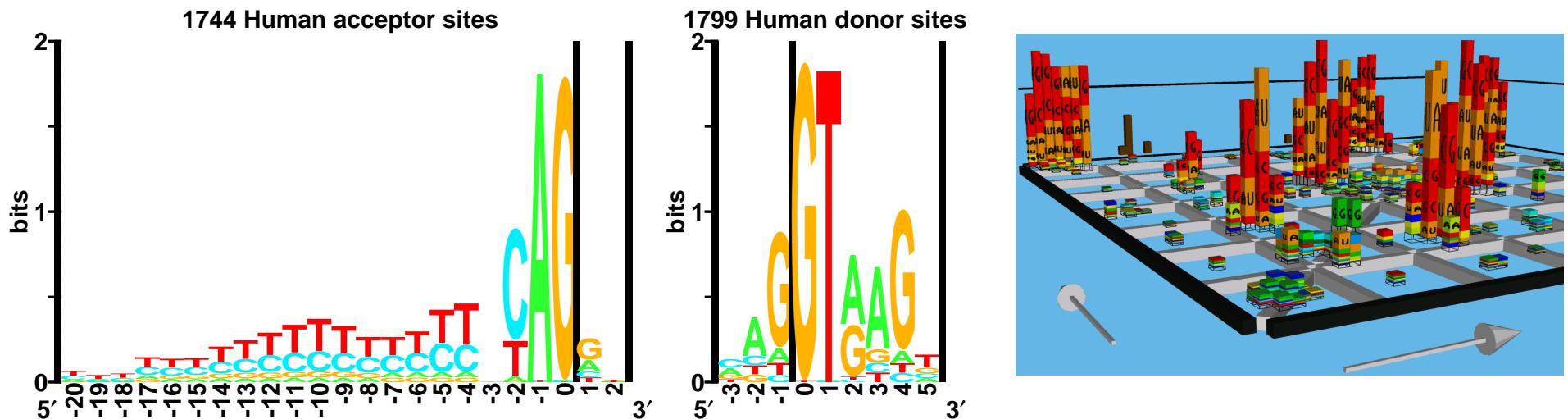
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Important Discovery



- Area under a sequence logo is the total information.
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- Isothermal efficiency

Important Discovery

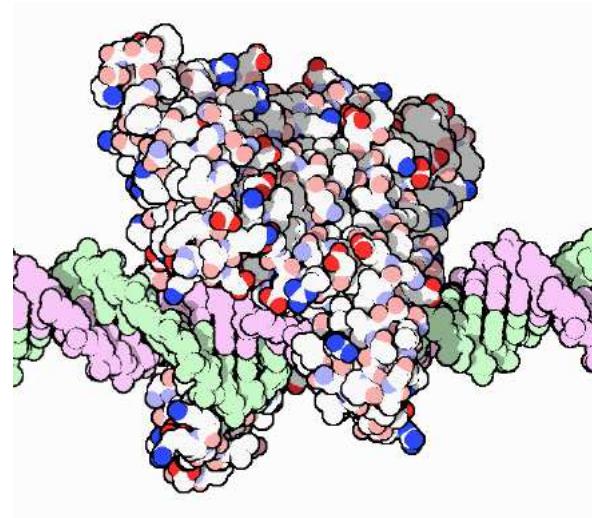


- Area under a sequence logo is the total information.
- How is that related to the binding energy?
- Information gained for energy dissipated
- Isothermal efficiency
- My most important discovery:

Molecules are often 70% efficient

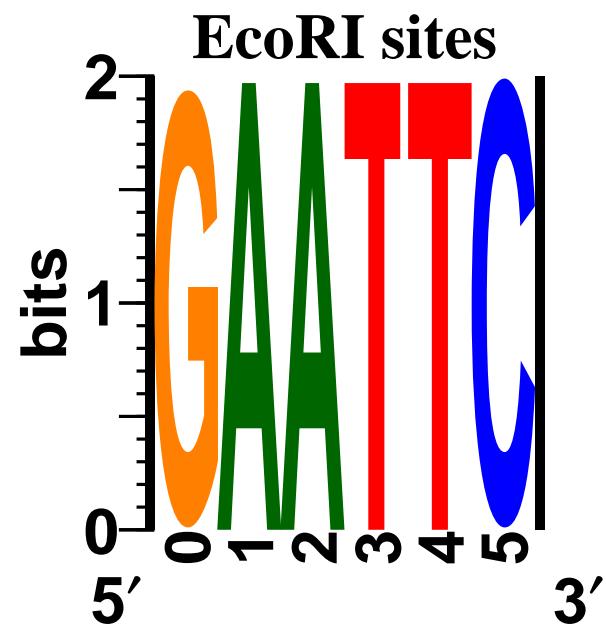
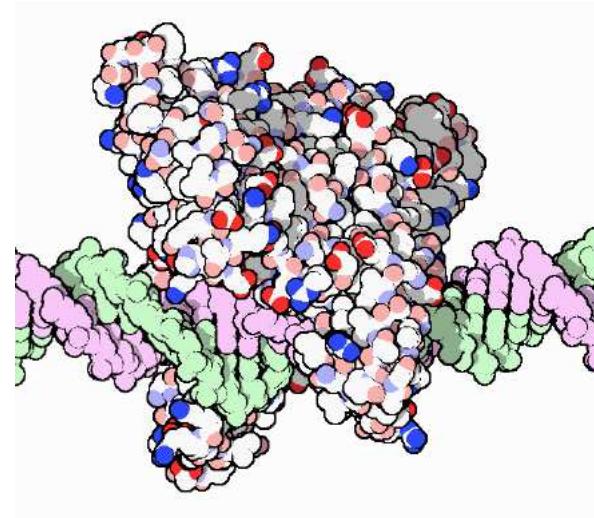
Information of EcoRI DNA Binding

- EcoRI - restriction enzyme



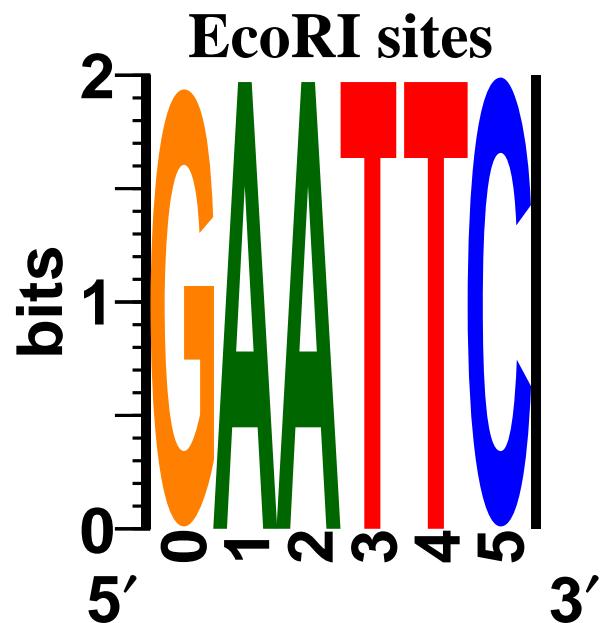
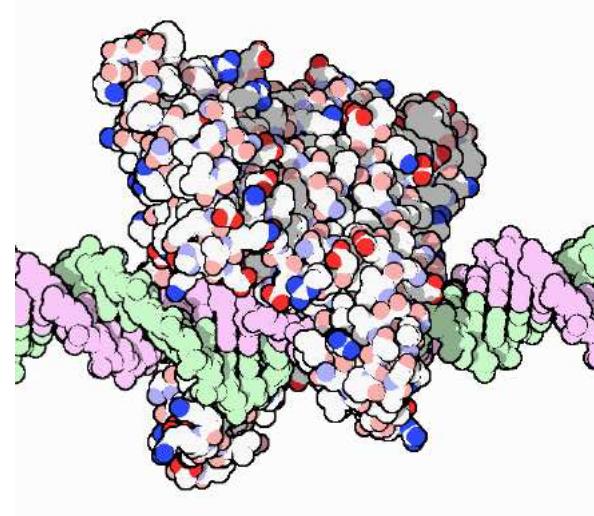
Information of EcoRI DNA Binding

- EcoRI - restriction enzyme
- EcoRI binds DNA at 5' GAATTC 3'



Information of EcoRI DNA Binding

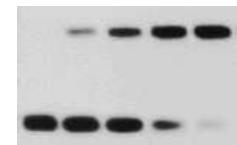
- EcoRI - restriction enzyme
- EcoRI binds DNA at 5' GAATTC 3'
- information required:
 $6 \text{ bases} \times 2 \text{ bits per base} = 12 \text{ bits}$



Energy Dissipation by EcoRI

- Measured specific binding constant:

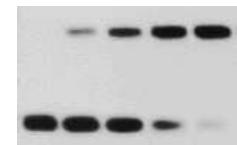
$$K_{spec} = 1.6 \times 10^5$$



Energy Dissipation by EcoRI

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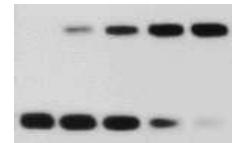
- Average energy dissipated by one molecule as it binds:

$$\Delta G_{spec}^\circ = -k_B T \ln K_{spec} \quad (\text{joules per binding})$$

Energy Dissipation by EcoRI

- Measured specific binding constant:

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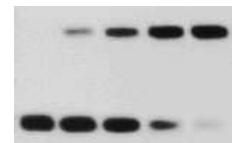
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- Number of bits that could have been selected:

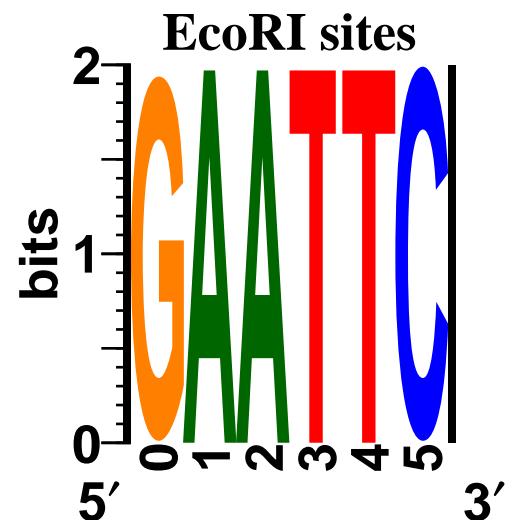
$$\begin{aligned} R_{energy} &= -\Delta G^\circ / \mathcal{E}_{min} \\ &= k_B T \ln K_{spec} / k_B T \ln 2 \\ &= \log_2 K_{spec} \qquad \qquad \Leftarrow \text{SO SIMPLE!} \\ &= \boxed{17.3 \text{ bits per binding}} \end{aligned}$$

Information/Energy = Efficiency of EcoRI

EcoRI could have made 17.3 binary choices

Information/Energy = Efficiency of EcoRI

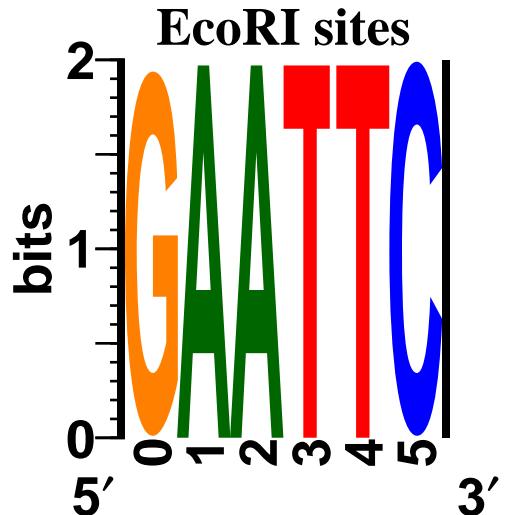
EcoRI could have made 17.3 binary choices
... but it only made 12 choices.



Information/Energy = Efficiency of EcoRI

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Efficiency is
'WORK' DONE / ENERGY DISSIPATED

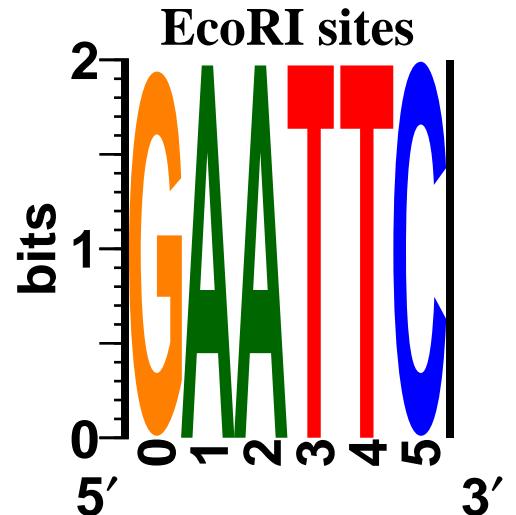


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$$\frac{12 \text{ bits per binding}}{17.3 \text{ bits per binding}} = 0.7$$

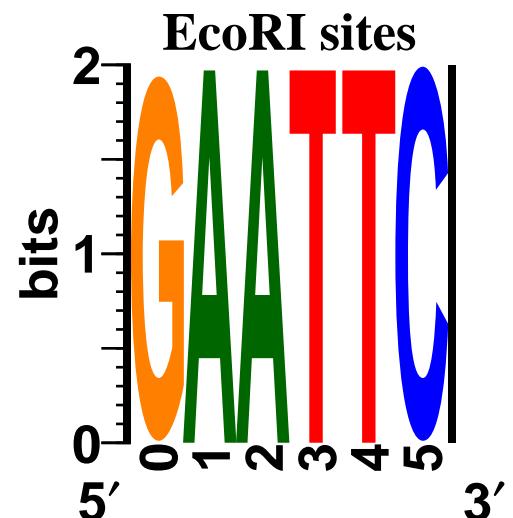


Information/Energy = Efficiency of EcoRI = 70%

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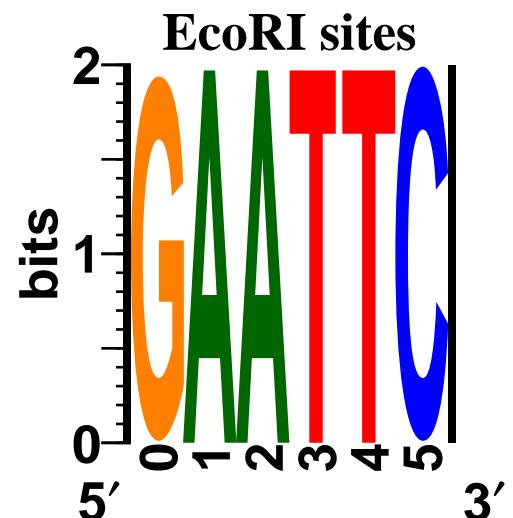
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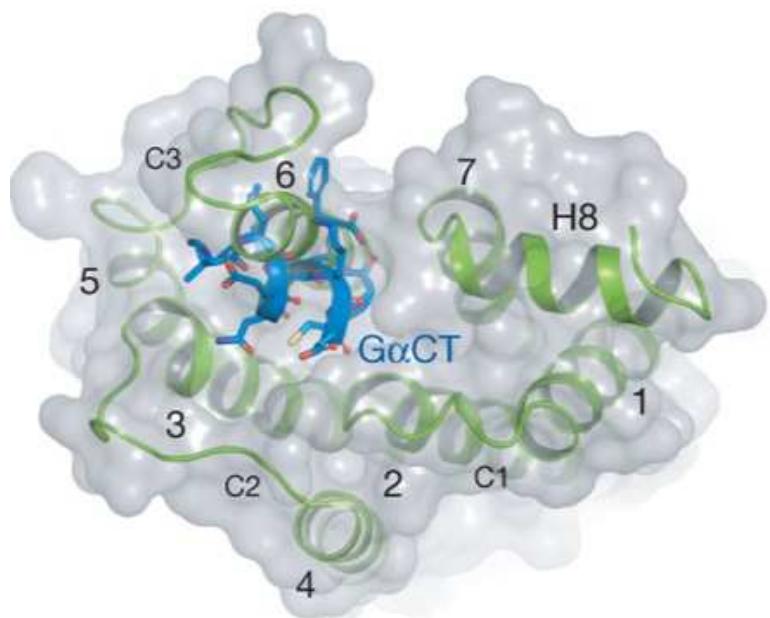


The efficiency is 70%.

18 out of 19 DNA binding proteins give ~70% efficiency.

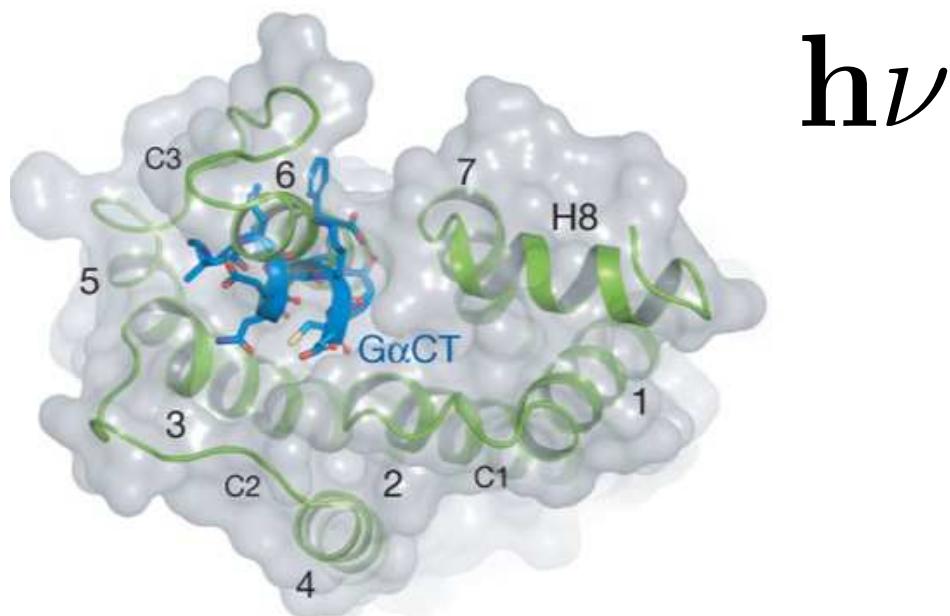
Rhodopsin Shape Change

Dark State



Rhodopsin Shape Change

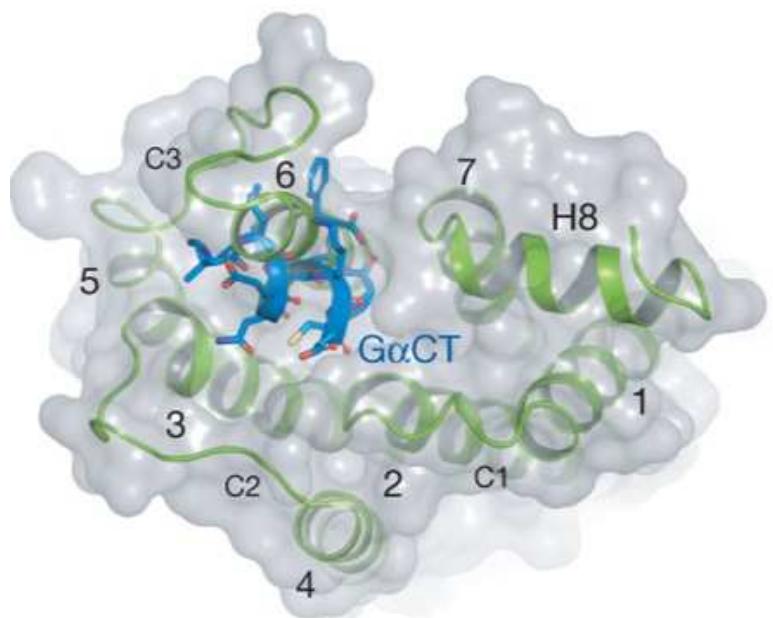
Dark State



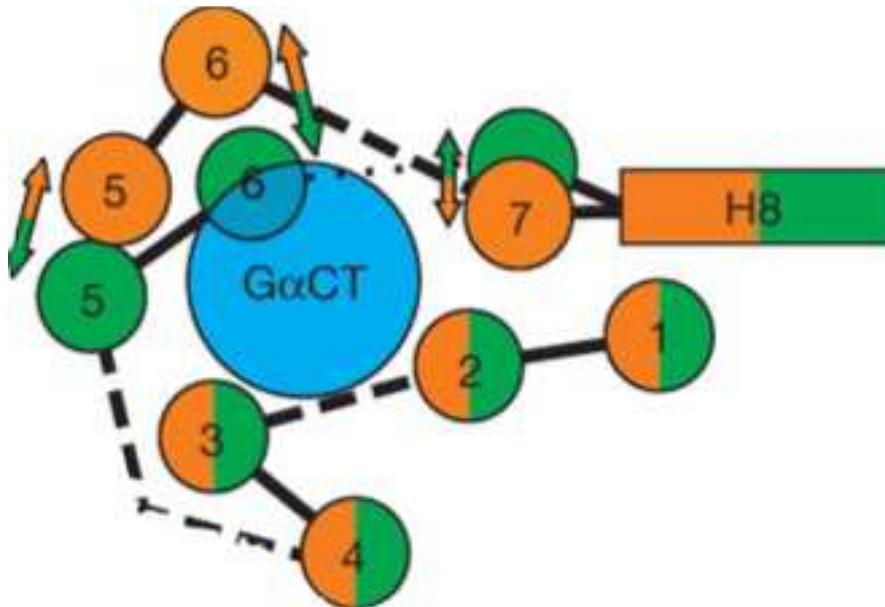
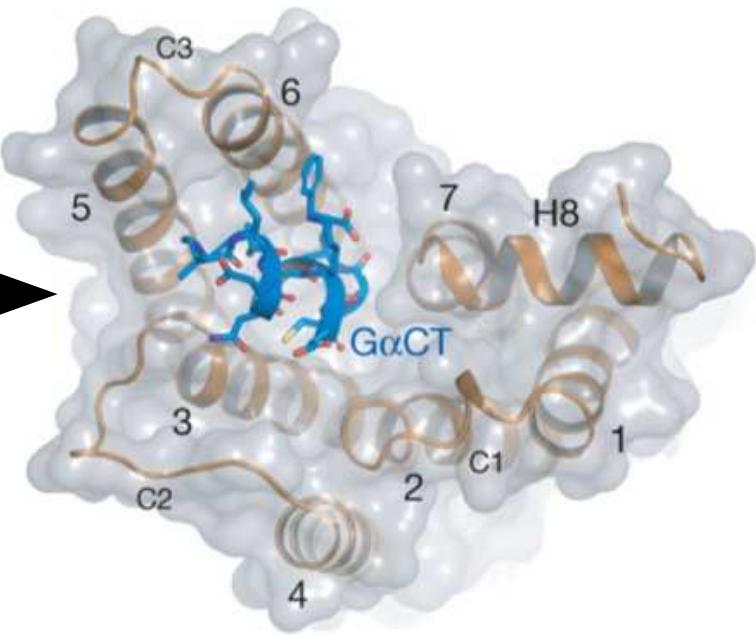
$h\nu$

Rhodopsin Shape Change

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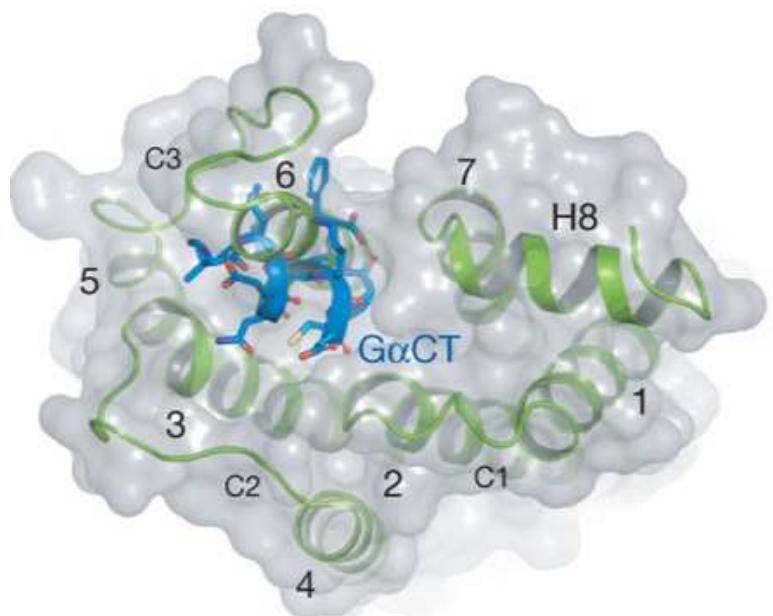


After Photon - Light State

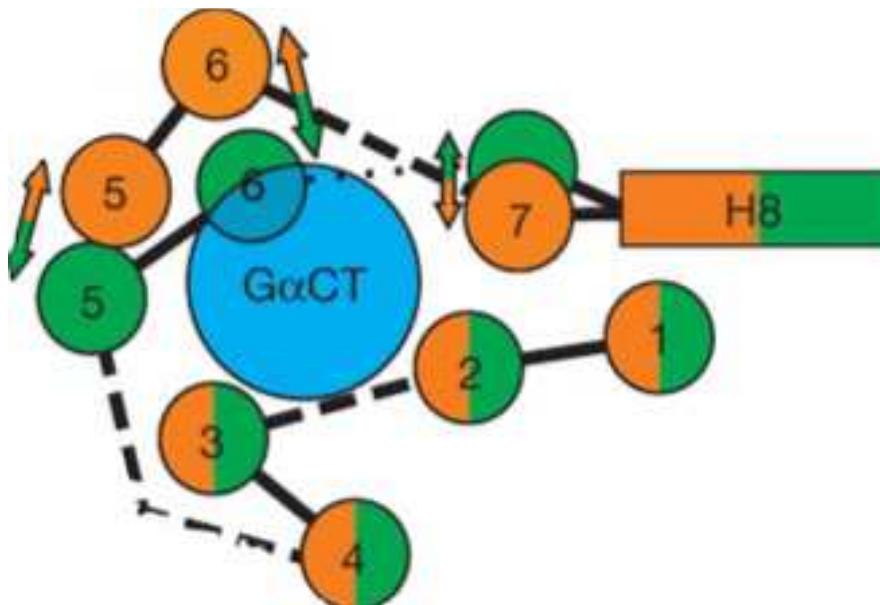
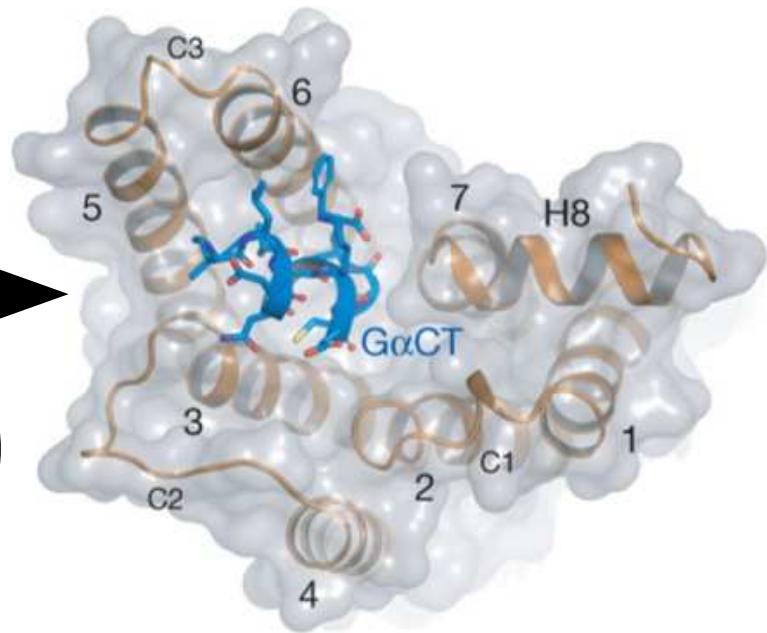


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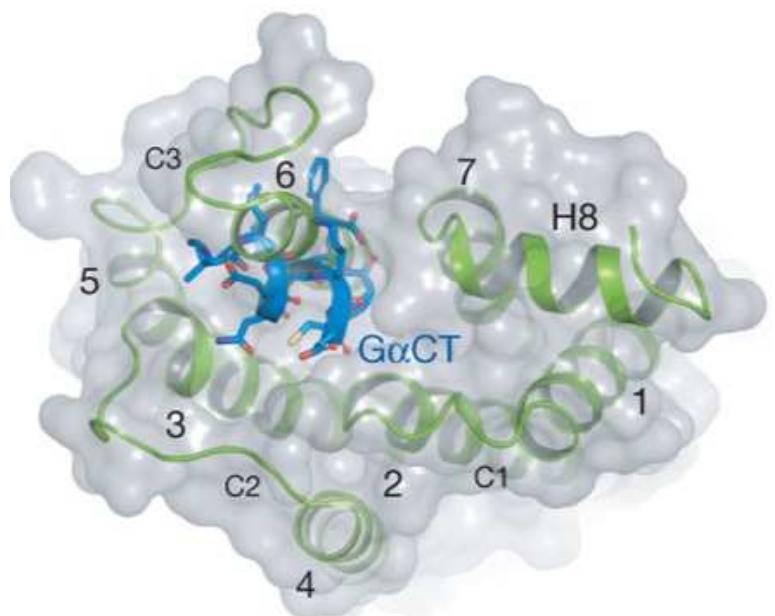


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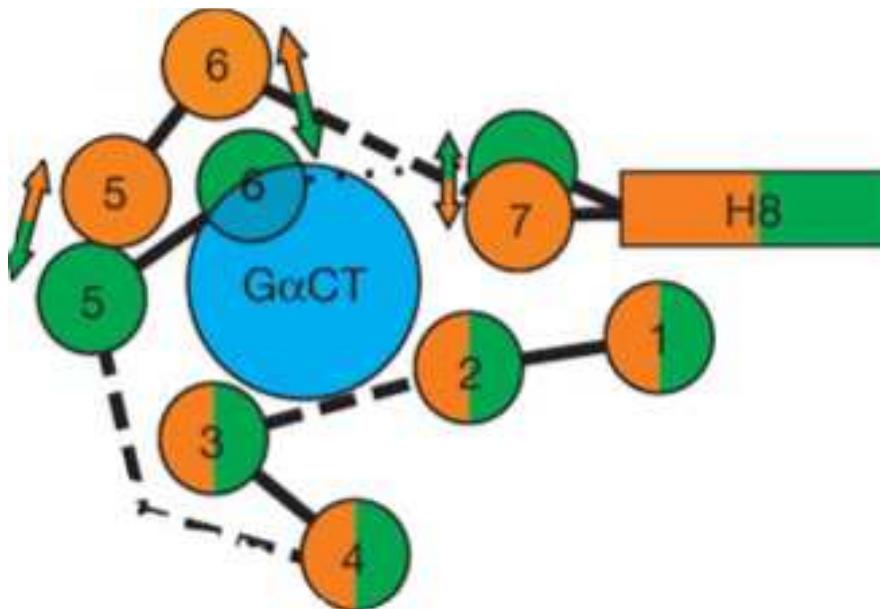
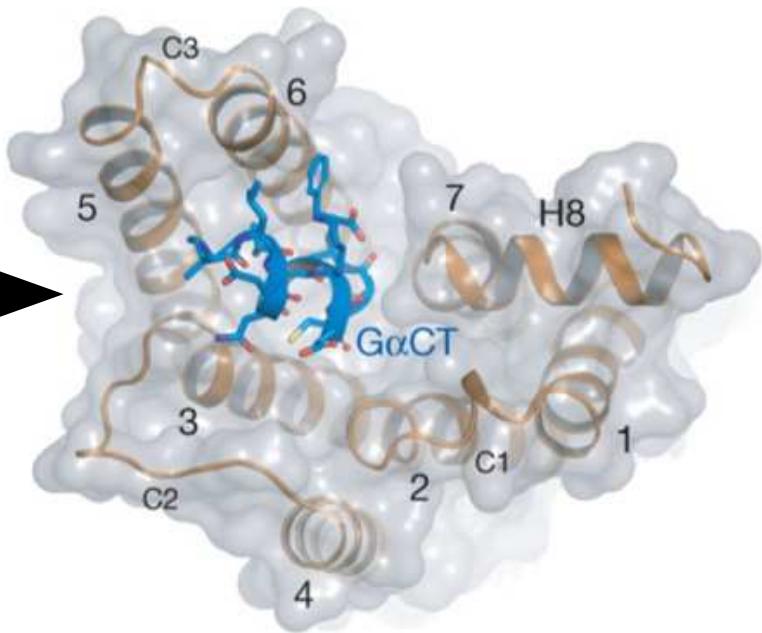


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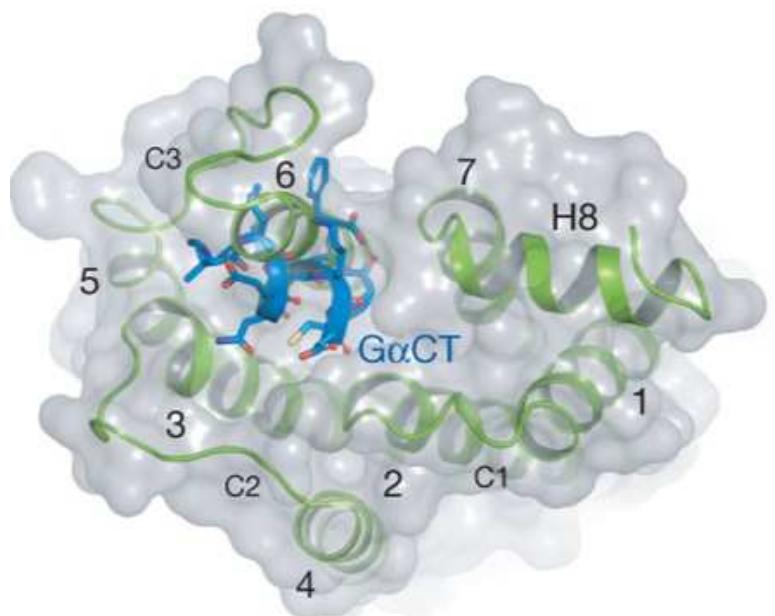


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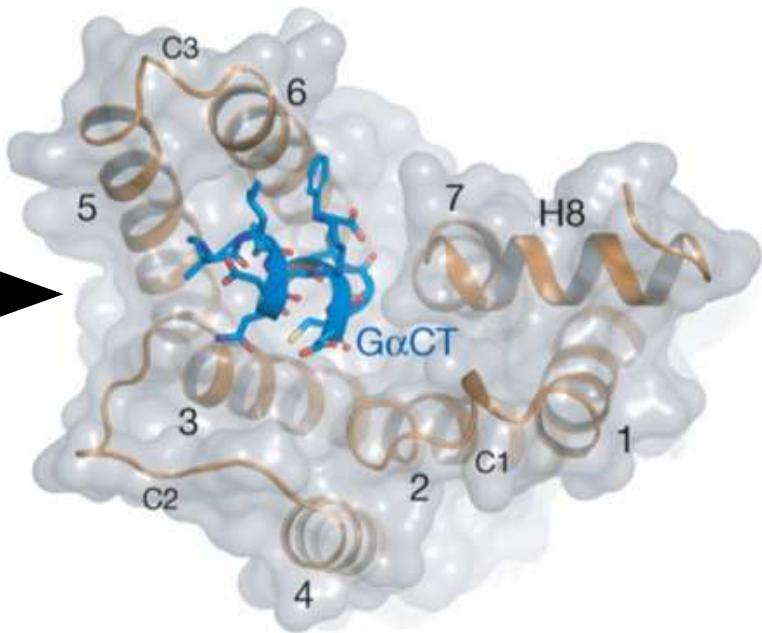


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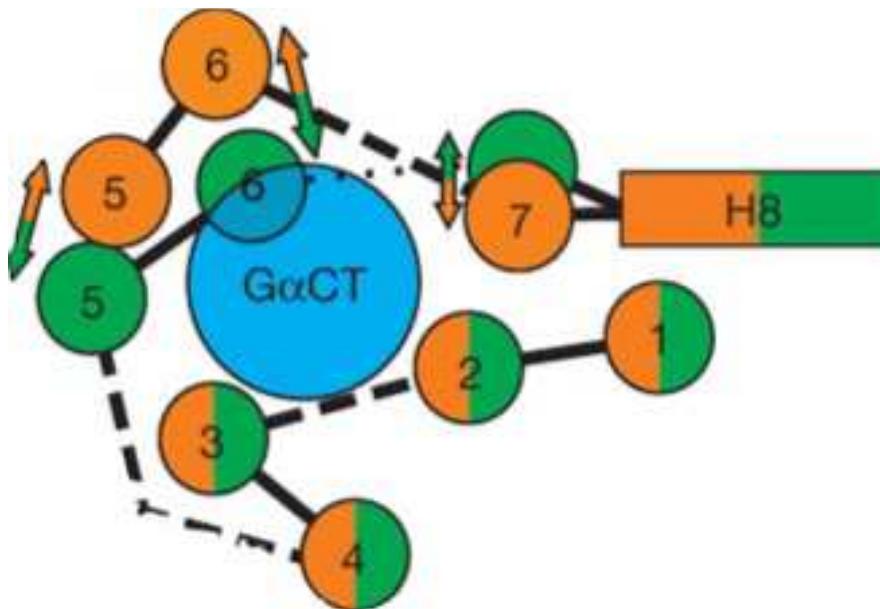
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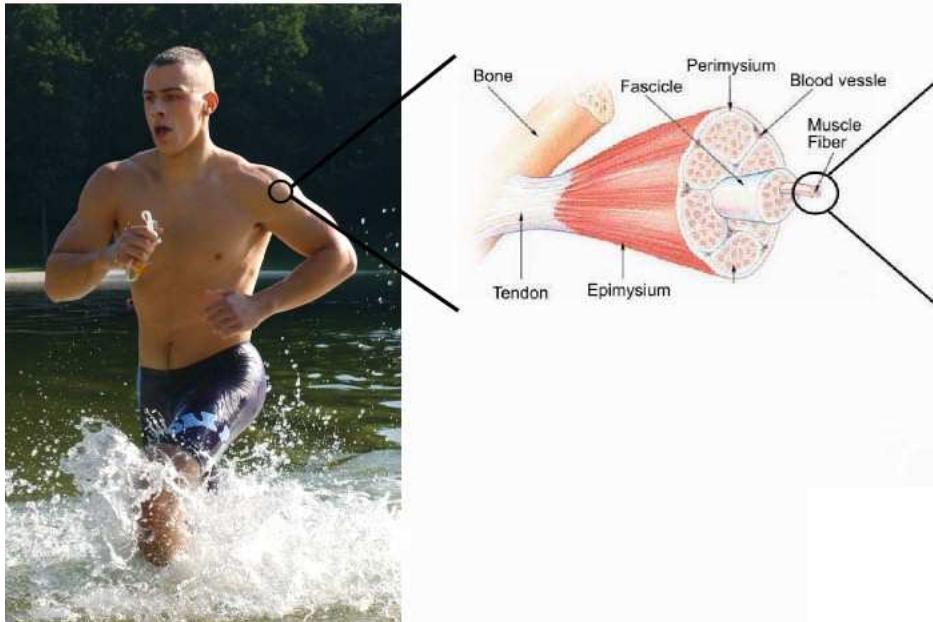
$h\nu$
70%
30%



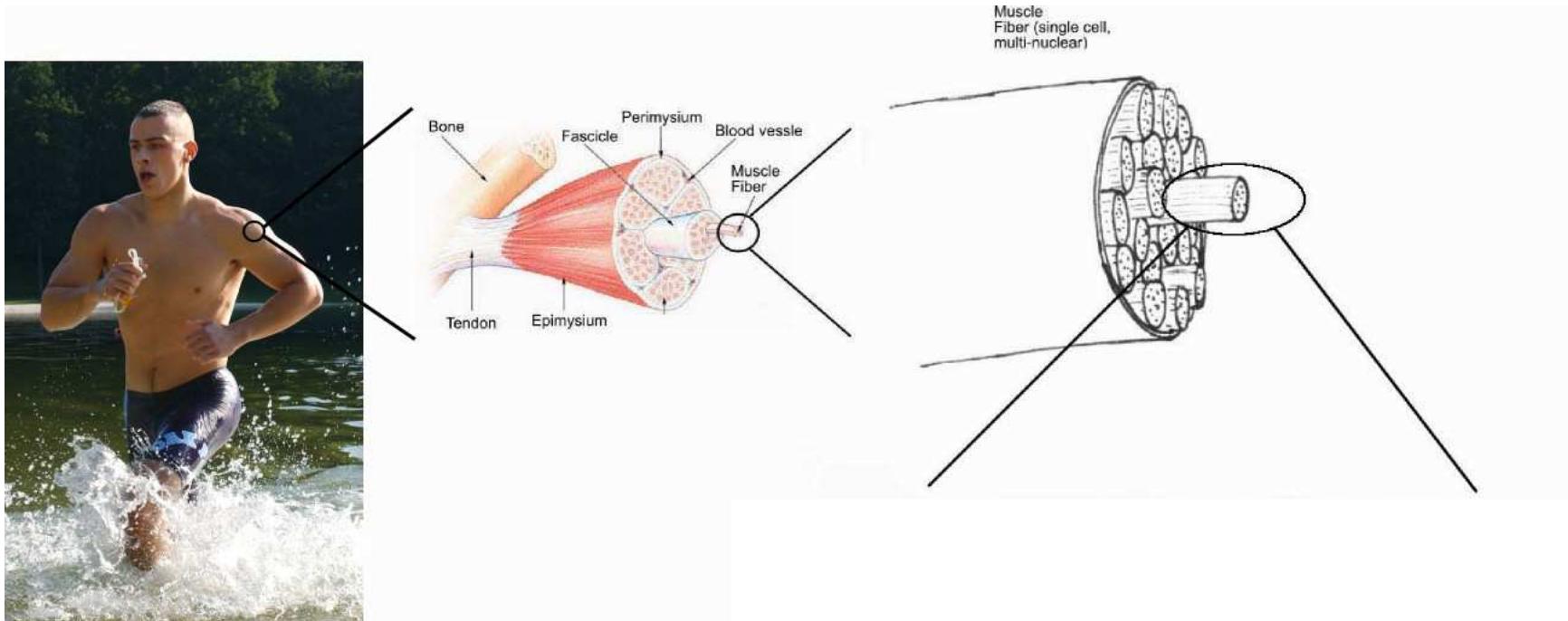
Muscle Structure



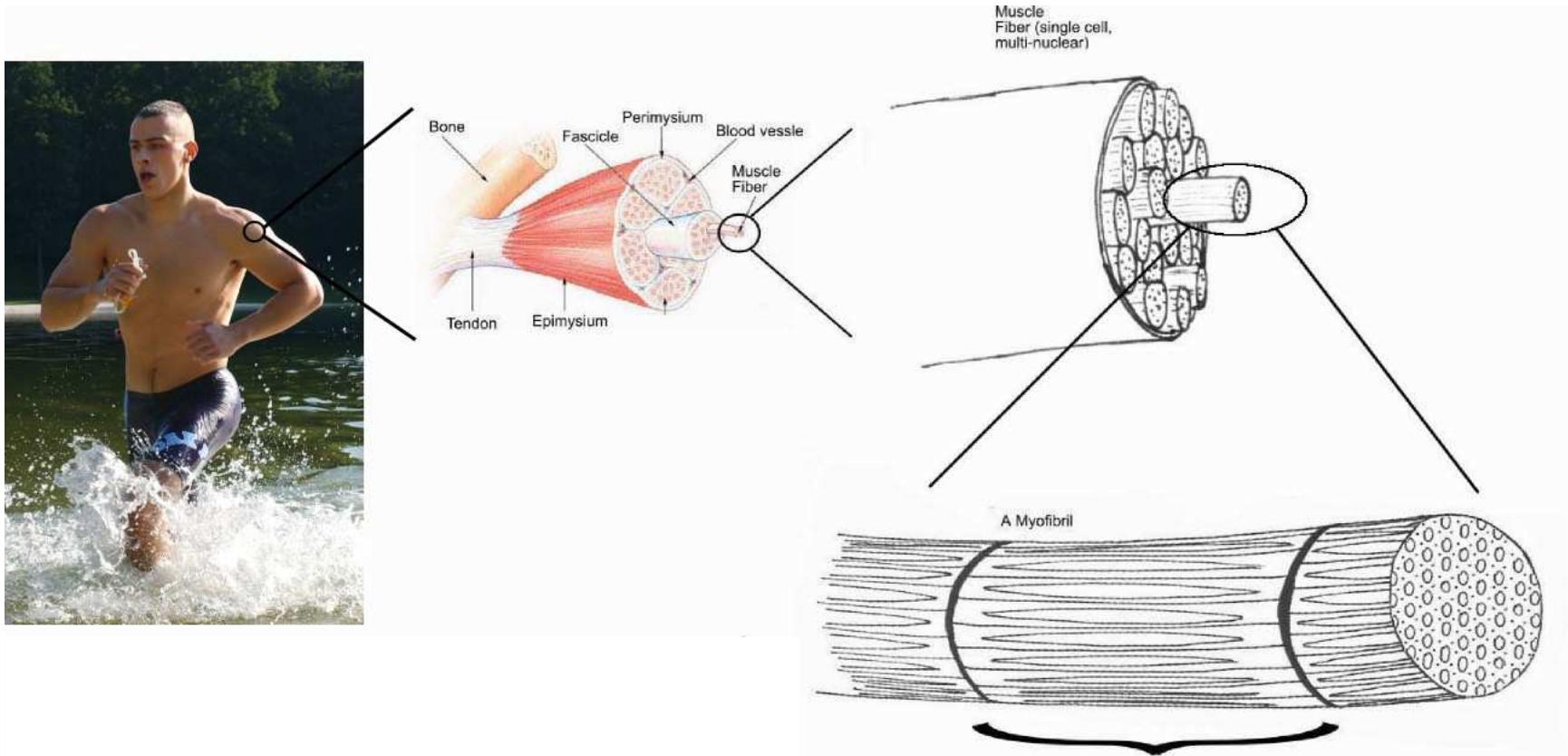
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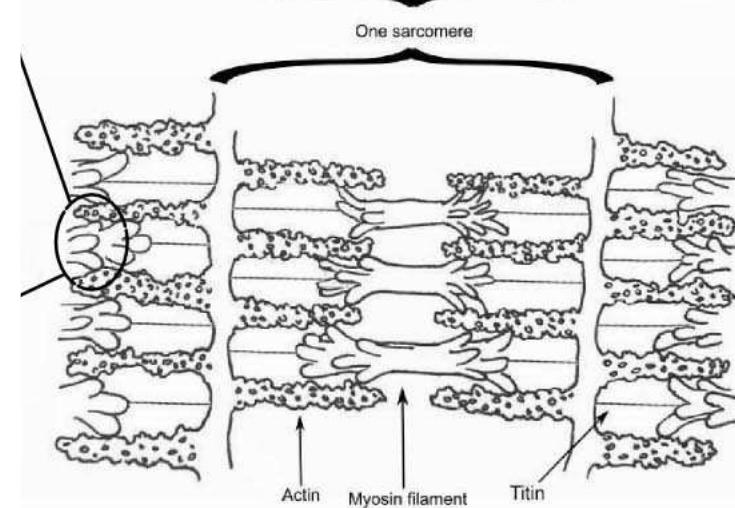
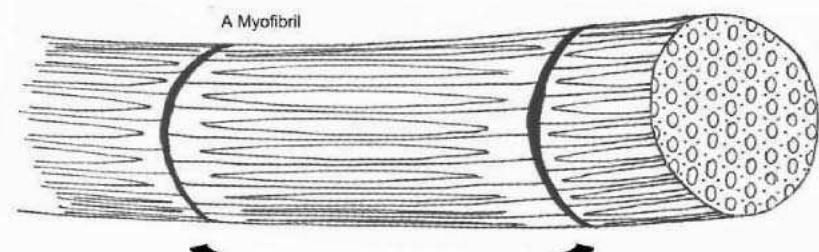
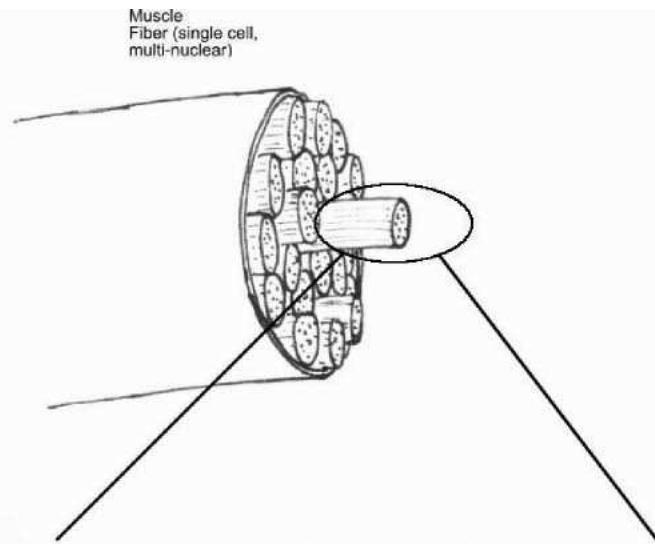
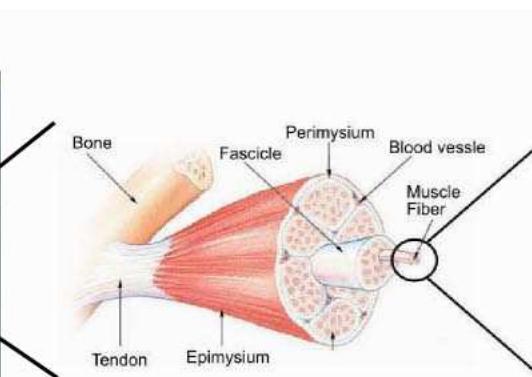
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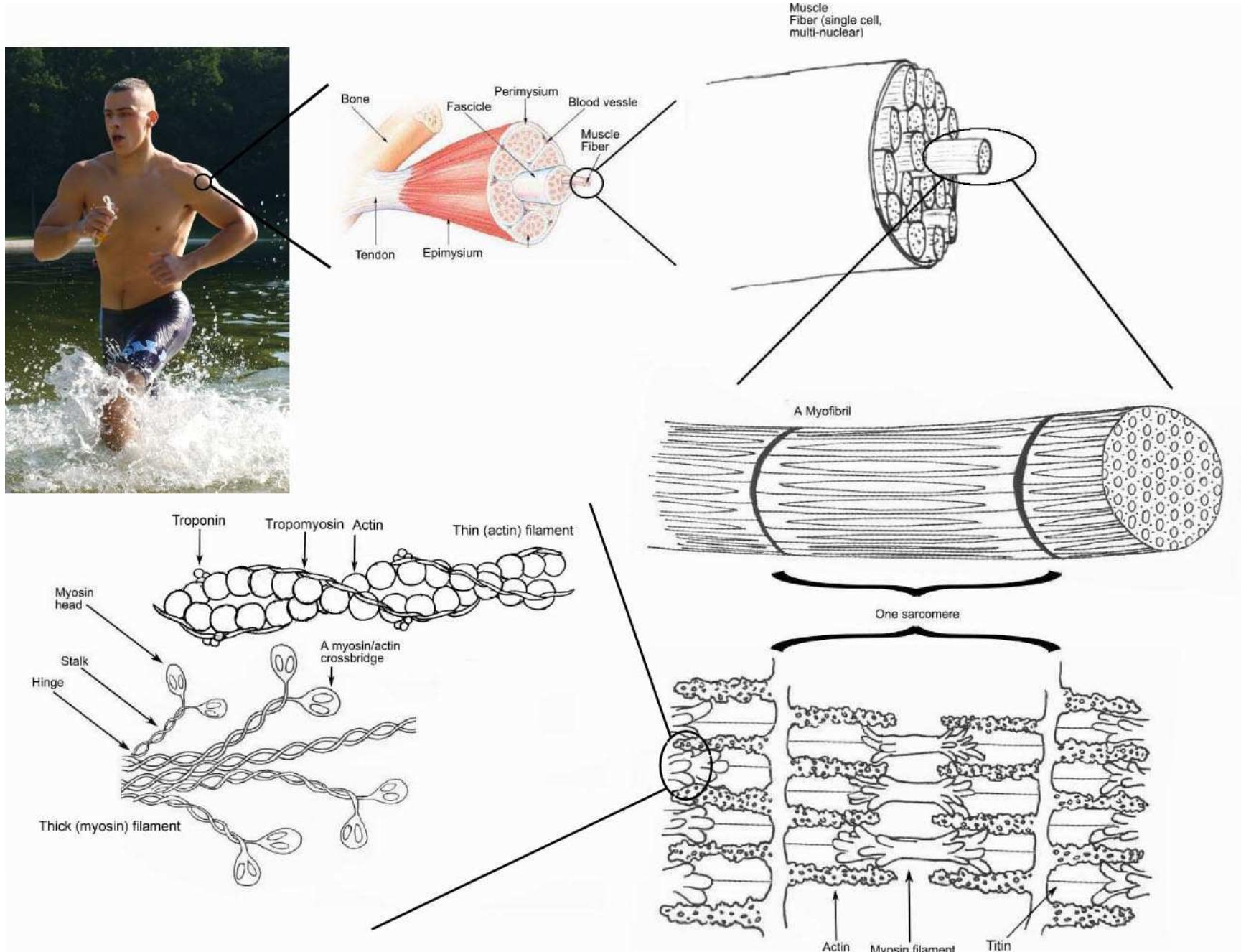
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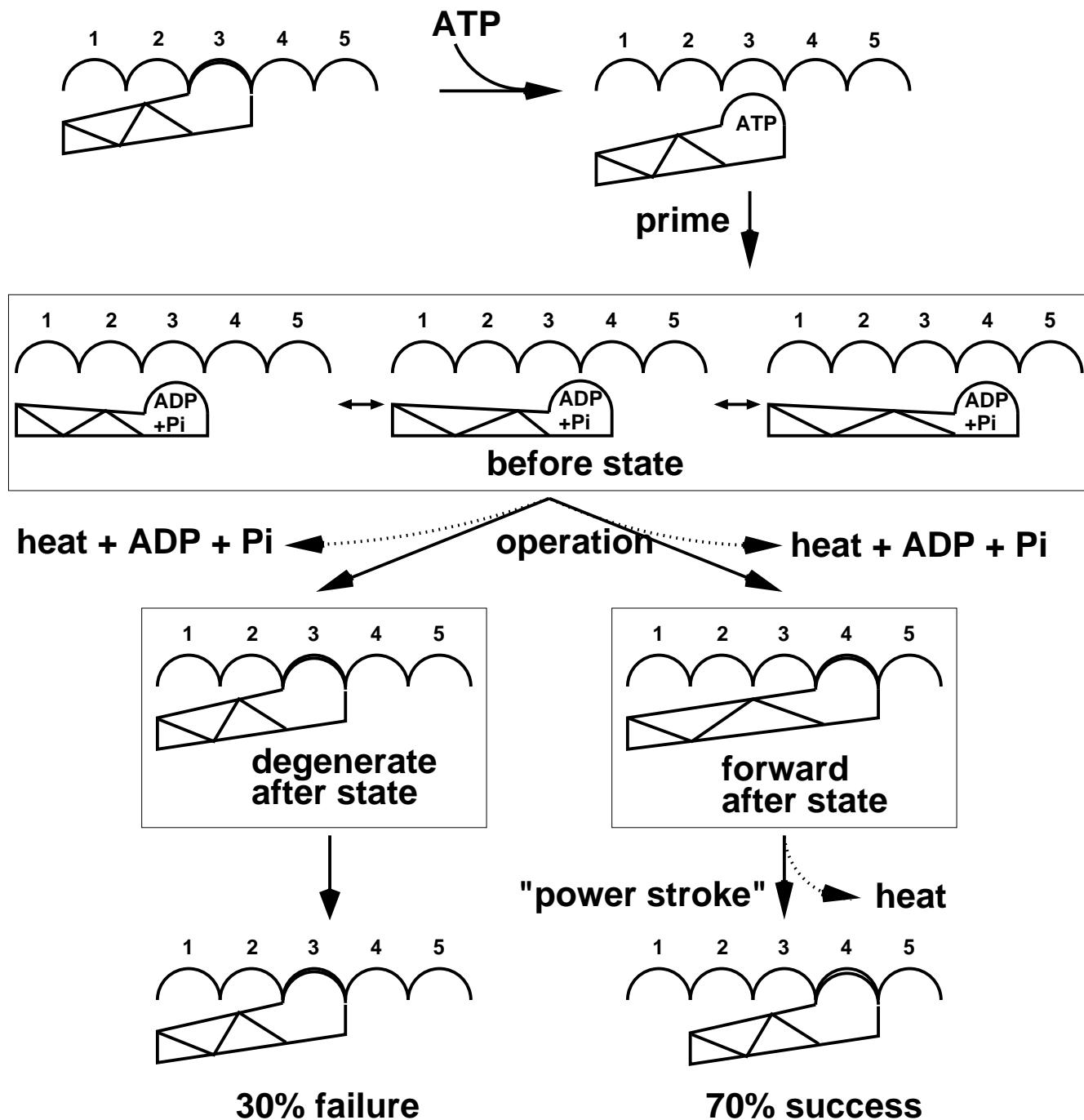
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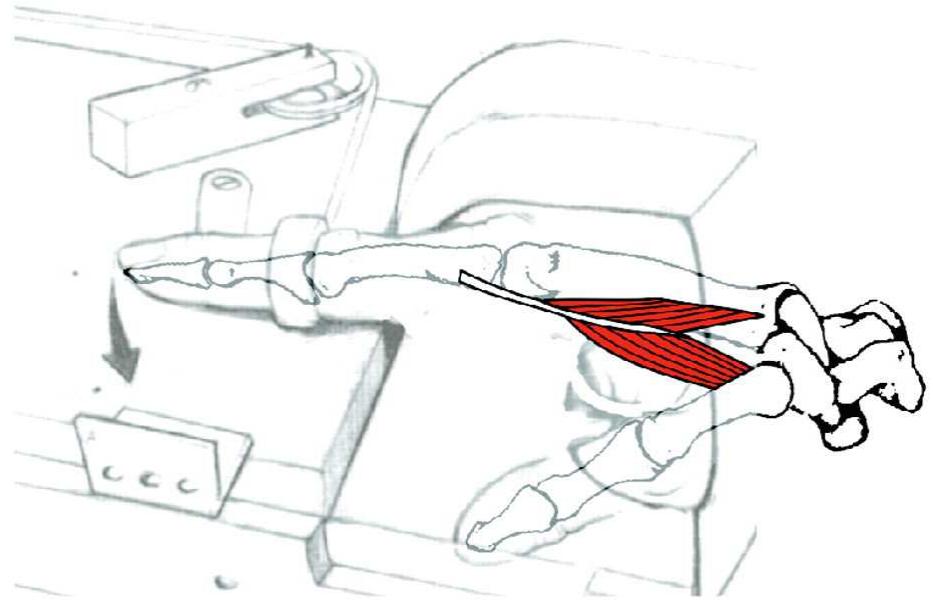
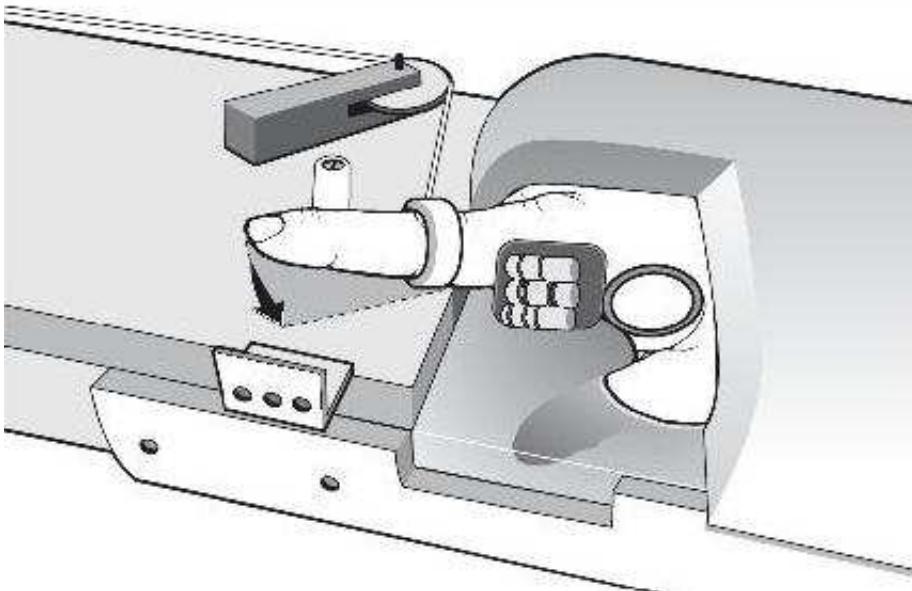
Muscle Structure



Tom's Model of Muscle Mechanism

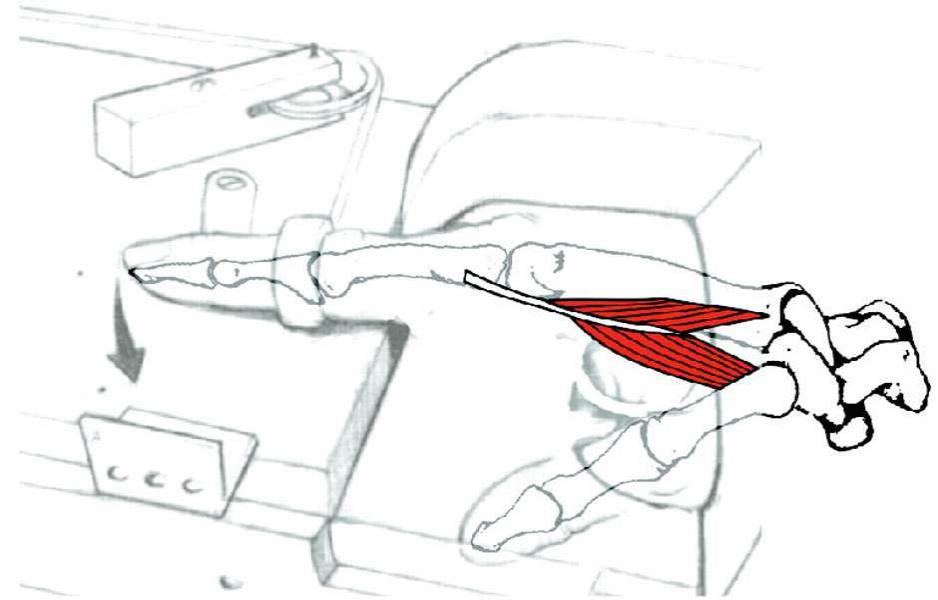
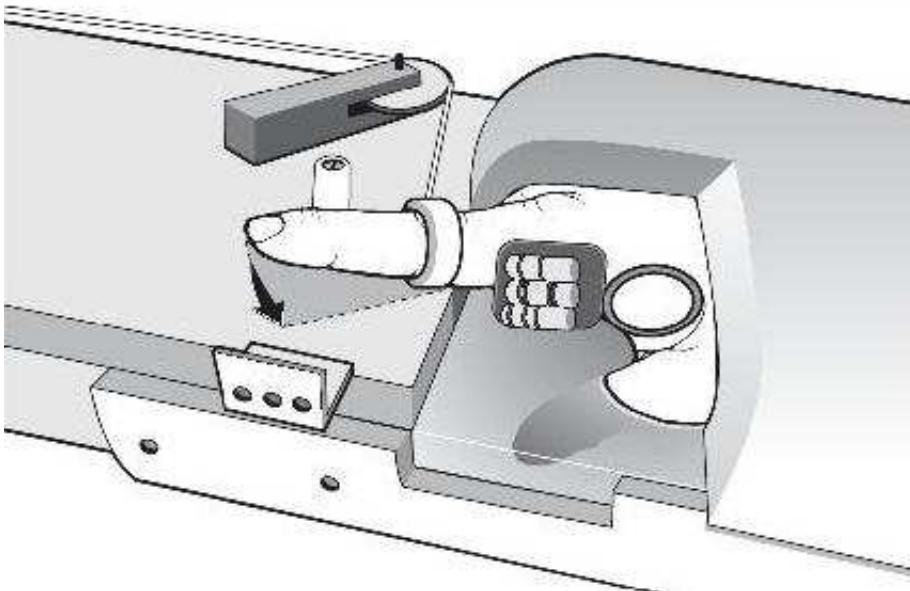


Efficiency of Muscle



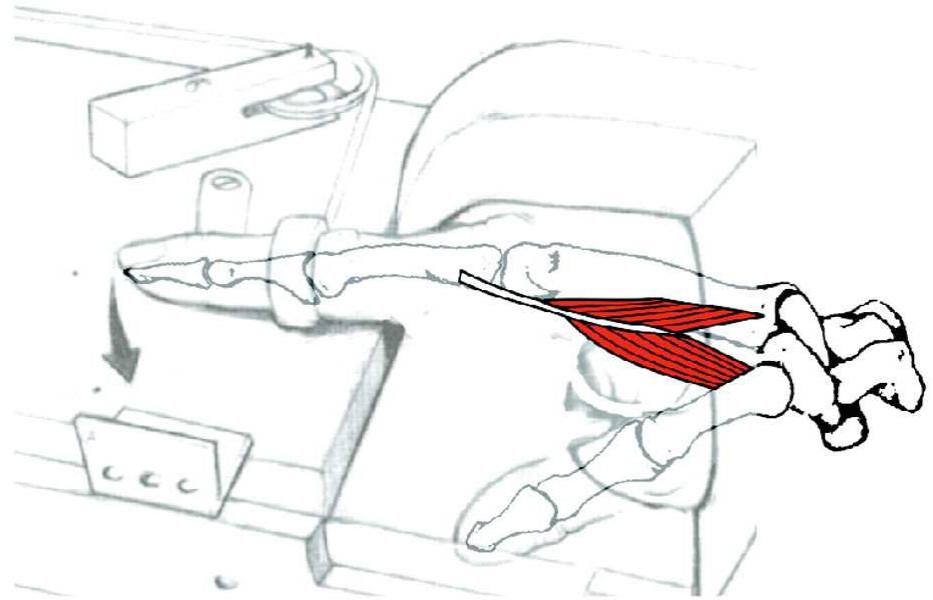
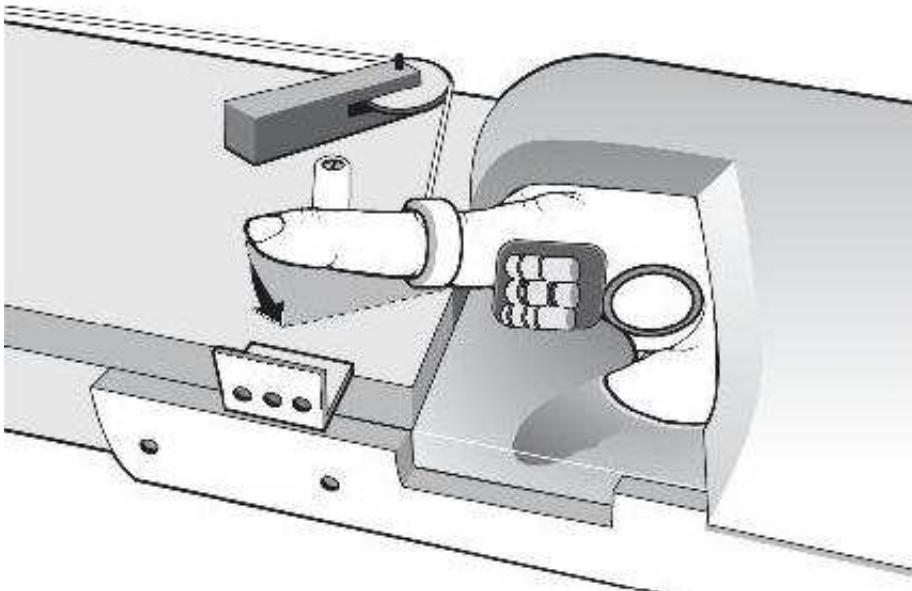
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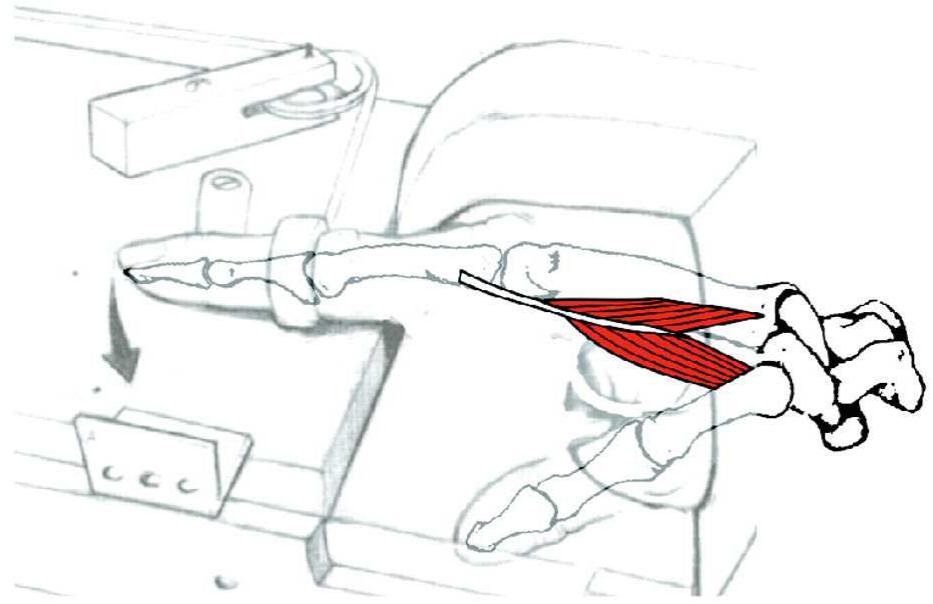
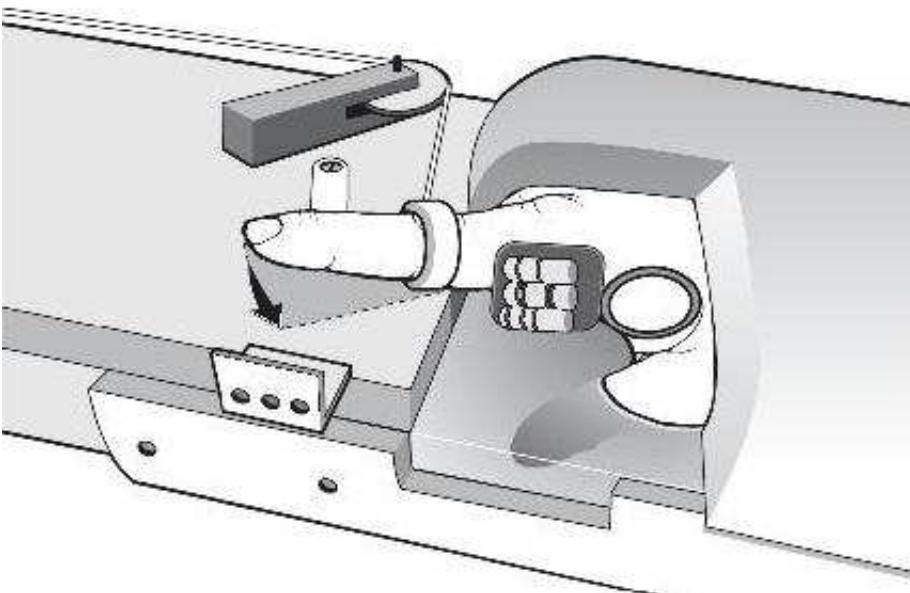
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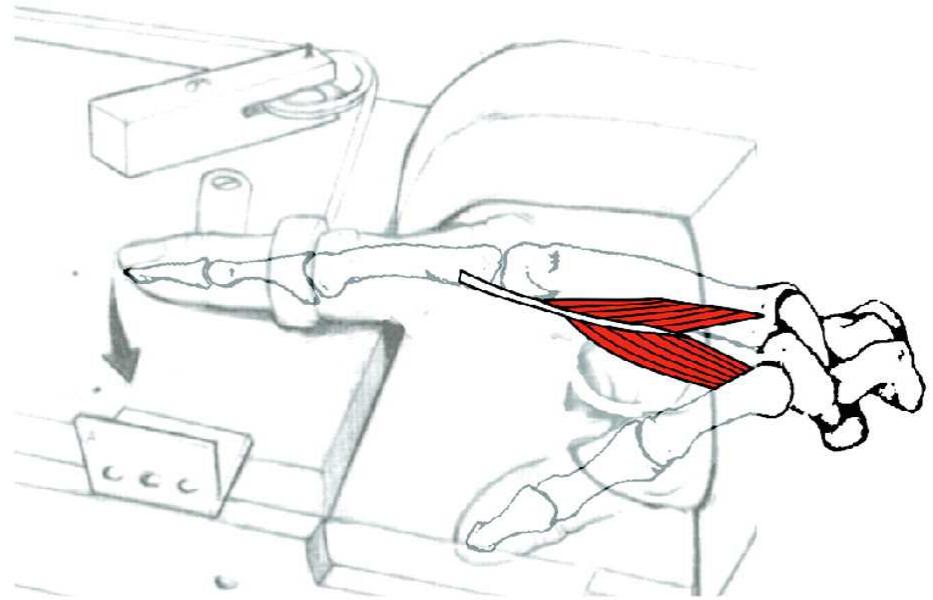
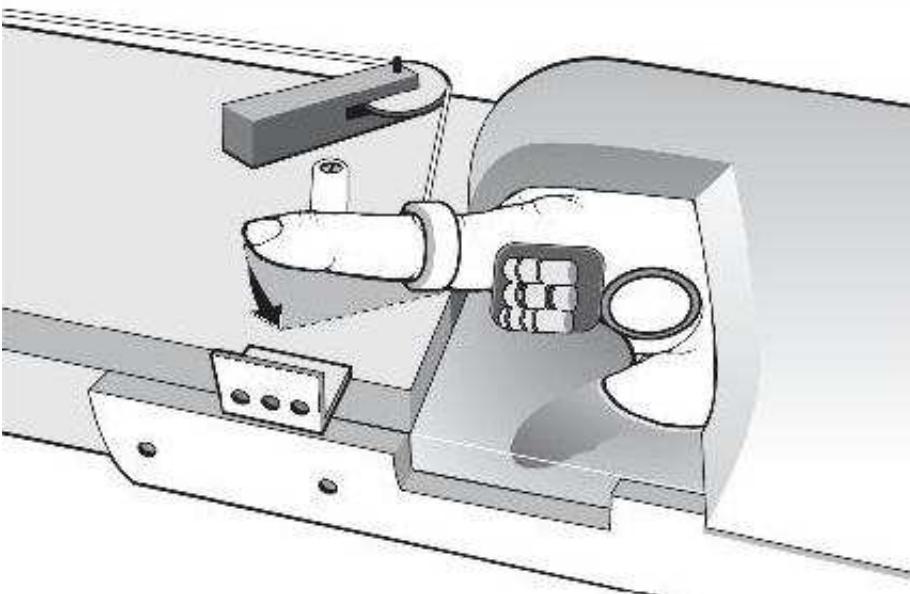
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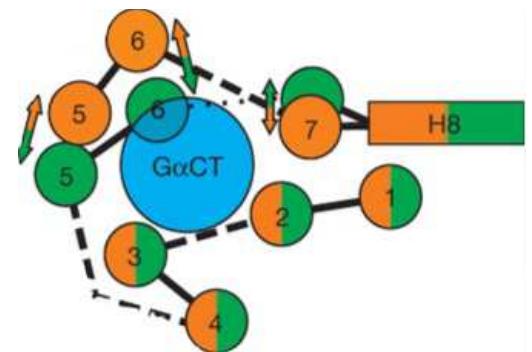
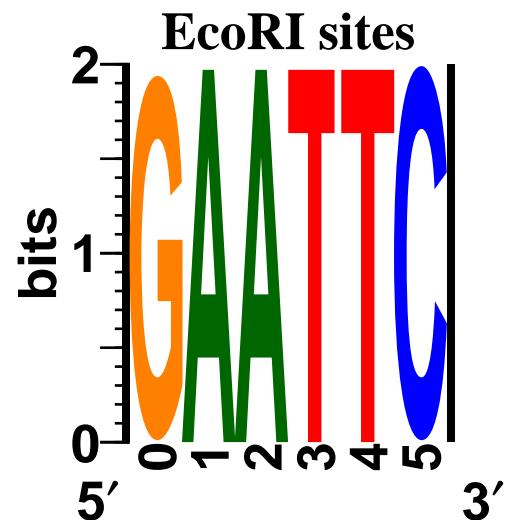
Efficiency of Muscle



- Experiments by Kushmerick's lab since (at least) 1969
- new work: 2008, 2011
- Weight lifting gives work done
- NMR coil gives ATP = energy used
- Efficiency: 0.68 ± 0.09

Why are molecular machines 70% efficient?

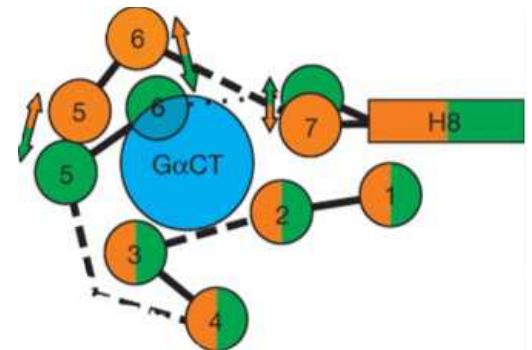
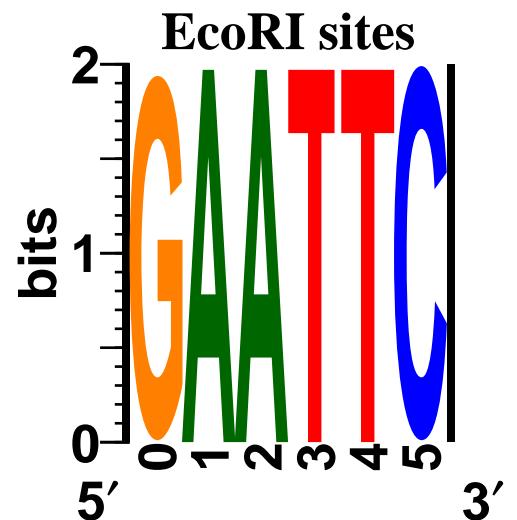
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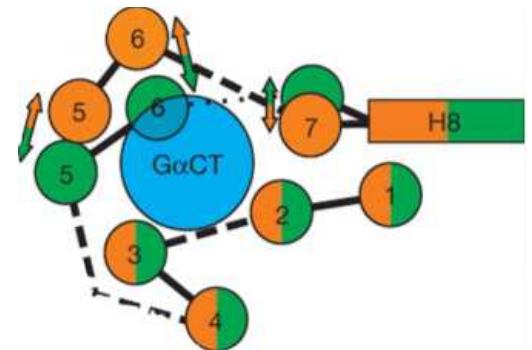
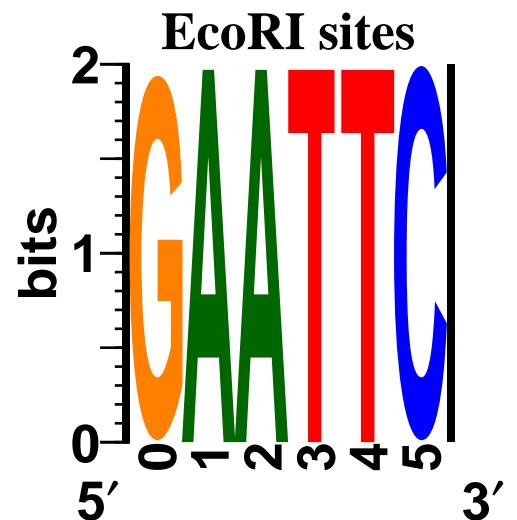
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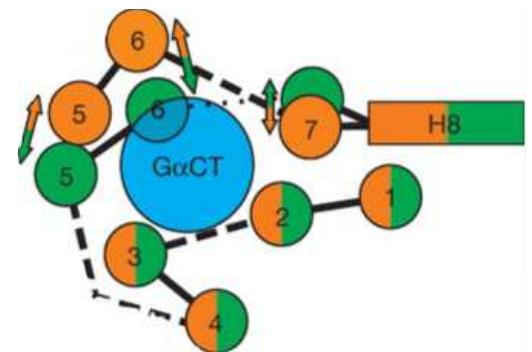
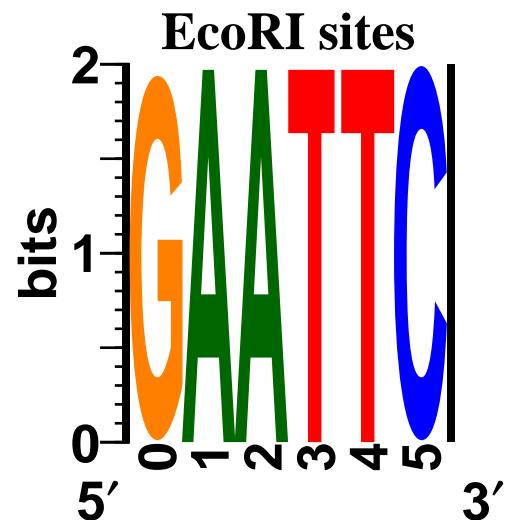
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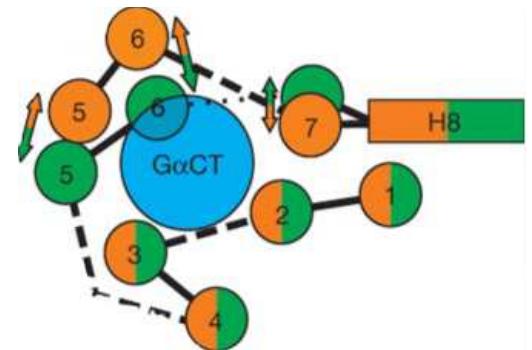
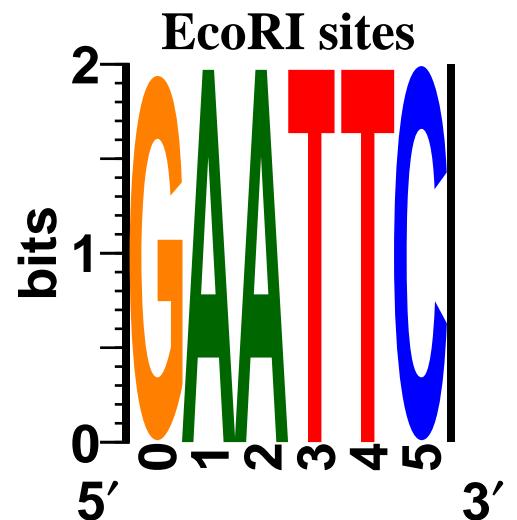
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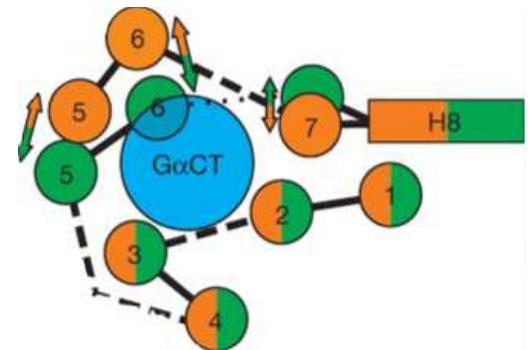
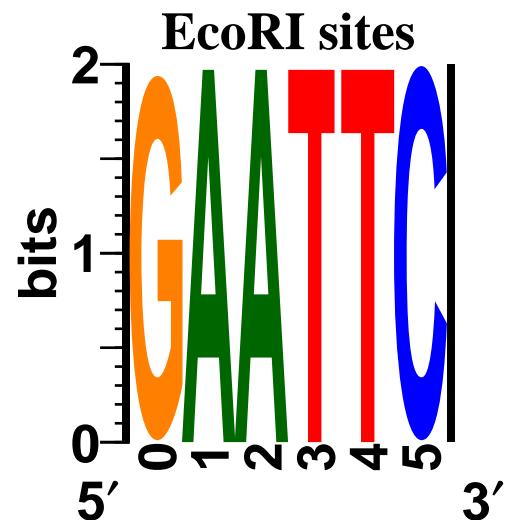


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Why 70% efficiency?



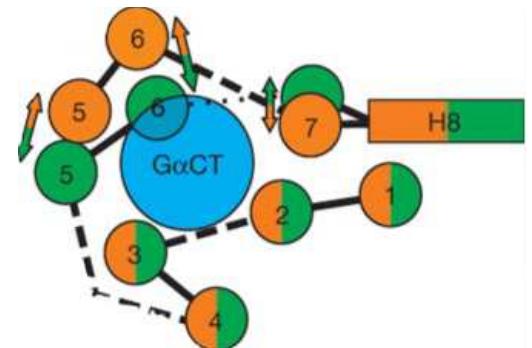
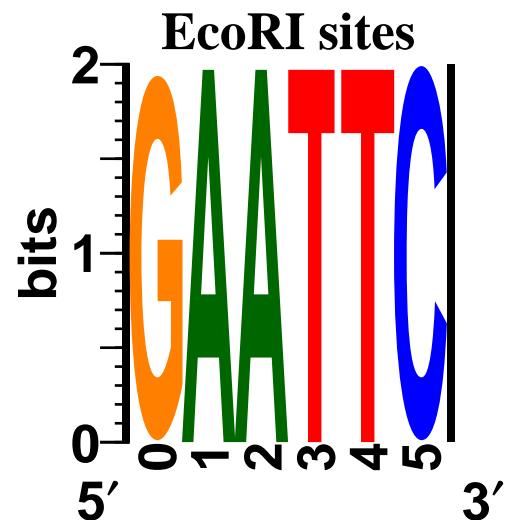
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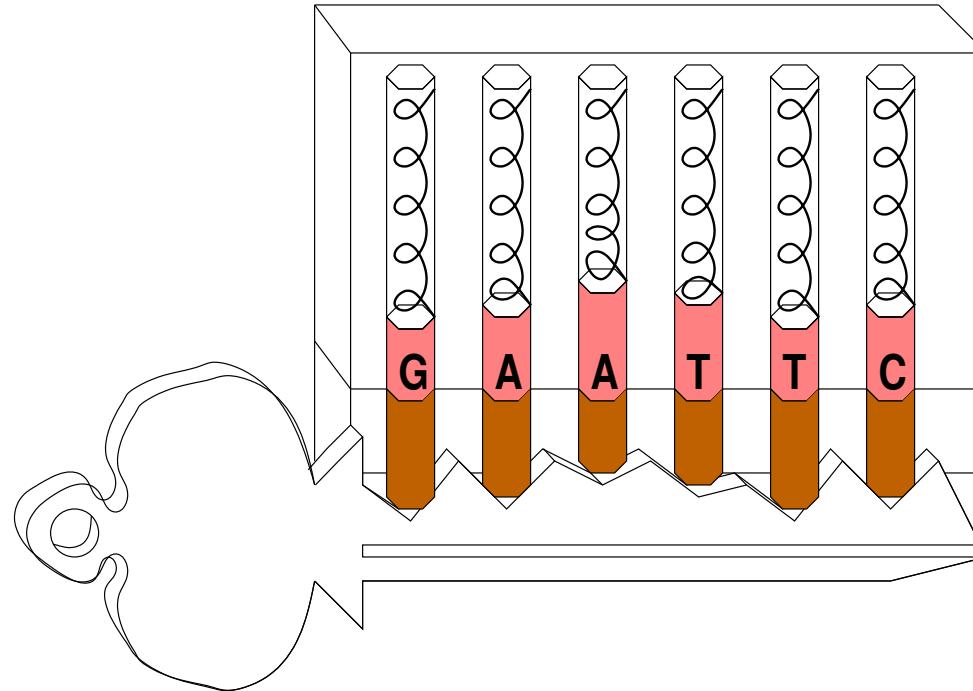
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Why 70% efficiency?

Information theory explanation



Lock and Key

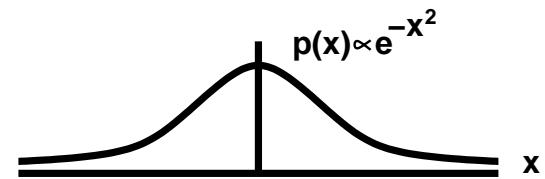


**Like a key in a lock
which has many independent pins,
it takes many numbers
to describe the vibrational state
of a molecular machine**

Gaussians

- Pin motion x has a Gaussian distribution:

$$p(x) = \frac{1}{\sqrt{2\pi\sigma^2}} e^{-\frac{(x-\mu)^2}{2\sigma^2}}$$

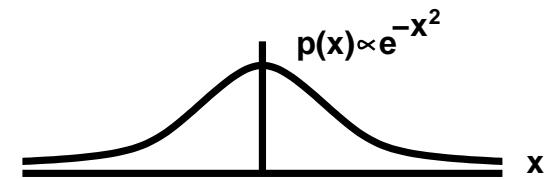


μ = mean, σ = standard deviation

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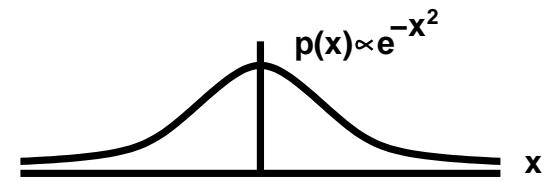
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Gaussians

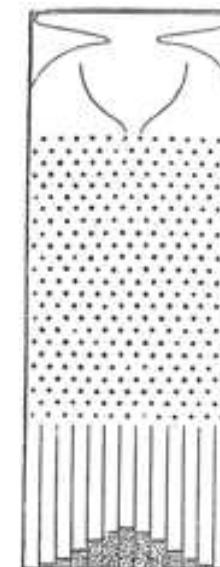
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- Gaussian distributions are generated by the sum of many small random variables
- Drunkard's walk: Galton's quincunx device!



Two Gaussians

$$p(x) = \frac{1}{\sqrt{2\pi\sigma^2}} e^{-\frac{(x-\mu)^2}{2\sigma^2}} \quad (1)$$

$$p(y) = \frac{1}{\sqrt{2\pi\sigma^2}} e^{-\frac{(y-\mu)^2}{2\sigma^2}} \quad (2)$$

Two Gaussians

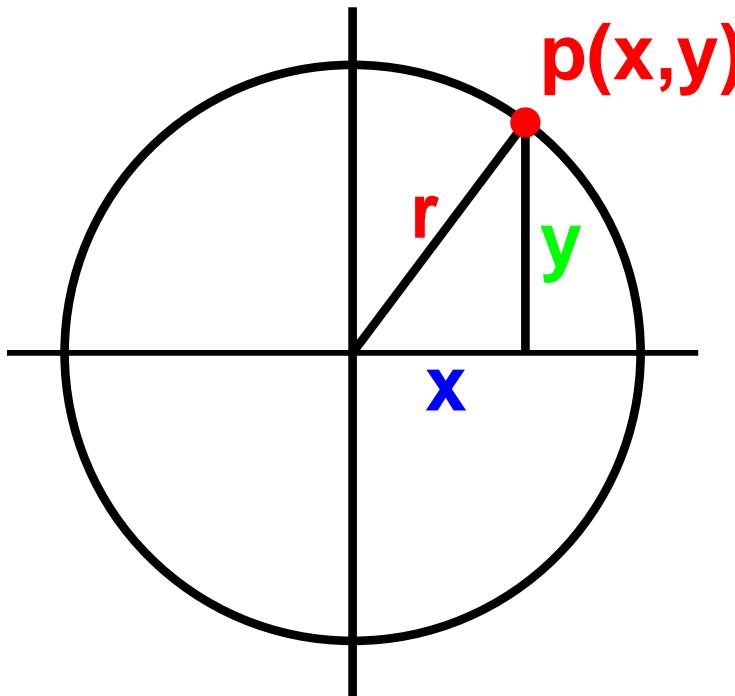
$$p(x) = \frac{1}{\sqrt{2\pi\sigma^2}} e^{-\frac{(x-\mu)^2}{2\sigma^2}} \propto e^{-x^2} \quad (1)$$

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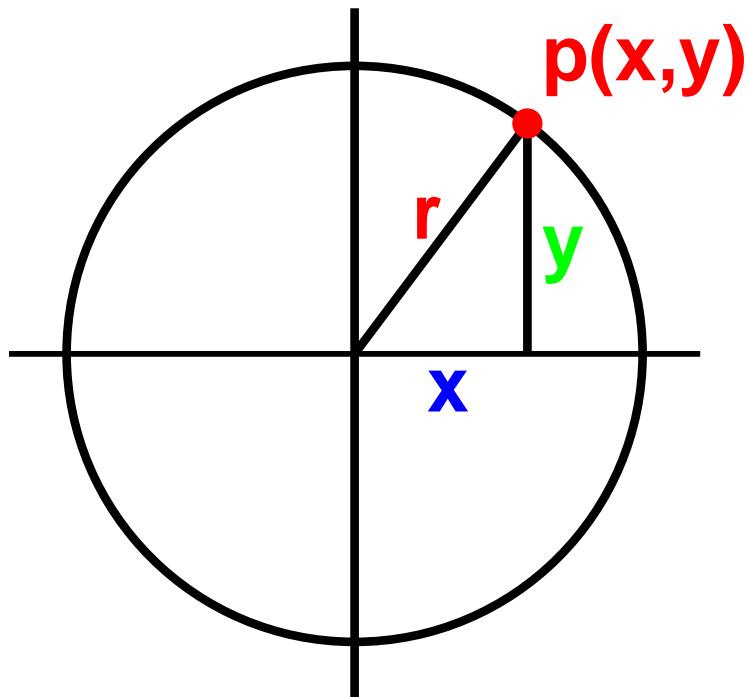
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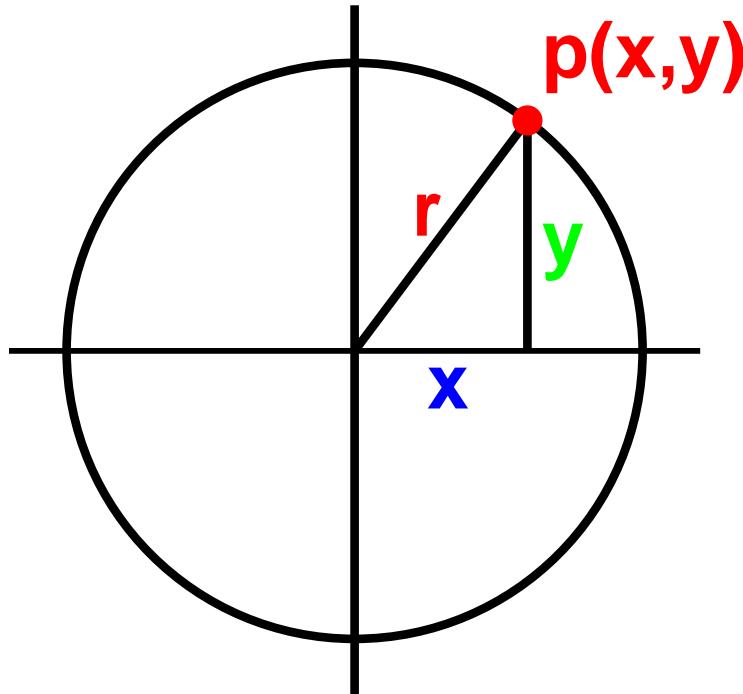


$$p(x, y) = p(x) \times p(y) \quad (3)$$

Two Gaussians

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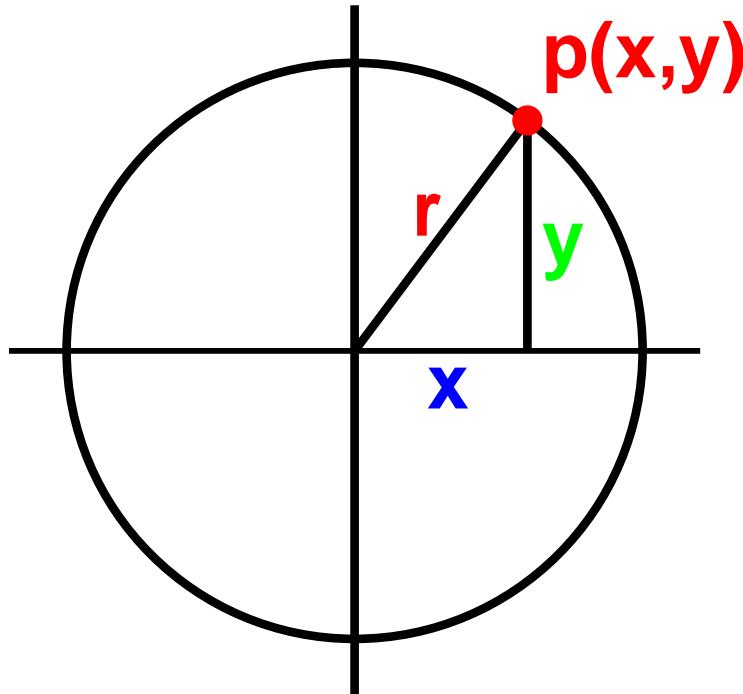
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$$\propto e^{-x^2} \times e^{-y^2} \quad (4)$$

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$$p(y) = \frac{1}{\sqrt{2\pi\sigma^2}} e^{-\frac{(y-\mu)^2}{2\sigma^2}} \propto e^{-y^2} \quad (2)$$



$$p(x, y) = p(x) \times p(y) \quad (3)$$

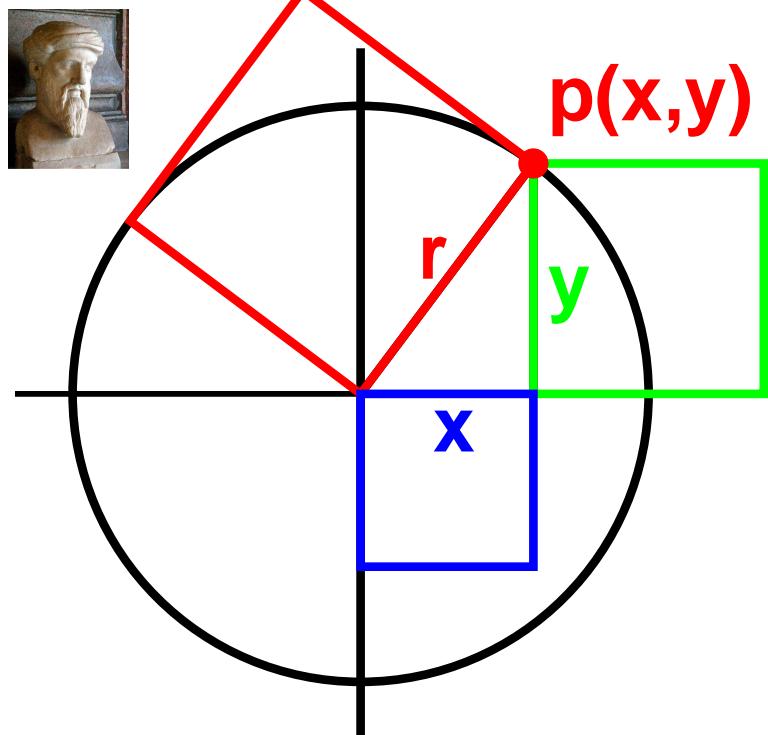
$$\propto e^{-x^2} \times e^{-y^2} \quad (4)$$

$$\propto e^{-(x^2+y^2)} \quad (5)$$

Two Gaussians

$$p(x) = \frac{1}{\sqrt{2\pi\sigma^2}} e^{-\frac{(x-\mu)^2}{2\sigma^2}} \propto e^{-x^2} \quad (1)$$

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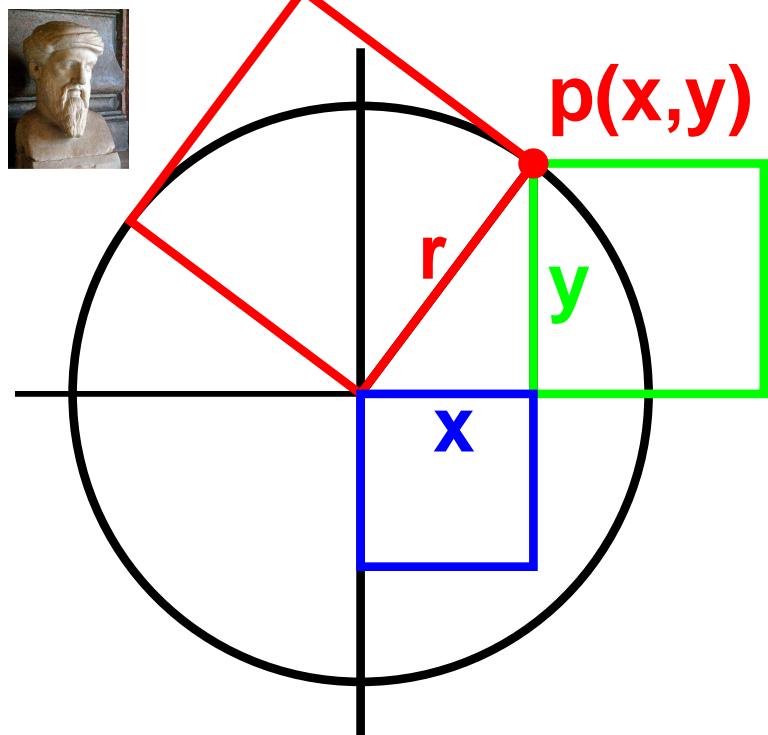
$$\propto e^{-(x^2+y^2)} \quad (5)$$

$$\propto e^{-r^2} \quad (6)$$

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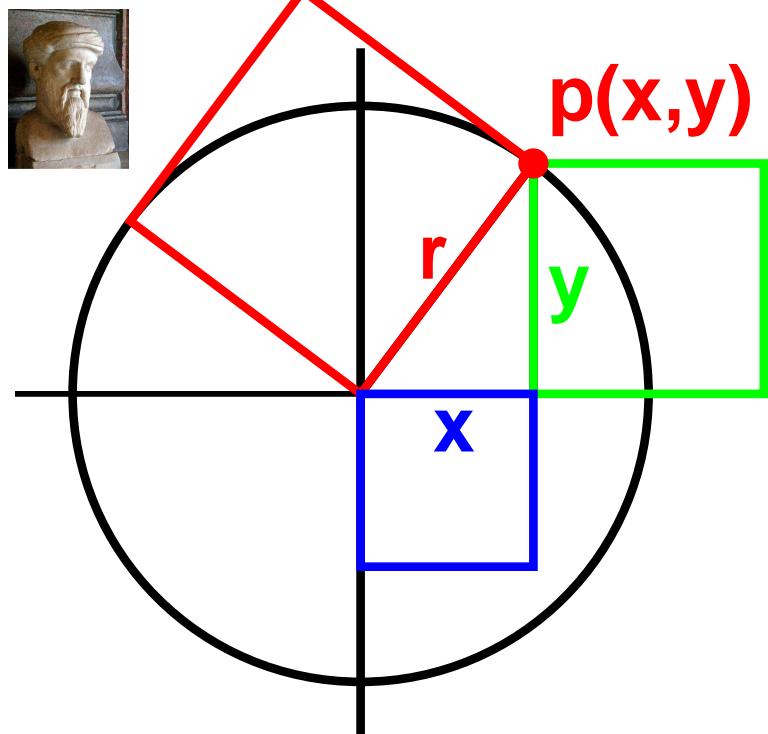
$$\propto e^{-r^2} \quad (6)$$

If $p(x, y)$ is a constant, then r is a constant.

Two Gaussians

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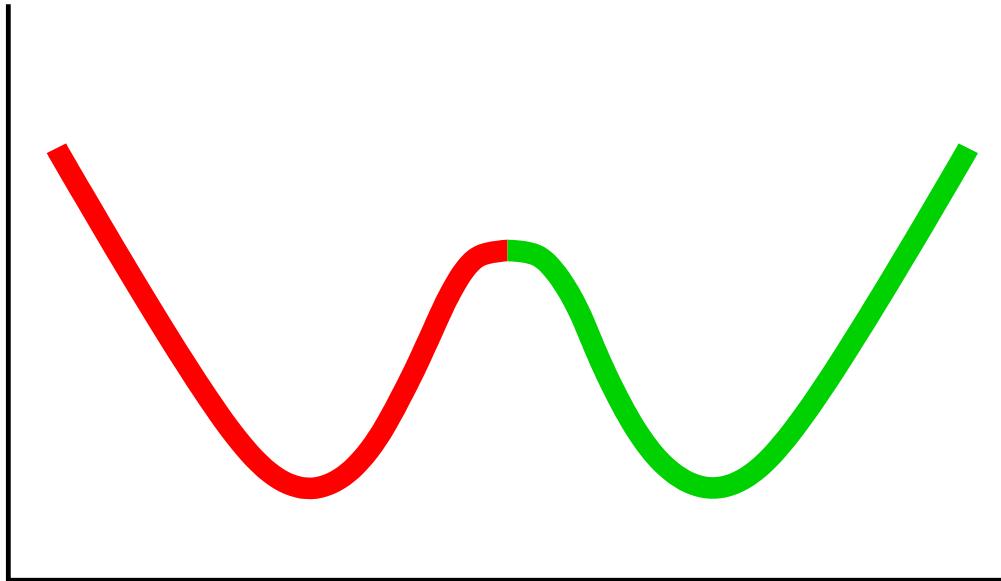
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If $p(x, y)$ is a constant, then r is a constant.

Circular distribution!

1 Dimension

Energy



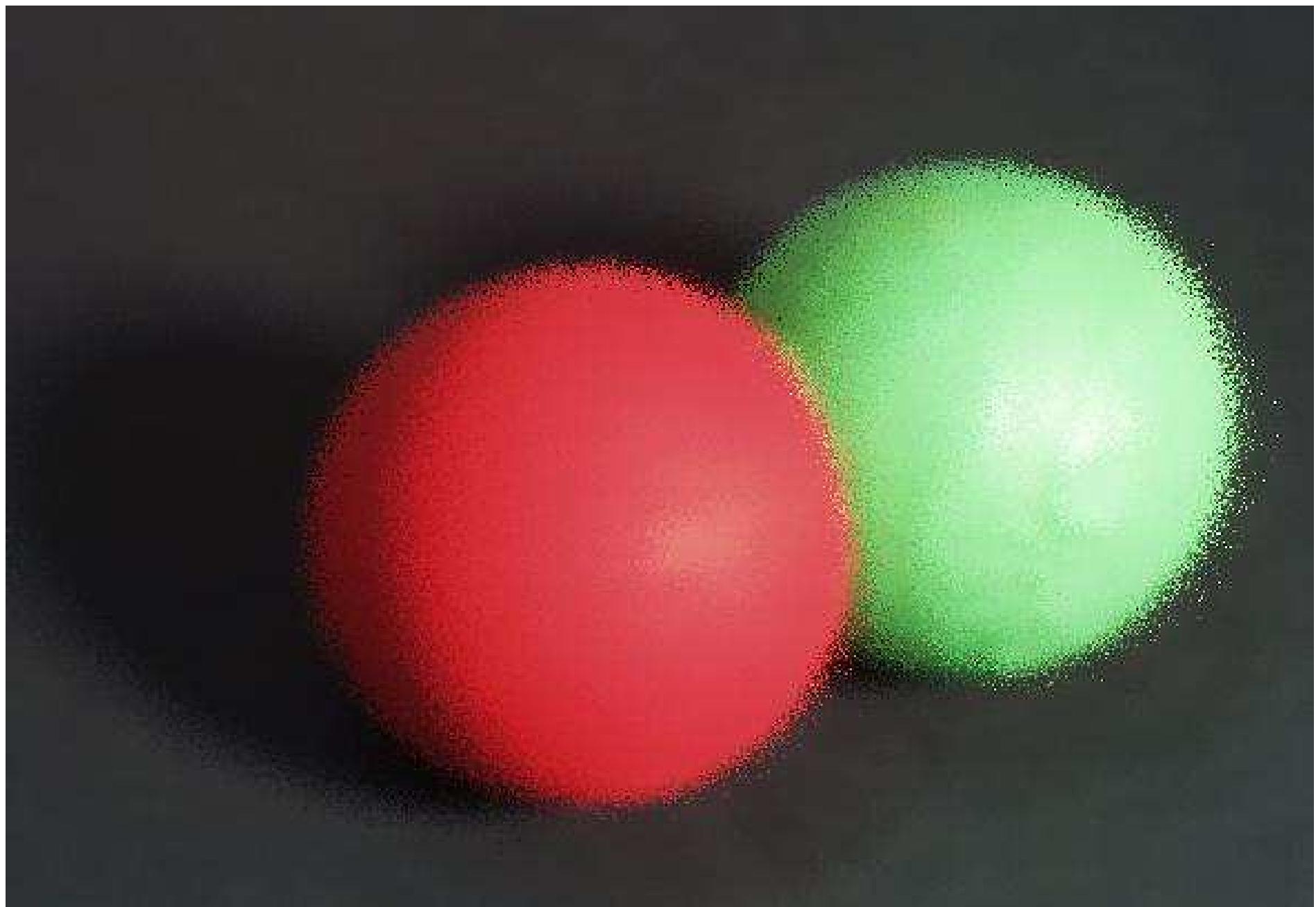
States

1 dimension is too simple!

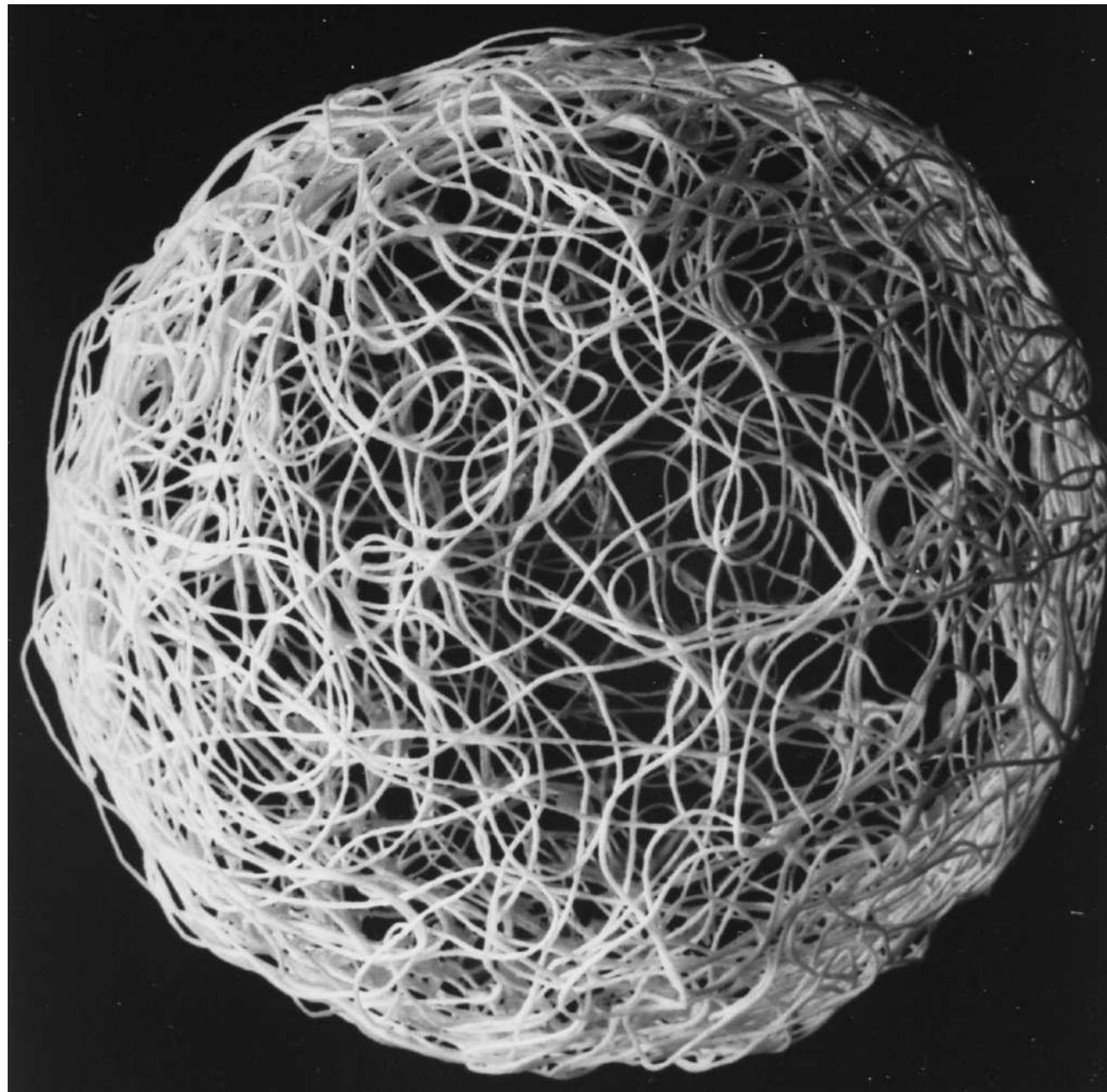
Bowls in 2 Dimensions



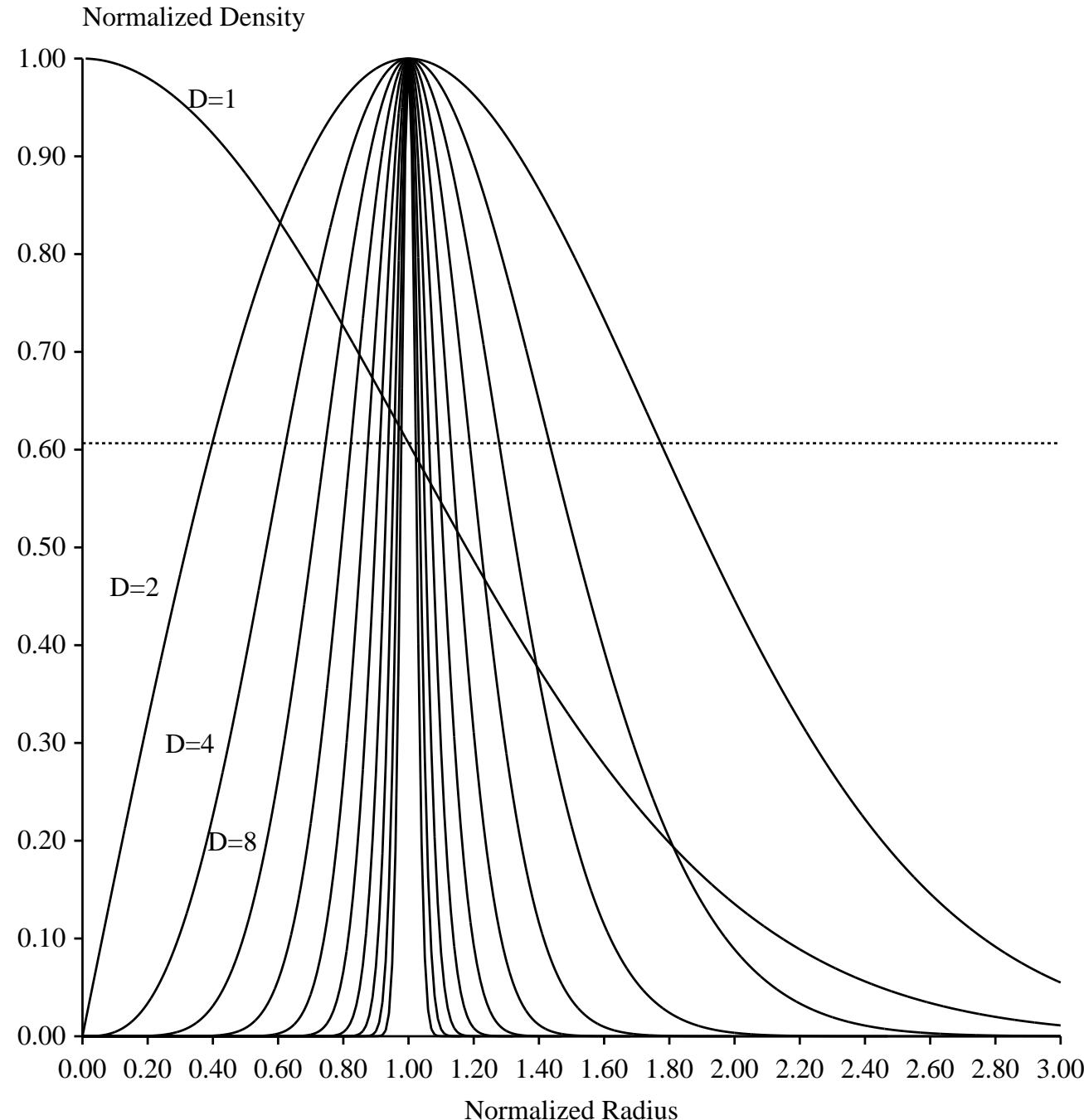
Spheres in 3 Dimensions



N Dimensional Sphere



Spheres tighten in high dimensions



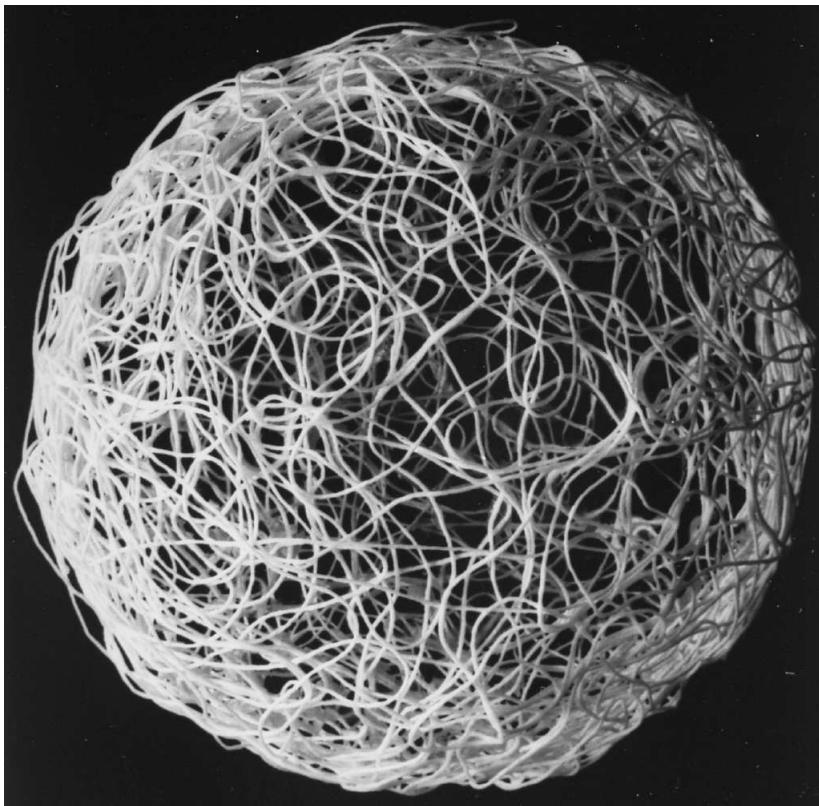
Good Sphere Packing



Good packing of spheres gives a molecule the capacity to make selections efficiently

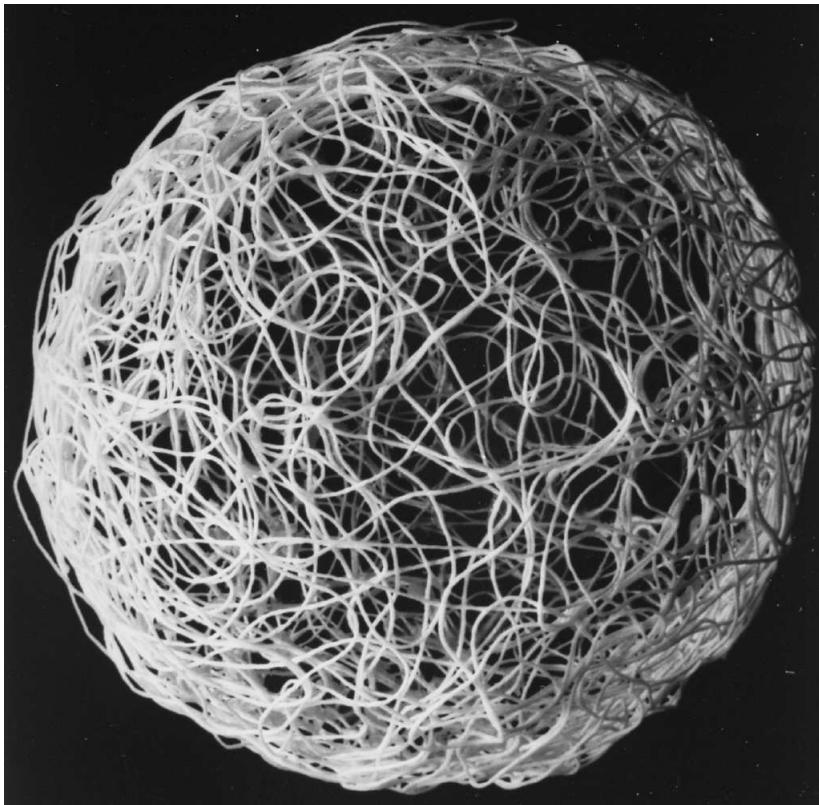
N Dimensional Sphere Separation

Degenerate Sphere

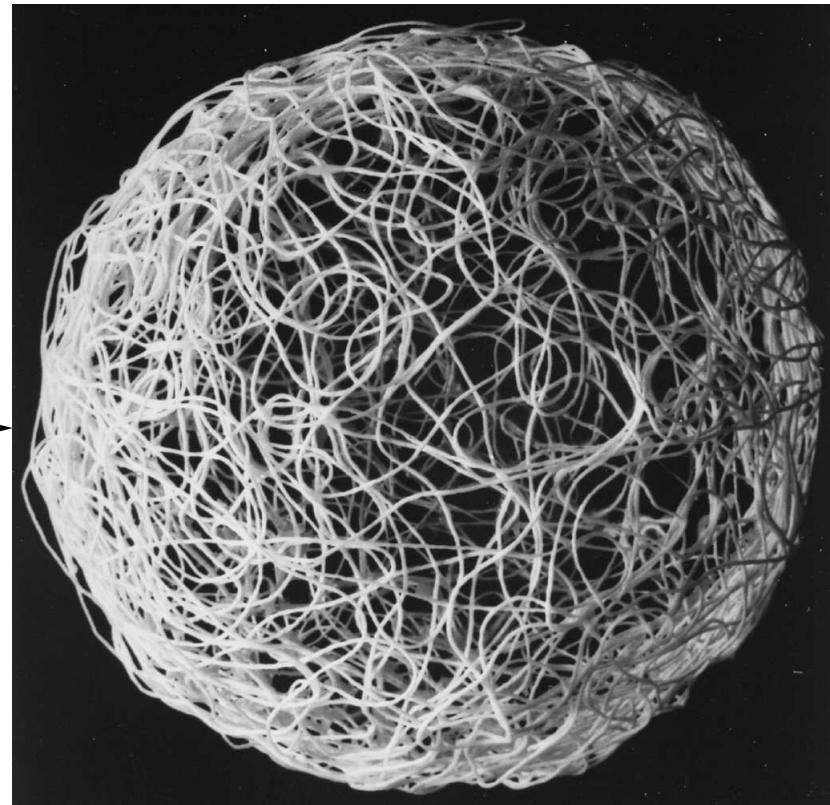


N Dimensional Sphere Separation

Degenerate Sphere

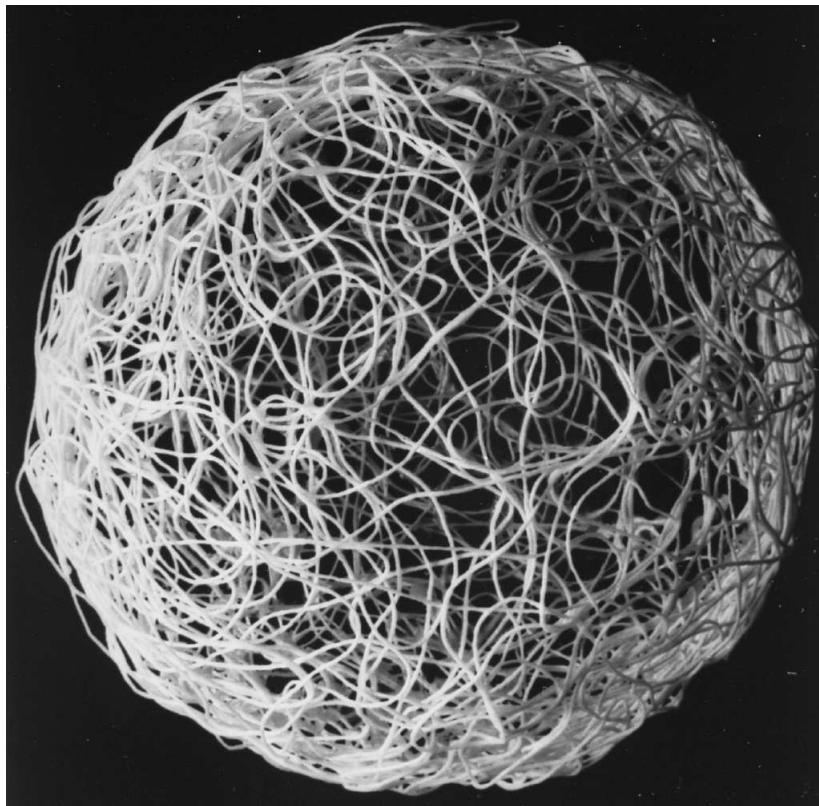


Forward Sphere

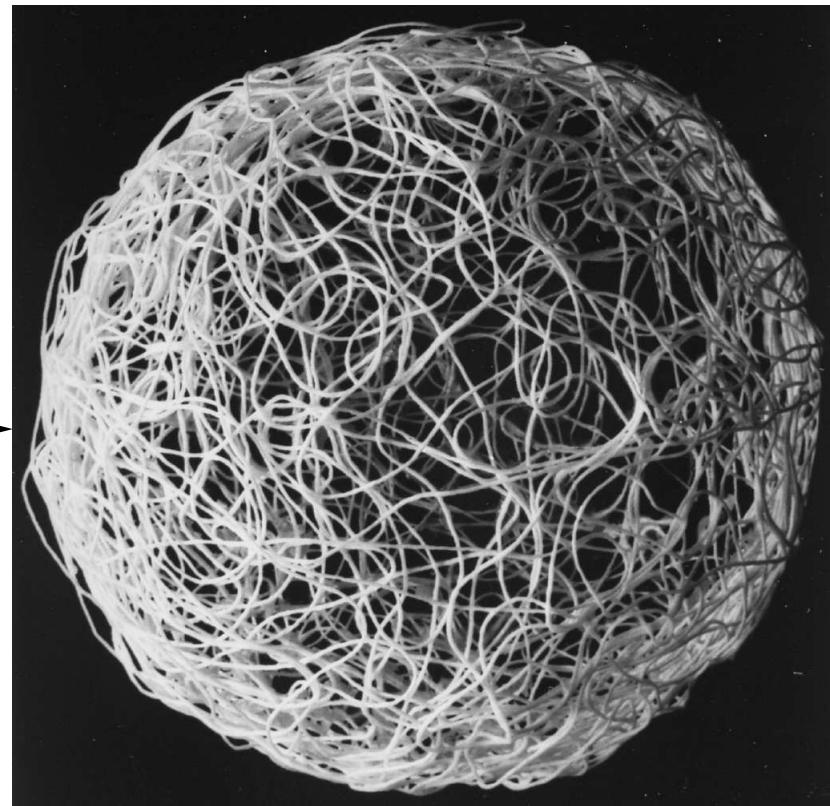


N Dimensional Sphere Separation

Degenerate Sphere



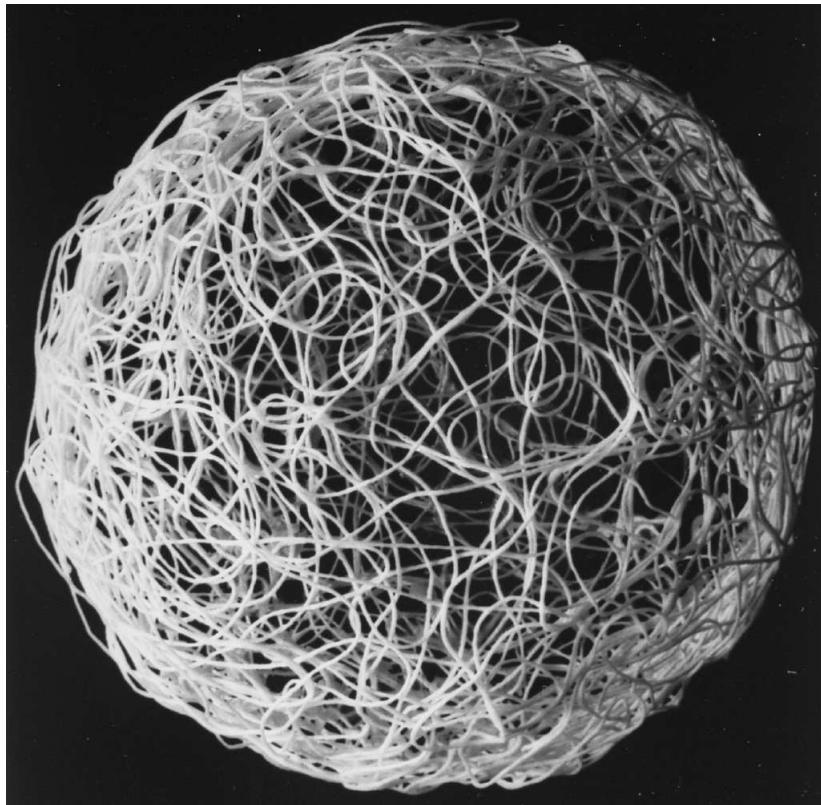
Forward Sphere



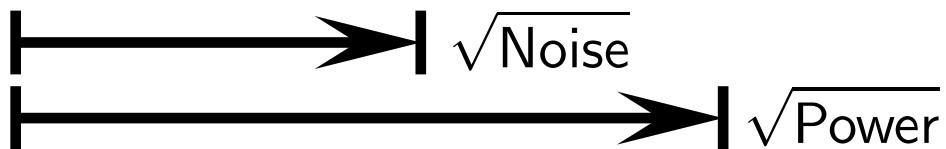
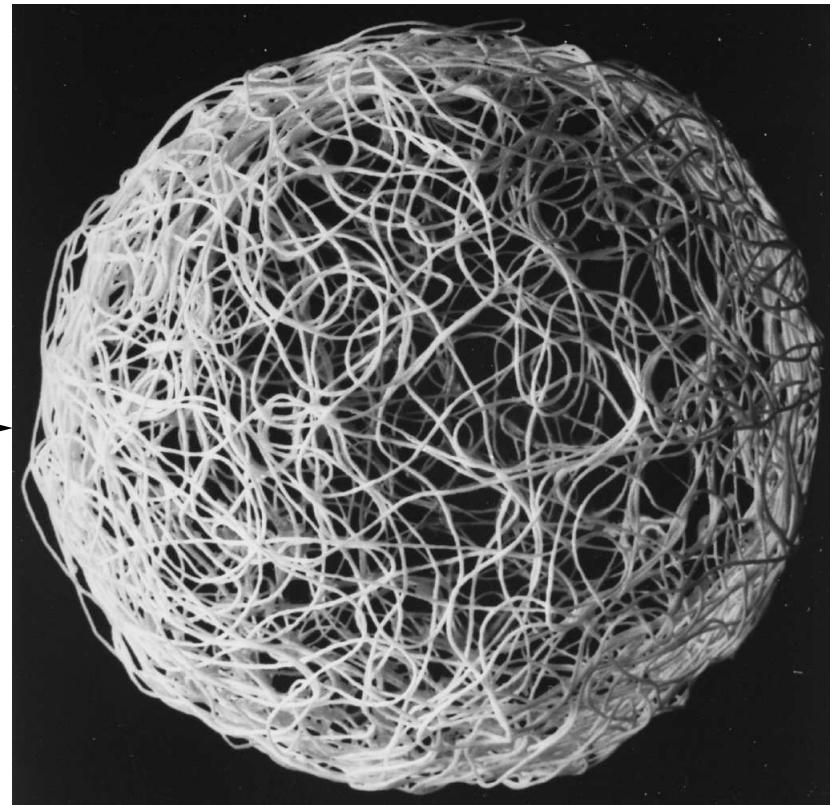
→ $\sqrt{\text{Noise}}$

N Dimensional Sphere Separation

Degenerate Sphere

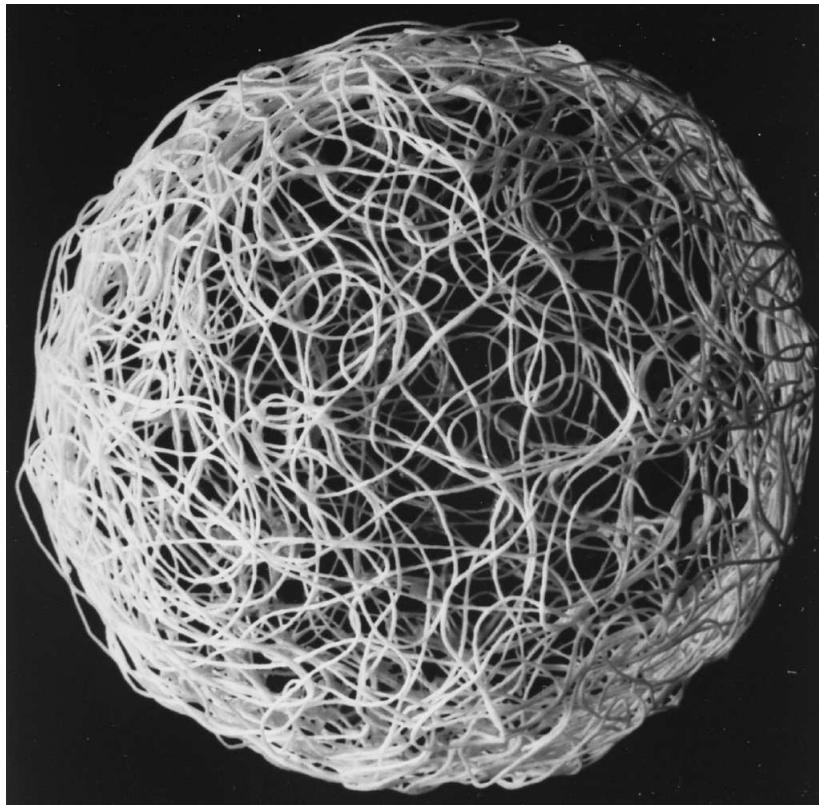


Forward Sphere

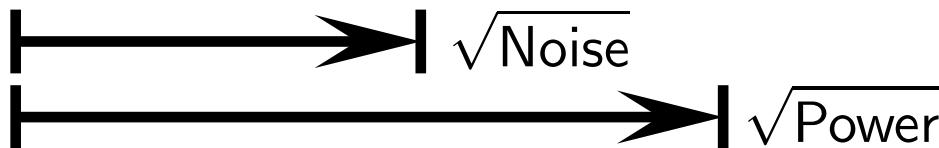
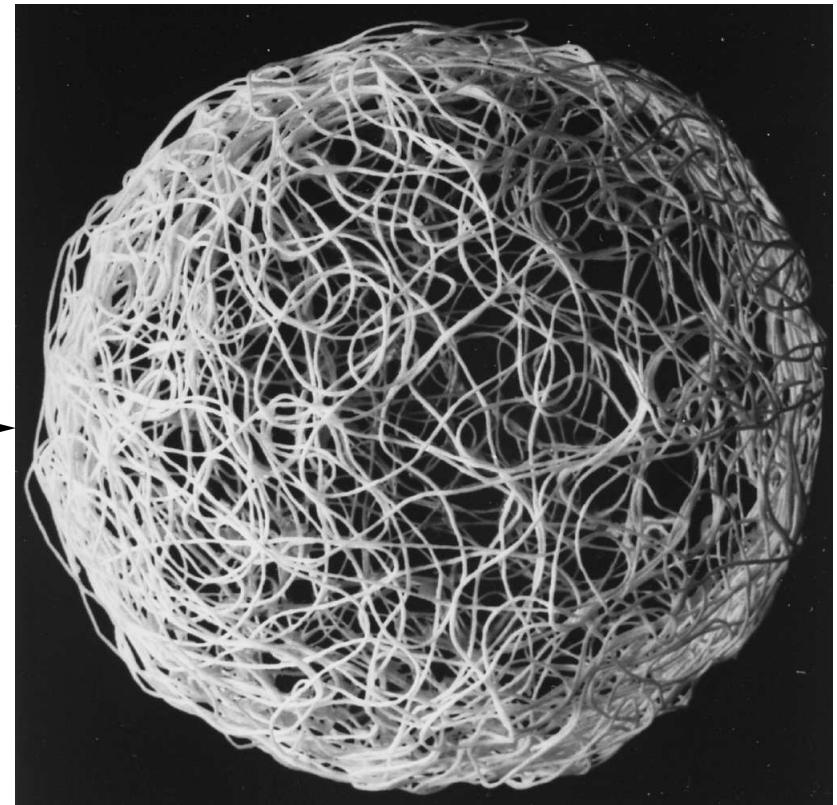


N Dimensional Sphere Separation

Degenerate Sphere



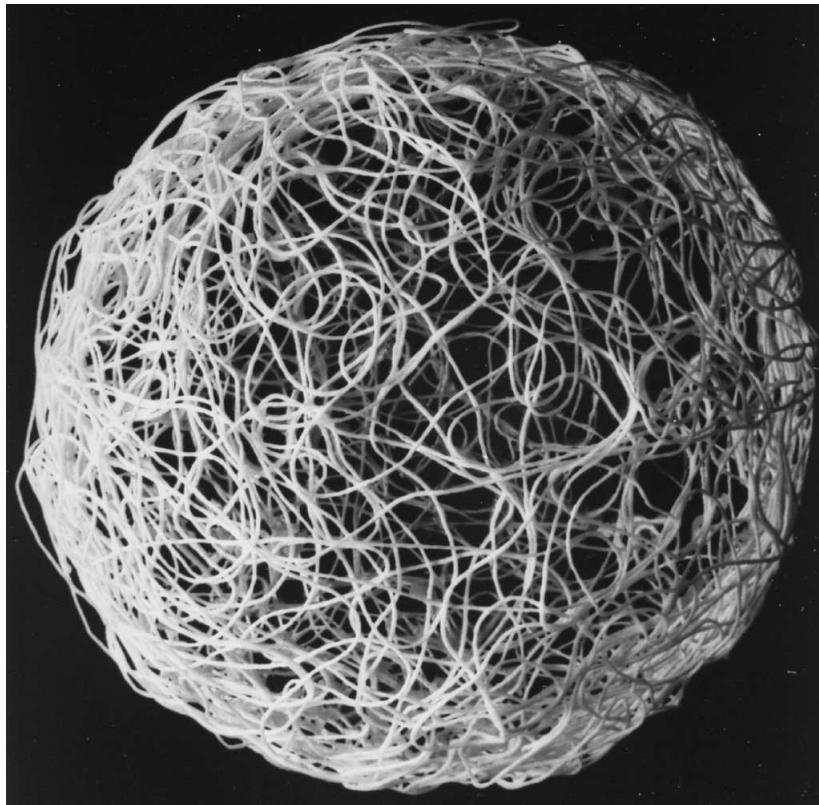
Forward Sphere



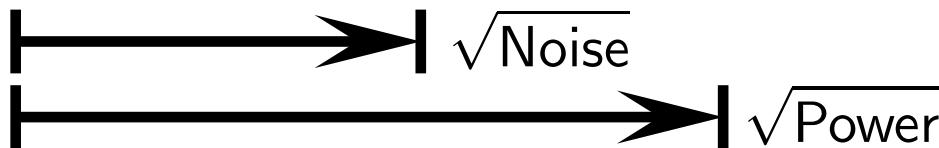
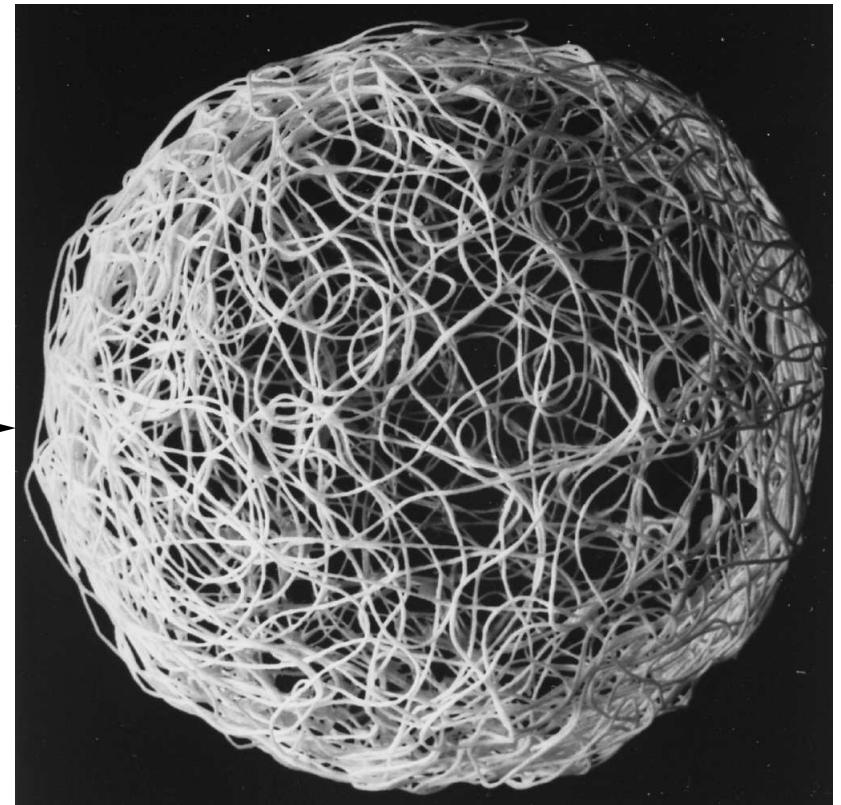
Energy dissipated to escape the Degenerate Sphere must exceed the Noise

N Dimensional Sphere Separation

Degenerate Sphere



Forward Sphere



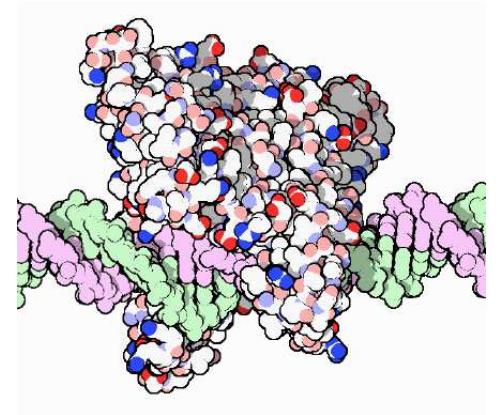
Energy dissipated to escape the Degenerate Sphere must exceed the Noise

$$\sqrt{\text{Power}} > \sqrt{\text{Noise}}$$

Theoretical Isothermal Efficiency

- For molecular states of molecules with d_{space} ‘parts’ P_y energy is dissipated for noise N_y and

$$C_y = d_{space} \log_2(P_y/N_y + 1) \leftarrow \text{machine capacity}$$

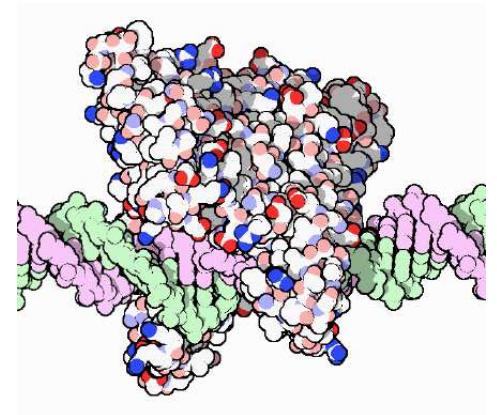


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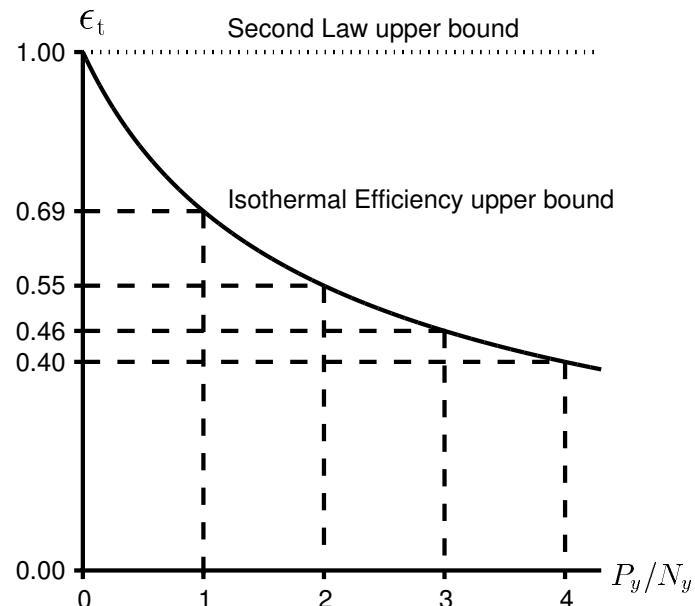
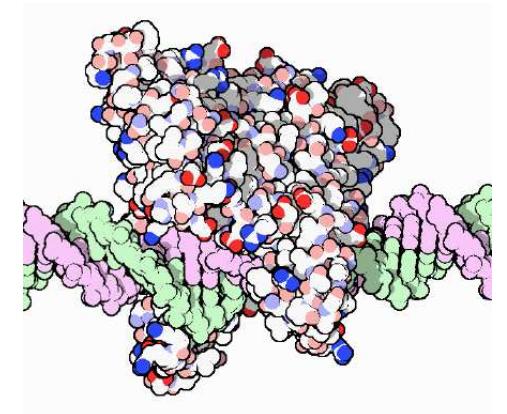


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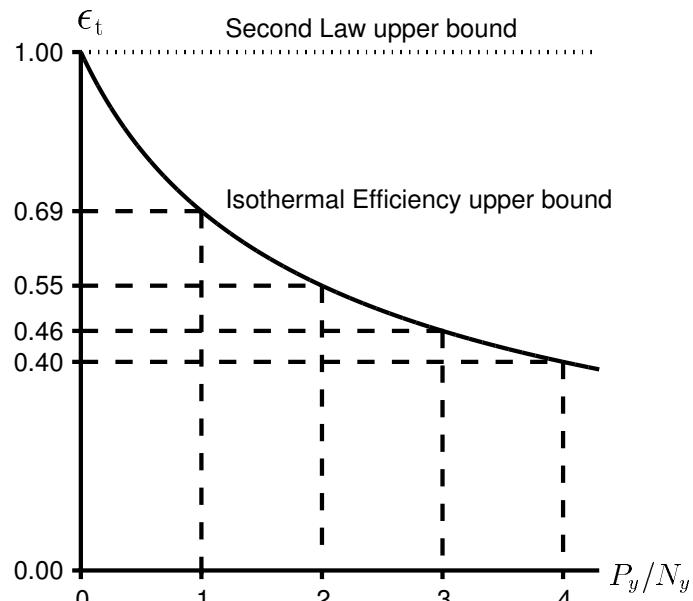
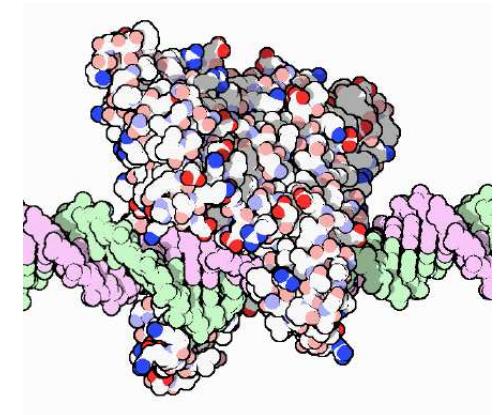
The curve is an upper bound

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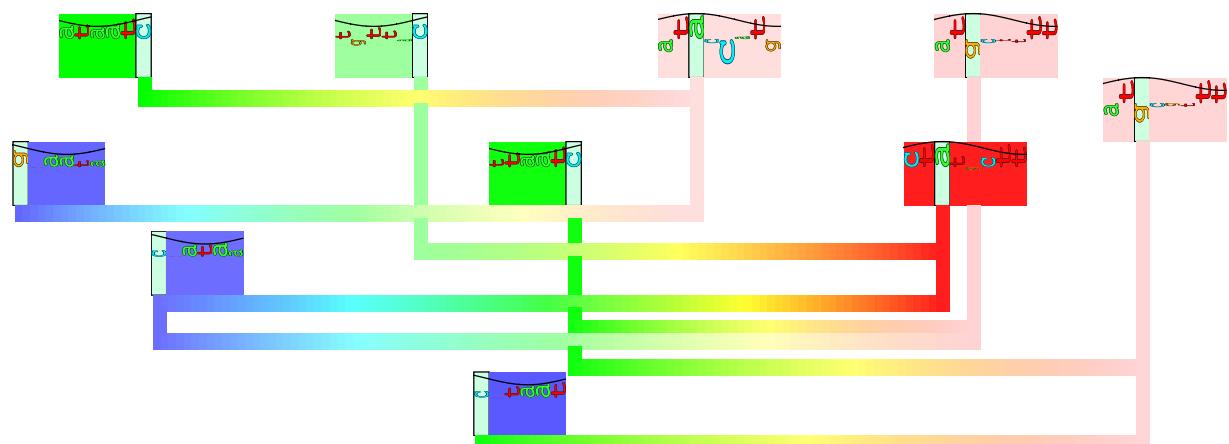


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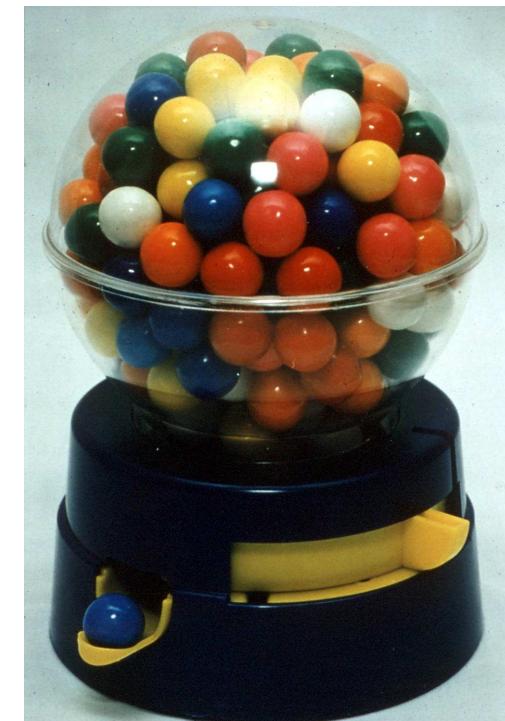
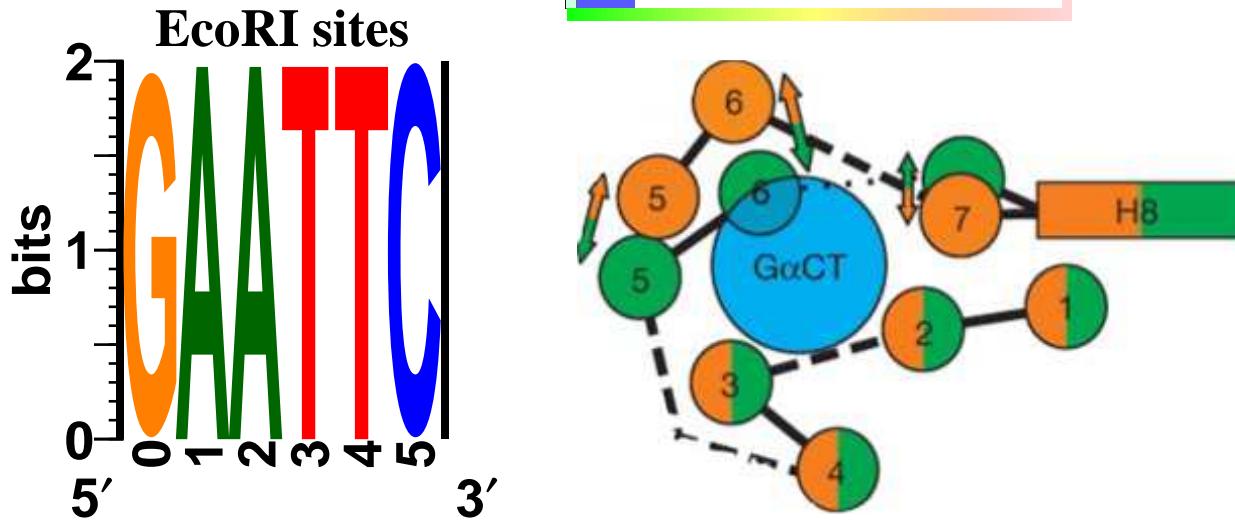
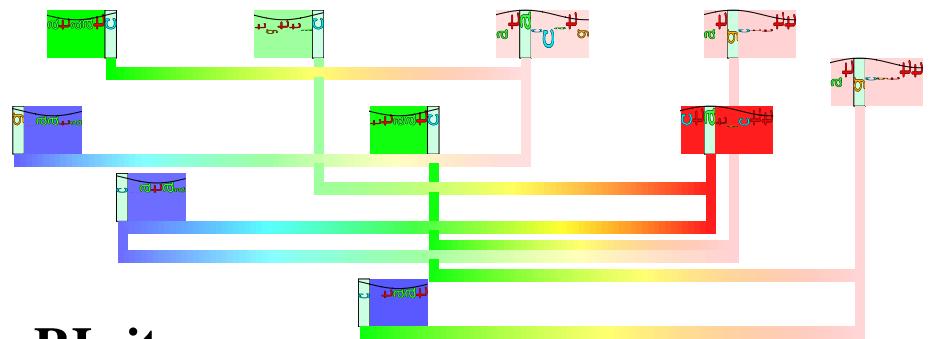
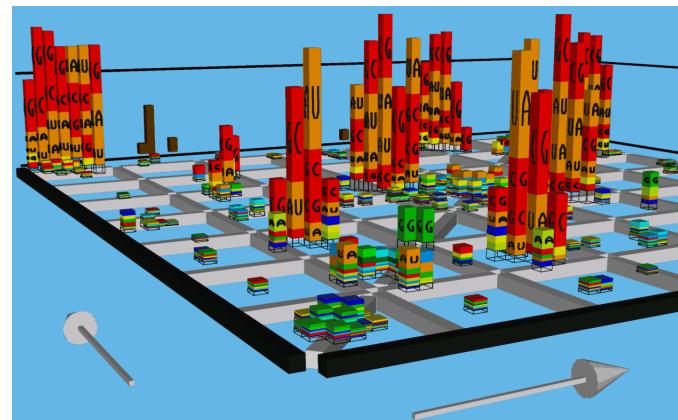
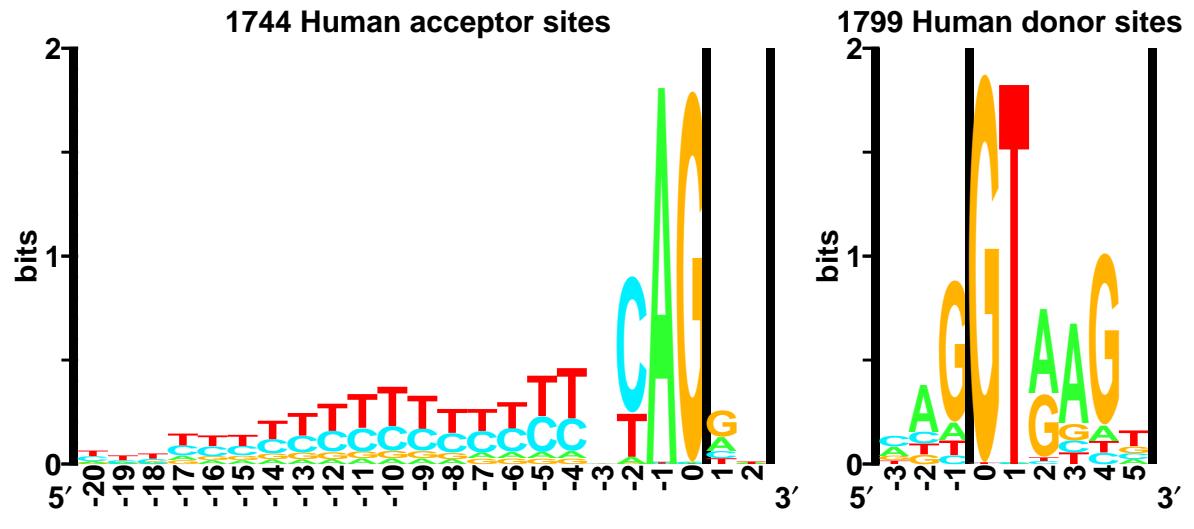
- If $P_y/N_y = 1$ the efficiency is 70%!

Acknowledgements

- Mentors:
 - Larry Gold (graduate school mentor)
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 - Ding Jin (σ^{38})
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Information theory: the mathematics of biology



Version

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